

Communication

Pollination Contribution Differs among Insects Visiting *Cardiocrinum cordatum* Flowers

Riko Komamura ¹, Kohei Koyama ^{1,*} , Takeo Yamauchi ¹, Yasuo Konno ¹ and Lingshuang Gu ²

¹ Department of Agro-environmental Science, Obihiro University of Agriculture and Veterinary Medicine, Inadacho, Obihiro, Hokkaido 080-8555, Japan

² Safety and Emergency Management Research Center, Henan Polytechnic University, Jiaozuo 454000, China

* Correspondence: koyama@obihiro.ac.jp

Abstract: (1) Background: *Cardiocrinum cordatum* (Thunb.) Makino (Liliaceae) is a forest perennial herb distributed in East Asia. Although flower visitors for this plant species have been well reported, their contribution to pollination remains unknown. (2) Methods: We evaluated pollination contribution for visitors of *C. cordatum* flowers in a natural cool temperate forest. We investigated visiting frequency, the number of pollen grains per body surface, fruit set, and the mean number of seeds per fruit produced after a single visit of each visiting species. Combining the results of these experiments, we determined the most important pollinators of this species. (3) Results: For the population investigated in the study, the three most essential pollinators were the bumblebee (*Bombus diversus tersatus*) (Apidae), sweat bee (Halictidae sp.), and marmalade hoverfly (*Episyrrhus balteatus*) (Syrphidae). Additionally, we found that the contribution of a flower-visiting ant species (*Myrmica ruginodis* Nylander (s.l.)) (Formicidae) is small. (4) Conclusions: Pollinator contributions differed among flower visitors. Our results underscore the insufficiency of current information about flower-visiting species to evaluate pollination contribution.



Citation: Komamura, R.; Koyama, K.; Yamauchi, T.; Konno, Y.; Gu, L. Pollination Contribution Differs among Insects Visiting *Cardiocrinum cordatum* Flowers. *Forests* **2021**, *12*, 452. <https://doi.org/10.3390/f12040452>

Academic Editor: David F. Greene

Received: 23 January 2021

Accepted: 7 April 2021

Published: 9 April 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Keywords: pollination; pollinator; plant–animal interaction; mutualism; pollen theft; pollination effectiveness; ant pollination; reproductive biology; floral larceny; forest understory

1. Introduction

The maintenance of genetic diversity is important for the conservation of wild plant species, and successful cross-pollination plays a major role in gene flow to maintain the genetic diversity of plant populations [1–9], especially when a population is fragmented [5–10]. Additionally, pollination is a central issue in studies on plant–animal interactions [11–15], plant reproductive biology [15–20], and on improvement of agricultural production [15,20–28]. For animal-pollinated flowers, pollination effectiveness differs among visitors of different species [3,5,12–15,19,29]. Some visitors obtain pollen or nectar from flowers without contributing as pollinators for some plant species, phenomena known as pollen or nectar theft [30], or nectar robbing for cases where a visitor damages a flower [31]. Therefore, the information on flower-visiting frequency may not be sufficient to evaluate the pollinator contribution of each animal species. Hence, it is necessary to identify which species significantly contribute as pollinators for specific plants.

Cardiocrinum cordatum (Thunb.) Makino (Liliaceae) is a temperate forest perennial herb distributed in East Asia. Although *C. cordatum* is still a common species in forest understories, most of the populations are highly fragmented due to the fragmentation of forests in lowland urban areas [32]. The situation is similar in the urban area of Obihiro city in Hokkaido, in which forests are fragmented into small pieces [33,34]. As the pollination process is critical for the maintenance of genetic diversity of fragmented populations [5,7–10], quantification of pollinator contribution is of utmost importance for future conservation of this species. Ohara et al. [32] suggested that the large nectar quantity and mild floral fragrance of *C. cordatum* attract pollinators; however, they did not investigate

the contribution of flower visitors to pollination. In addition, they suggested that although this species is self-compatible, pollen limitation may also be present for this species. However, although flower visitors have been well documented for this species [32,35,36], their contribution to pollination has not been quantitatively investigated. Here, we examined the pollination contribution of each flower-visiting species using visiting frequency, the number of pollen grains on body surfaces, fruit set, and the mean number of seeds per fruit produced after a single visit in a wild plant population in a natural cool temperate forest in Hokkaido, Japan.

2. Materials and Methods

2.1. Study Species

Cardiocrinum cordatum (Thunb.) Makino (Liliaceae), including var. *glehni*, is a forest perennial herb of East Asia [32,37,38]. Hermaphroditic flowers (Figure 1a–e) have six stamens and one pistil and are self-compatible [7,32,39]. Although their basic structure is similar to lily flowers (e.g., *Lilium auratum* Lindl.), their flower tubes do not open as widely as those of *L. auratum* (Figure 1a–d). Seeds, contained in a capsule, have wings (Figure 1f,g) and are suggested to be dispersed by winds [32]. This species is classified as a monocarpic perennial; the entire aboveground part of a bolting rosette dies after producing seeds, often leaving (usually, one or two) small bulblets at the base of the parent bulbs [6,7,32,40–43]. Therefore, offspring from bulblets compensate for the loss of the parent plant after (apparently) monocarpic sexual reproduction, and occupy the same place as the parental plant [6,43]. Traditionally, the bulb of *C. cordatum* has been used as food [44,45].

2.2. Study Site

The study was performed using a wild population in a natural cool temperate deciduous forest (45°52' N 143°11' E, altitude: 69 m a.s.l.) located in the campus area of the Hokkaido Obihiro Agricultural High School. The mean annual temperature was 7.2 °C and the precipitation was 937 mm at the Japan Meteorological Agency Obihiro Weather Station (6 km from the study site) during 1998–2017 [46]. The understory vegetation comprises a mixture of native species, which includes *C. cordatum*, *Sasa chartacea* (Makino) Makino & Shibata, *Pachysandra terminalis* Siebold & Zucc. [47], and *Phryma esquirolii* H.Lév. The population of *C. cordatum* at the site was comprised of approximately 150–200 individuals and separated from other populations in Obihiro city. The canopy openness before canopy closure was 27.1% [47] and after canopy closure was 4.9% (estimated with hemispherical photographs and the software CanopOn2 [48]).

2.3. Monitoring of Flower Visitors

In lowland forests in Obihiro, flowering of *C. cordatum* occurs during a relatively short period during mid to late July. Each flower lasts approximately one week at the study site. According to our field observation, the pollen was mature and the pistil was receptive simultaneously at the beginning of the first day of the flowering. We marked flowers and determined its age (0 day = the first day of flowering) by monitoring them every day. We investigated the flower visit frequency during midday hours in July in 2020 (11:00 AM–12:40 PM, 9:50 AM–11:00 AM, and 10:50 AM–12:10 PM on 17–19 July, respectively). On each day, an observation was made by one or two observers. Each observer sat in front of one or two individual plants, monitored the flowers on them for 10 min, and moved to next one or two plants. During each 10-min observation, the tentative name of every species that visited the flowers was recorded. Species visits were classified either as (1) those that entered the flower (i.e., those that arrived at the adaxial surface of the corolla) or (2) those that did not enter the flower (i.e., those that arrived only at the external (abaxial) surface of the corolla). Whenever possible, each individual flower-visiting species was captured into either a vial (5, 15, or 20 mL) or a plastic bag (Ziploc, Asahi Kasei, Tokyo, Japan), depending on the body size. Some visitors were recorded as described but were not successfully captured. Separate containers were used for each individual visitor.

Immediately after capturing, each container was stored in a Styrofoam box with ice packs. These containers were moved to a household freezer within the same day and kept until the pollen count assays, as described below. After the pollen counts, one of the authors (T.Y.) identified captured species morphologically.

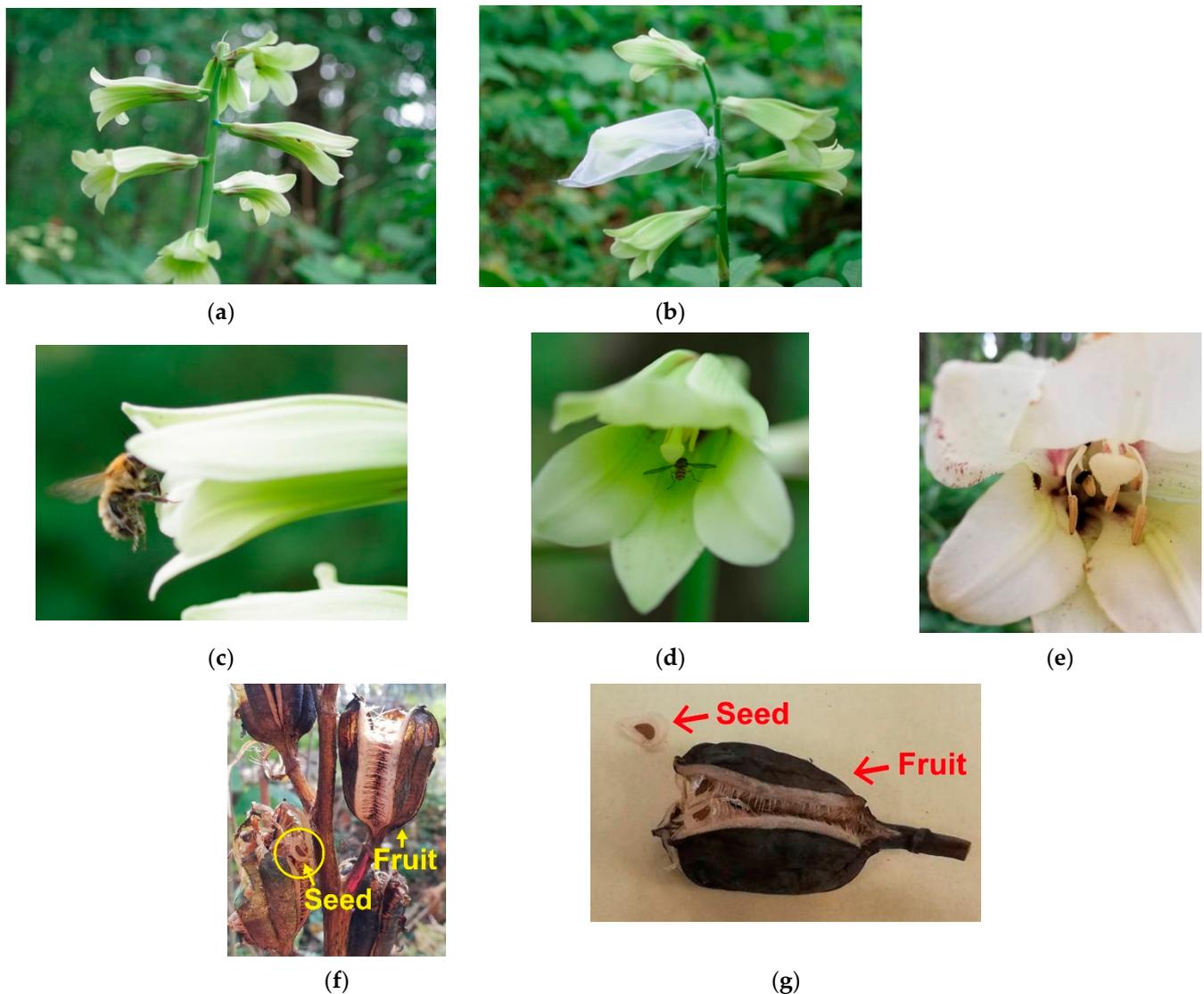


Figure 1. Flowers of *Cardiocrinum cordatum* (Thunb.) Makino. (a) An inflorescence; (b) pollinator exclusion experiment using polyester bags; (c) a bumblebee (*Bombus diversus tersatus*); (d) a marmalade hoverfly (*Episyrphus balteatus*); (e) leaf beetles (*Chrysomelidae* sp.); (f,g) fruits and seeds. Photographs were taken at Hokkaido Obihiro Agricultural High School (a–e) in July and (f,g) in October, in 2020, by (a–d) Kohei Koyama and (e–g) Riko Komamura. The original high-resolution images are available as Supplementary Materials.

2.4. Pollen Counts

We followed the pollen count procedure as described in [29], with some modifications. The hind legs of the bees and bumblebees were removed before counting, because pollen grains on these parts do not contribute to pollination [29,49]. Distilled water (1.0–6.0 mL, depending on the body size) was pipetted into each sample-containing vial. Samples from plastic bags were moved into 15 or 20 mL vials. These vials were shaken manually for 1 min. Distilled water was used instead of an isotonic sucrose solution [29] because the pollen grains of this species do not explode in distilled water. Because the number of *C. cordatum* pollen grains attached to the insect body differed greatly, we adopted different methods

depending on insect species. For the bumblebee (*Bombus diversus tersatus*), sweat bee (*Halictidae* sp.), and marmalade hoverfly (*Episyrphus balteatus*), which contained many pollen grains attached to them, we took subsamples (60 μ L per body) from the shaken water inside each vial with a micropipette (M200; MonotaRO, Amagasaki, Japan) and counted the number of pollen grains under an optical microscope (ECLIPSE E600; Nikon, Tokyo, Japan) (Figure 2). This process was repeated five times, and the total number of pollen grains in the vial was estimated by the volumetric ratio. If Ziploc plastic bags were used to capture the flower visitor, we also washed the inner surface of the bags with 6 mL distilled water. The bags were shaken, and the number of pollen grains inside each bag was calculated by sampling five 60 μ L subsamples, as described above, and the values obtained were added to those on the body surfaces. For remaining species with small pollen counts, we counted all pollen found in the distilled water. After shaking, the distilled water in the vial was put on a small open-top transparent plastic container. A piece of graph paper was attached to the bottom surface of the container to facilitate counting, and the pollen grains were counted under the optical microscope.

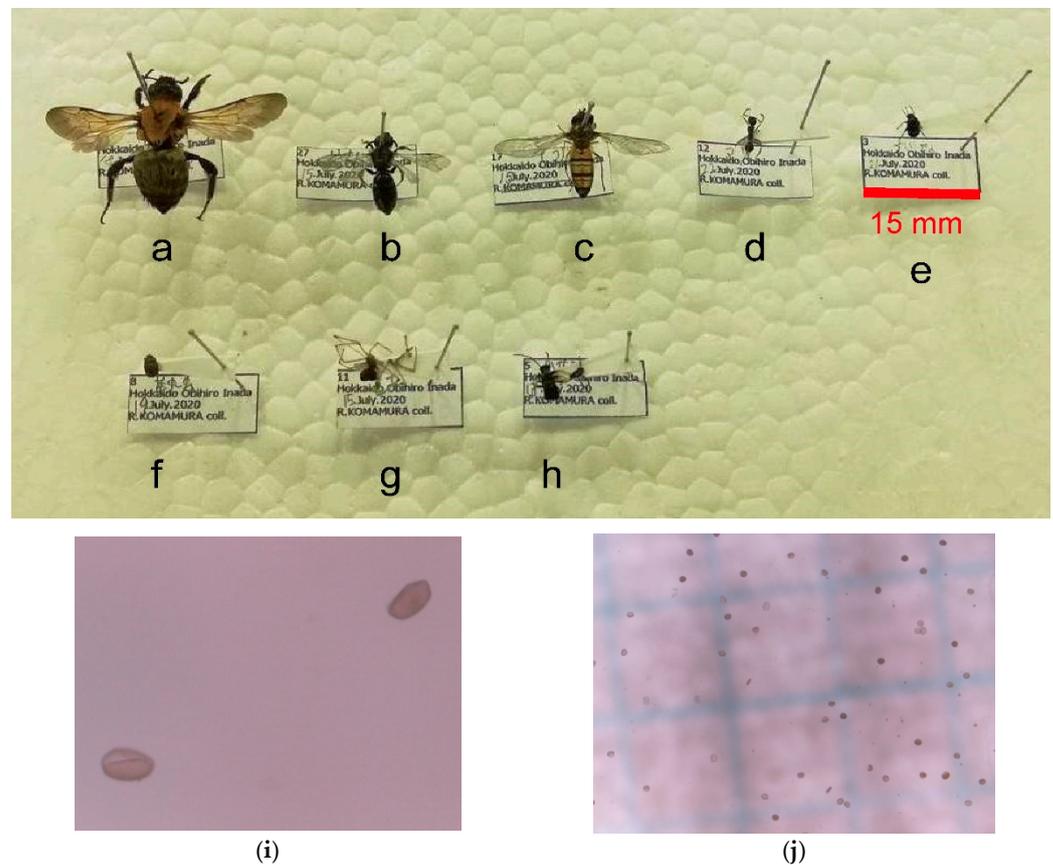


Figure 2. Different species visiting the flowers of *Cardiacrinum cordatum* (Thunb.) Makino. (a) A bumblebee (*Bombus diversus tersatus*), (b) a sweat bee (*Halictidae* sp.), (c) a marmalade hoverfly (*Episyrphus balteatus*), (d) an ant (*Myrmica ruginodis* (s.l.)), (e) a leaf beetle (*Chrysomelidae* sp.), (f) a sap beetle (*Nitidulidae* sp.), (g) a spider (*Araneae* sp.), and (h) an earwig (*Dermaptera* sp.). (i,j) Pollen grains of *C. cordatum* were observed during the counting. The photographs were taken by Riko Komamura in 2020. The original high-resolution images are available as Supplementary Materials.

2.5. Pollinator Exclusion Experiment

Following the procedures as described in [2,7,12,17,18,25,50], we performed a pollinator exclusion experiment by covering the flower with polyester bags (0.6 mm \times 0.9 mm mesh) (Figure 1b). One selected flower was covered for each individual. Before blooming, 85 flower buds were covered with bags. Among them, 15 flowers were assigned as com-

plete pollinator exclusion treatment [18,25], and were covered with the bags during the entire flowering period. The remaining 70 flowers were used to estimate the pollination contribution per visit as described below.

Following the procedures as described in [50], the pollination contribution per visit was investigated during the daytimes of 13–22 July 2020. On each observation day, for each target flower for observation, the bag on one flower was temporarily removed, and an observer sat in front of it until the first visitor entered that flower. In this particular experiment, only those that entered the flower (as described above) were counted as visitors. After the visitor left, the flower was covered again with the same bag, and the observer moved to the next target flower. These visited flowers were kept bagged until the end of the flowering period. Whenever possible, flower-visiting species were captured just after they left the flowers. These captured species were also used for pollen counts as described above.

2.6. Estimating Fruit Set and Seed Number per Fruit

We calculated fruit set (i.e., fruit-to-flower ratio [51,52]) and the seed number per fruit for the 85 bagged fruits used for the pollinator exclusion experiment and, additionally, for different 84 uncovered flowers (i.e., open pollination treatment [15,18,29]) from 15 individuals in the same population. The bags were removed when the flowering period was complete. Then, the fruits were covered again with the same bags before fruit maturation to prevent a loss of seeds by wind dispersal. After maturation, all fruits from the bagged flowers and 22 selected fruits from uncovered flowers were sampled and their seeds were oven-dried at 70 °C for 70 h. For each fruit, the dry mass of 20 selected seeds and the total seed mass were measured with a precision balance and, the total number of seed in each fruit was estimated by the mass ratio. All statistical analyses were performed with statistical software R Version 4.0.4 [53]. All the datasets and the R codes used in this article are available in the Supplementary Materials.

3. Results

3.1. Flower-Visiting Species

The species composition differed among flowers of different ages (Tables 1 and 2). For young flowers, just after the beginning of flowering (flower age, 0–1 day), the most frequent visitors were bumblebees and marmalade hoverflies, followed by ants, sweat bees, and leaf beetles. By contrast, old flowers (age, >2 days) were seldom visited by bumblebees, whereas the same ant species continued to visit the old flowers. Other species (sap beetles, earwigs, and spiders) occasionally arrived at the outer surface of the corolla but they did not enter the flower tube.

3.2. Number of Pollen Grains on the Body Surface

The number of pollen grains on body surfaces differed greatly among the flower visitors (Table 3). Bumblebees and sweat bees carried much more pollen than did the rest of the species, and marmalade hoverflies carried a moderate amount of pollen. For the rest of the species, including the ants, the number of pollen grains on the body surfaces was very small.

3.3. Pollination Contribution per Single Visit

Although this species is known to be self-compatible [7,32,39], in our experiment, only one undeveloped fruit that contained no seed was produced from 15 flowers used for complete pollinator exclusion, and all the other ovules were aborted after flowering. Pollination contribution was confirmed by the observation of fruits produced after single visits for bumblebees, sweat bees, and marmalade hoverflies (Table 4). The fruit set of flowers visited by bumblebees (31.8%) was comparable to those of uncovered flowers (32.1%). Among single-visited flowers, the mean number of seeds per fruit was the highest for the bumblebee-visited flowers. However, the mean number of seeds per fruit produced

after a single visit of bumblebee was lower than those of uncovered flowers. This may be due to pollen limitation, as previously suggested (but not directly tested) [32]. One flower visited by an ant produced a fruit with a small number of seeds, suggesting a possible small contribution of ants as pollinators, but the present limited data do not allow us to confirm the contribution of ants.

Table 1. The observed frequency of flower-visiting species per unit observational time (visits hour⁻¹ flower⁻¹). Flowering age = 0 is defined as the first day of the flowering for each individual flower. In each cell, the fraction X/Y indicates that a total of Y individuals arrived at the flower per hour (including those that only arrived at the outer (abaxial) surface of the corolla), and among them, X entered inside a flower tube (i.e., the adaxial surface of a corolla).

Flower Age (Day)	0	1	2	3	4	Total
Bumblebee (<i>Bombus diversus tersatus</i>)	3.92/3.92	1.64/1.64	0.78/0.78	0	0.23/0.23	6.57/6.57
Sweat bee (<i>Halictidae</i> sp.)	0.23/0.23	0.68/0.82	0.78/0.91	0.77/0.92	0.46/1.15	2.93/4.04
Marmalade hoverfly (<i>Episyrphus balteatus</i>)	1.15/1.62	3.55/4.91	1.70/3.13	1.08/2.15	0.69/0.92	8.16/12.73
Ant (<i>Myrmica ruginodis</i> (s.l.))	1.15/2.77	1.64/4.09	1.04/3.39	2.46/4.77	2.54/4.15	8.83/19.17
Leaf beetle (<i>Chrysomelidae</i> sp.)	0.46/0.46	0.27/0.41	0.65/0.78	0.77/0.77	0.46/0.46	2.62/2.88
Sap beetle (<i>Nitidulidae</i> sp.)	0	0	0	0	0.00/0.23	0.00/0.23
Spider (<i>Araneae</i> sp.) ¹	0	0.00/0.14	0	0.00/0.15	0	0.00/0.29
Earwig (<i>Dermaptera</i> sp.) ¹	0	0	0	0.00/0.15	0.00/0.23	0.00/0.38
Mosquito (<i>Culicidae</i> sp.) ^{1,2}	0.00/0.23	0.00/0.55	0	0.00/0.15	0.00/0.23	0.00/1.16
Unidentified species ¹	0	0.00/0.14	0	0.15/0.15	0.00/0.46	0.15/0.75
Total	6.92/9.23	7.77/12.68	4.96/9.00	5.23/9.23	4.38/8.08	29.27/48.22

¹ Each of these may include multiple species. ² Mosquitoes may have been attracted by the observers and temporarily arrived at the flower.

Table 2. The total number of visits of each species in relation to flower age.

Flower Age (Day)	0	1	2	3	4	Total
Bumblebee (<i>Bombus diversus tersatus</i>)	17/17	12/12	6/6	0/0	1/1	36/36
Sweat bee (<i>Halictidae</i> sp.)	1/1	5/6	6/7	5/6	2/5	19/25
Marmalade hoverfly (<i>Episyrphus balteatus</i>)	5/7	26/36	13/24	7/14	3/4	54/85
Ant (<i>Myrmica ruginodis</i> (s.l.))	5/12	12/30	8/26	16/31	11/18	52/117
Leaf beetle (<i>Chrysomelidae</i> sp.)	2/2	2/3	5/6	5/5	2/2	16/18
Sap beetle (<i>Nitidulidae</i> sp.)	0/0	0/0	0/0	0/0	0/1	0/1
Spider (<i>Araneae</i> sp.) ¹	0/0	0/1	0/0	0/1	0/0	0/2
Earwig (<i>Dermaptera</i> sp.) ¹	0/0	0/0	0/0	0/1	0/1	0/2
Mosquito (<i>Culicidae</i> sp.) ^{1,2}	0/1	0/4	0/0	0/1	0/1	0/7
Unidentified species ¹	0/0	0/1	0/0	1/1	0/2	1/4
Total	30/40	57/93	38/69	34/60	19/35	178/297

¹ Each of these may include multiple species. ² Mosquitoes may have been attracted by the observers and temporarily arrived at the flower.

Table 3. The total number of pollen grains attached on the surface of the flower-visiting animals.

Species	Total No. of Individual Visitors Captured	No. of Pollens on Each Body Surface	
		Mean	SD
Bumblebee (<i>Bombus diversus tersatus</i>)	15	54,328	44,745.5
Sweat bee (<i>Halictidae</i> sp.)	10	25,913	25,902.5
Marmalade hoverfly (<i>Episyrphus balteatus</i>)	15	1468.5	1813.8
Ant (<i>Myrmica ruginodis</i> (s.l.))	4	14	15.7
Leaf beetle (<i>Chrysomelidae</i> sp.)	4	48.8	14.9
Sap beetle (<i>Nitidulidae</i> sp.)	1	0	-
Spider (<i>Araneae</i> sp.)	1	166	-
Earwig (<i>Dermaptera</i> sp.)	1	10	-

Table 4. Pollination contribution of each species, estimated as the total number of seeds in each fruit produced after a single visit of each individual insect, with the prior exclusion and subsequent exclusion of the other visitors by covering the flowers with bags.

Visitor	No. of Flowers Investigated (FL)	No. of Fruits Produced (FR)	Fruit Set (FR/FL) (%)	No. of Fruits Investigated for Seed Counts	Mean No. of Seeds per Fruit	
					Mean	SD
Uncovered flowers ¹	84	27	32.1	22	516.3	93.2
Bumblebee (<i>Bombus diversus tersatus</i>)	22	7	31.8	7	364.6	193.4
Sweat bee (<i>Halictidae</i> sp.)	13	3	23.1	3	215.2	246.3
Marmalade hoverfly (<i>Episyrphus balteatus</i>)	22	5	22.7	5	194.8	259.7
Ant (<i>Myrmica ruginodis</i> (s.l.))	1	1	100	1	78.6	-
Complete pollinator exclusion	15	0	0	-	-	-

¹ The total number of visits varied among uncovered flowers.

4. Discussion

Our results suggest that bumblebees (*Bombus diversus tersatus* [syn. *Bombus tersatus*]), which feed on both nectar [54] and pollen [55], are the most important pollinators for this species for the population investigated in the present study. Generally, insects that visit young flowers with abundant pollen are important pollinators [50]. Bumblebees visited young flowers (flower age, 0–1 day) (Table 1). Accordingly, bumblebees carried more pollen than other pollinators did (Table 3). Pollination efficiency is also determined by match and mismatch between flower morphology (including the size of flower parts) and animal morphology [30,31]. All of the observed bumblebees entered into the inner (adaxial) surface of the corolla (Table 1), and their large body length (ca. 1–2 cm), together with the tube-shaped morphology of the *C. cordatum* flower (Figure 1), forced them into contact with the stigma when they entered the flower to extract the nectar. These observations are consistent with the results that they are important pollinators, as confirmed with the pollinator exclusion experiment (Table 4). Several species of bumblebees (genus *Bombus*) are generally well known to be effective pollinators [29,56], and are recognized as flower visitors of *C. cordatum* [32,35]. However, in previous studies on *C. cordatum*, the contribution of bumblebees to pollination remains elusive. The results of the present study are consistent with the aforementioned previous findings and provide further quantitative evidence confirming that *B. diversus tersatus* is an important pollinator for *C. cordatum*. Additionally, *B. diversus* has wide foraging range [33]. This indicates that the fragmented populations of *C. cordatum* in the lowland urban area might be genetically connected to the nearby populations with the contribution of bumblebees as pollinators.

Ohara et al. [32] and Nishizawa and Ohara [6] discussed that the large nectar quantity and mild flower fragrance of *C. cordatum* attract pollinators. Nevertheless, not all flower visitors may have access to nectar because the length of the proboscis is known to limit the ability of flower-visiting insects to access nectar [5,57,58]. Among the flower visitors observed in the present study, bumblebees (*B. diversus*) have the longest proboscis (approximate size 10.5–13.5 mm [59]), which is longer than that of sweat bees (*Halictidae* sp., typically, <2 mm [60]) or marmalade hoverflies (*Episyrphus balteatus*, 2.1–2.6 mm [57]). The flower tube of *C. cordatum* (Liliaceae) does not open as widely as that of *Lilium auratum* (Figure 1a,c,d). Therefore, a flower visitor with a short proboscis may not have access to the nectar of *C. cordatum*. The long, tube-shaped flower morphology of *C. cordatum* with narrow opening corolla may serve as a mechanism that limits visitor access to its nectar, thus restricting the nectar access to bumblebees that are the most efficient pollinators.

van Rijn et al. [61] reported that marmalade hoverflies feed on both pollen and nectar. Consistent with their results, we observed that they gathered or fed on pollen dropped from the anthers to the surface of the petals (Figure 1d). These behaviors may explain the observed high pollen amount on their body surface (Table 3). We also observed that sweat bees and marmalade hoverflies were occasionally attached to the stigma, consistent with their contribution as pollinators (Table 4). Ants are generally considered as nectar thieves [62], and their contribution to plant reproduction is thought to be low in many

cases [62], although they indeed in some cases contribute as pollinators [63,64]. Although the ants observed in the present study (*Myrmica ruginodis* (s.l.)) were frequent visitors (Table 1), the number of pollen grains on the body surfaces of the ant species was small (Table 3). Additionally, we observed that the ants, because of their small body size (ca. 5 mm), rarely came into contact with the stigma. These results indicate that the ant species may not play a major role as pollinators. Other species, including sap beetles (Nitidulidae sp.), earwigs (Dermaptera sp.), and spiders (Araneae sp.) occasionally arrived at the outer surface of the corolla, but they did not enter inside the flower tube (Table 1). The number of pollen grains on their body surfaces was small (Table 3), indicating that they are not pollinators, at least in our present results.

Our study has several limitations. First, we investigated only a single population for a single flowering period. In general, reproductive patterns of plants differ among populations of the same species from different environments [6,7,36,39]. Second, the existence of nocturnal floral visitors was not investigated in the present study. Although we are unaware of any reports on nocturnal visitors for this plant species, further studies, including nighttime investigations, are needed in future studies. Third, we did not investigate the genetic composition or quality component of pollens. Self-pollination causes inbreeding depression, and hence outcross pollen is generally considered as of higher quality than self-pollen [1,3,8,25,62,63,65]. Matsuki et al. [3] reported that certain beetle species carry genetically more diverse outcross pollen than bumblebees. Rostás et al. [62,63] further argued that fruit or seed set may not give a sufficient estimate of contribution as pollinators and emphasized the importance of investigation of seedling viability and offspring vigor, which are determined by the genetic composition of pollen. Given these important limitations, further studies are needed to reconfirm our findings before generalization.

Supplementary Materials: The following are available online at: <https://www.mdpi.com/article/10.3390/f12040452/s1>.

Author Contributions: Conceptualization, R.K. and K.K.; methodology, R.K., K.K. and Y.K.; investigation, R.K. and K.K.; identification of insect species, T.Y.; writing—original draft preparation, R.K. and K.K.; writing—review and editing, R.K., K.K., Y.K. and L.G.; Supervision, K.K. All authors have read and agreed to the published version of the manuscript.

Funding: This work was funded by the Japan Society for the Promotion of Science (KAKENHI Grant Number 18K06406).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: All the datasets and the R codes used in this article are available in the Supplementary Materials.

Acknowledgments: We thank Norikuni Kumano for useful advices on pollen counts. We also thank staff members of Hokkaido Obihiro Agricultural High School for permitting us to perform the fieldwork at the study site.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Collevatti, R.G.; Estolano, R.; Garcia, S.F.; Hay, J.D. Seed abortion in the bat pollinated Neotropical tree species, *Caryocar brasiliense* (Caryocaraceae). *Botany* **2009**, *87*, 1110–1115. [[CrossRef](#)]
2. Kunitake, Y.K.; Hasegawa, M.; Miyashita, T.; Higuchi, H. Role of a seasonally specialist bird *Zosterops japonica* on pollen transfer and reproductive success of *Camellia japonica* in a temperate area. *Plant Spec. Biol.* **2004**, *19*, 197–201. [[CrossRef](#)]
3. Matsuki, Y.; Tateno, R.; Shibata, M.; Isagi, Y. Pollination efficiencies of flower-visiting insects as determined by direct genetic analysis of pollen origin. *Am. J. Bot.* **2008**, *95*, 925–930. [[CrossRef](#)]
4. Nakamura, M.; Nanami, S.; Okuno, S.; Hirota, S.K.; Matsuo, A.; Suyama, Y.; Tokumoto, H.; Yoshihara, S.; Itoh, A. Genetic diversity and structure of apomictic and sexually reproducing *Lindera* species (Lauraceae) in Japan. *Forests* **2021**, *12*, 227. [[CrossRef](#)]
5. Yoshida, Y.; Nikkeshi, A.; Chishiki, A. Identification of effective pollinators of *Primula sieboldii* E. Morren in a wild habitat in Hiroshima, Japan. *Plant Spec. Biol.* **2020**. [[CrossRef](#)]

6. Nishizawa, M.; Ohara, M. The role of sexual and vegetative reproduction in the population maintenance of a monocarpic perennial herb, *Cardiocrinum cordatum* var. *glehnii*. *Plant Spec. Biol.* **2018**, *33*, 289–304. [[CrossRef](#)]
7. Narumi, T.; Ohara, M. Variation in reproductive modes and population genetic structures of a monocarpic perennial herb, *Cardiocrinum cordatum*, in relation to habitat fragmentation. *Plant Spec. Biol.* **2018**, *33*, 248–258. [[CrossRef](#)]
8. Al-Qathanin, R.N.; Alharbi, S.A. Spatial structure and genetic variation of a mangrove species (*Avicennia marina* (Forssk.) Vierh) in the Farasan Archipelago. *Forests* **2020**, *11*, 1287. [[CrossRef](#)]
9. Lu, J.-T.; Qiu, Y.-H.; Lu, J.-B. Effects of landscape fragmentation on genetic diversity of male-biased dioecious plant *Pistacia chinensis* bunge populations. *Forests* **2019**, *10*, 792. [[CrossRef](#)]
10. Collevatti, R.G.; Grattapaglia, D.; Hay, J.D. High resolution microsatellite based analysis of the mating system allows the detection of significant biparental inbreeding in *Caryocar brasiliense*, an endangered tropical tree species. *Heredity* **2001**, *86*, 60–67. [[CrossRef](#)]
11. Geerts, S.; Coetzee, A.; Rebelo, A.G.; Pauw, A. Pollination structures plant and nectar-feeding bird communities in Cape fynbos, South Africa: Implications for the conservation of plant–bird mutualisms. *Ecol. Res.* **2020**, *35*, 838–856. [[CrossRef](#)]
12. Herrera, C.M. Components of pollinator “quality”: Comparative analysis of a diverse insect assemblage. *Oikos* **1987**, *50*, 79–90. [[CrossRef](#)]
13. Herrera, C.M. Pollinator abundance, morphology, and flower visitation rate: Analysis of the “quantity” component in a plant–pollinator system. *Oecologia* **1989**, *80*, 241–248. [[CrossRef](#)] [[PubMed](#)]
14. Schupp, E.W.; Jordano, P.; Gómez, J.M. A general framework for effectiveness concepts in mutualisms. *Ecol. Lett.* **2017**, *20*, 577–590. [[CrossRef](#)] [[PubMed](#)]
15. Conceição, S.I.R.; Fernandes, J.; Borges da Silva, E.; Caperta, A.D. Reproductive output and insect behavior in hybrids and apomicts from *Limonium ovalifolium* and *L. binervosum* complexes (Plumbaginaceae) in an open cross-pollination experiment. *Plants* **2021**, *10*, 169. [[CrossRef](#)]
16. Tokumoto, Y.; Matsushita, M.; Kishimoto-Yamada, K.; Nikkeshi, A.; Isogimi, T.; Nakagawa, M. Floral visitors and reproductive success in two sequentially flowering *Lindera shrubs* (Lauraceae) of central Japan. *J. For. Res.* **2019**, *24*, 42–51. [[CrossRef](#)]
17. Yamasaki, E.; Sakai, S. Wind and insect pollination (ambophily) of *Mallotus* spp. (Euphorbiaceae) in tropical and temperate forests. *Aust. J. Bot.* **2013**, *61*, 60–66. [[CrossRef](#)]
18. Giblin, D.E. Variation in floral longevity between populations of *Campanula rotundifolia* (Campanulaceae) in response to fitness accrual rate manipulation. *Am. J. Bot.* **2005**, *92*, 1714–1722. [[CrossRef](#)]
19. Soley, N.M.; Sipes, S.D. Reproductive biology and pollinators of the invasive shrub Autumn olive (*Elaeagnus umbellata* Thunberg). *Plant Spec. Biol.* **2020**. [[CrossRef](#)]
20. Hou, S.; Zhao, T.; Yang, D.; Li, Q.; Liang, L.; Wang, G.; Ma, Q. Selection and validation of reference genes for quantitative RT-PCR analysis in *Corylus heterophylla* Fisch. × *Corylus avellana* L. *Plants* **2021**, *10*, 159. [[CrossRef](#)]
21. Bentrup, G.; Hopwood, J.; Adamson, N.L.; Vaughan, M. Temperate agroforestry systems and insect pollinators: A review. *Forests* **2019**, *10*, 981. [[CrossRef](#)]
22. Tran, X.T.; Parks, S.E.; Nguyen, M.H.; Roach, P.D. Reduced pollination efficiency compromises some physicochemical qualities in gac (*Momordica cochinchinensis* Spreng.) fruit. *Agronomy* **2021**, *11*, 190. [[CrossRef](#)]
23. Fernández, F.J.; Garay, J.; Móri, T.F.; Csiszár, V.; Varga, Z.; López, I.; Gámez, M.; Cabello, T. Theoretical foundation of the control of pollination by hoverflies in a greenhouse. *Agronomy* **2021**, *11*, 167. [[CrossRef](#)]
24. Aizen, M.A.; Aguiar, S.; Biesmeijer, J.C.; Garibaldi, L.A.; Inouye, D.W.; Jung, C.; Martins, D.J.; Medel, R.; Morales, C.L.; Ngo, H.; et al. Global agricultural productivity is threatened by increasing pollinator dependence without a parallel increase in crop diversification. *Glob. Chang. Biol.* **2019**, *25*, 3516–3527. [[CrossRef](#)] [[PubMed](#)]
25. Cusser, S.; Neff, J.L.; Jha, S. Natural land cover drives pollinator abundance and richness, leading to reductions in pollen limitation in cotton agroecosystems. *Agr. Ecosyst. Environ.* **2016**, *226*, 33–42. [[CrossRef](#)]
26. McGrady, C.M.; Troyer, R.; Fleischer, S.J. Wild bee visitation rates exceed pollination thresholds in commercial *Cucurbita* agroecosystems. *J. Econ. Entomol.* **2020**, *113*, 562–574. [[CrossRef](#)]
27. Pfister, S.C.; Eckert, P.W.; Schirmel, J.; Cresswell, J.E.; Entling, M.H. Sensitivity of commercial pumpkin yield to potential decline among different groups of pollinating bees. *R. Soc. Open Sci.* **2017**, *4*, 170102. [[CrossRef](#)] [[PubMed](#)]
28. Senapathi, D.; Fründ, J.; Albrecht, M.; Garratt, M.P.D.; Kleijn, D.; Pickles, B.J.; Potts, S.G.; An, J.; Andersson, G.K.S.; Bänisch, S.; et al. Wild insect diversity increases inter-annual stability in global crop pollinator communities. *Proc. R. Soc. B Biol. Sci.* **2021**, *288*, 20210212. [[CrossRef](#)]
29. Nikkeshi, A.; Inoue, H.; Arai, T.; Kishi, S.; Kamo, T. The bumblebee *Bombus ardens ardens* (Hymenoptera: Apidae) is the most important pollinator of Oriental persimmon, *Diospyros kaki* (Ericales: Ebenaceae), in Hiroshima, Japan. *Appl. Entomol. Zool.* **2019**, *54*, 409–419. [[CrossRef](#)]
30. Hargreaves, A.L.; Harder, L.D.; Johnson, S.D. Consumptive emasculation: The ecological and evolutionary consequences of pollen theft. *Biol. Rev.* **2009**, *84*, 259–276. [[CrossRef](#)] [[PubMed](#)]
31. Irwin, R.E.; Bronstein, J.L.; Manson, J.S.; Richardson, L. Nectar robbing: Ecological and evolutionary perspectives. *Annu. Rev. Ecol. Evol. Syst.* **2010**, *41*, 271–292. [[CrossRef](#)]
32. Ohara, M.; Narumi, T.; Yoshizane, T.; Okayasu, T.; Masuda, J.; Kawano, S. 7: *Cardiocrinum cordatum* (Thunb.) Makino (Liliaceae). *Plant Spec. Biol.* **2006**, *21*, 201–207. [[CrossRef](#)]

33. Nagamitsu, T.; Tsukuba, S.-A.; Ushirokita, F.; Konno, Y. Foraging habitats and floral resource use by colonies of long- and short-tongued bumble bee species in an agricultural landscape with kabocha squash fields. *Appl. Entomol. Zool.* **2012**, *47*, 181–190. [[CrossRef](#)]
34. Konno, Y. Present status of remnant forests in Obihiro, eastern Hokkaido, Japan. In *Global Perspective in Forest Conservation and Sustainable Agriculture*; Obihiro Asia and the Pacific Seminar on Education for Rural Development (OASERD): Obihiro, Japan, 2002; pp. 39–46.
35. Matsumura, C.; Yokoyama, J.; Washitani, I. Invasion status and potential ecological impacts of an invasive alien bumblebee, *Bombus terrestris* L. (Hymenoptera: Apidae) naturalized in Southern Hokkaido, Japan. *Glob. Environ. Res.* **2004**, *8*, 51–66.
36. Cao, G.-X.; Kudo, G. Size-dependent sex allocation in a monocarpic perennial herb, *Cardiocrinum cordatum* (Liliaceae). *Plant Ecol.* **2008**, *194*, 99–107. [[CrossRef](#)]
37. Lu, R.-S.; Chen, Y.; Tamaki, I.; Sakaguchi, S.; Ding, Y.-Q.; Takahashi, D.; Li, P.; Isagi, Y.; Chen, J.; Qiu, Y.-X. Pre-quaternary diversification and glacial demographic expansions of *Cardiocrinum* (Liliaceae) in temperate forest biomes of Sino-Japanese Floristic Region. *Mol. Phylogenet. Evol.* **2020**, *143*, 106693. [[CrossRef](#)]
38. Lu, R.-S.; Li, P.; Qiu, Y.-X. The complete chloroplast genomes of three *Cardiocrinum* (Liliaceae) species: Comparative genomic and phylogenetic analyses. *Front. Plant Sci.* **2017**, *7*, 2054. [[CrossRef](#)]
39. Cao, G.X.; Worley, A.C. Life history trade-offs and evidence for hierarchical resource allocation in two monocarpic perennials. *Plant Biol.* **2013**, *15*, 158–165. [[CrossRef](#)]
40. Koyama, K.; Hidaka, Y.; Ushio, M. Dynamic scaling in the growth of a non-branching plant, *Cardiocrinum cordatum*. *PLoS ONE* **2012**, *7*, e45317. [[CrossRef](#)]
41. Araki, K.; Shimatani, K.; Nishizawa, M.; Yoshizane, T.; Ohara, M. Growth and survival patterns of *Cardiocrinum cordatum* var. *glehnii* (Liliaceae) based on a 13-year monitoring study: Life history characteristics of a monocarpic perennial herb. *Botany* **2010**, *88*, 745–752. [[CrossRef](#)]
42. Kondo, T.; Sato, C.; Baskin, J.M.; Baskin, C.C. Post-dispersal embryo development, germination phenology, and seed dormancy in *Cardiocrinum cordatum* var. *glehnii* (Liliaceae s. str.), a perennial herb of the broadleaved deciduous forest in Japan. *Am. J. Bot.* **2006**, *93*, 849–859. [[CrossRef](#)] [[PubMed](#)]
43. Hayafune, T.; Utech, F.H.; Ohara, M. Inter-populational variation, but no-annual variation within populations, in terms of reproductive size and genetic structure in a monocarpic perennial herb, *Cardiocrinum cordatum* var. *glehnii*. *Plant Spec. Biol.* **2019**, *34*, 27–30. [[CrossRef](#)]
44. Hori, K.; Watanabe, T.; Devkota, H.P. Phenolic acid derivatives, flavonoids and other bioactive compounds from the leaves of *Cardiocrinum cordatum* (Thunb.) Makino (Liliaceae). *Plants* **2021**, *10*, 320. [[CrossRef](#)] [[PubMed](#)]
45. Nishiumi, S.; Hosokawa, K.; Anetai, M.; Shibata, T.; Mukai, R.; Yoshida, K.-i.; Ashida, H. Antagonistic effect of the ainu-selected traditional beneficial plants on the transformation of an aryl hydrocarbon receptor. *J. Food Sci.* **2012**, *77*, C420–C429. [[CrossRef](#)]
46. Japan Meteorological Agency. Available online: <http://www.jma.go.jp> (accessed on 14 September 2020).
47. Iwabe, R.; Koyama, K.; Komamura, R. Shade avoidance and light foraging of a clonal woody species, *Pachysandra terminalis*. *Plants*. under review.
48. Takenaka, A. CanopOn 2 ver. 2.03c. 2009. Available online: <http://takenaka-akio.org/etc/canopon2/index.html> (accessed on 26 September 2020).
49. Parker, A.J.; Tran, J.L.; Ison, J.L.; Bai, J.D.K.; Weis, A.E.; Thomson, J.D. Pollen packing affects the function of pollen on corbiculate bees but not non-corbiculate bees. *Arthropod-Plant Int.* **2015**, *9*, 197–203. [[CrossRef](#)]
50. Sakai, S. *Handbook of Methods in Ecological Research 2: Field Methods in Pollination Ecology*; Kyritsu Publishing: Tokyo, Japan, 2015. (In Japanese)
51. Sutherland, S.D. Why hermaphroditic plants produce many more flowers than fruits: Experimental tests with *Agave mckelveyana*. *Evolution* **1987**, *41*, 750–759. [[CrossRef](#)]
52. Koyama, K.; Tashiro, M. No effect of selective maturation on fruit traits for a bird-dispersed species, *Sambucus racemosa*. *Plants* **2021**, *10*, 376. [[CrossRef](#)]
53. R Core Team. *R: A Language and Environment for Statistical Computing*; R Foundation for Statistical Computing: Vienna, Austria, 2021.
54. Kobayashi, S.; Inoue, K.; Kato, M. Evidence of pollen transfer efficiency as the natural selection factor favoring a large corolla of *Campanula punctata* pollinated by *Bombus diversus*. *Oecologia* **1997**, *111*, 535–542. [[CrossRef](#)]
55. Katayama, E. Studies on the development of the broods of *Bombus diversus* Smith (Hymenoptera, Apidae): II. Brood development and feeding habits. *Kontyu* **1966**, *34*, 8–17.
56. Darvill, B.; Knight, M.E.; Goulson, D. Use of genetic markers to quantify bumblebee foraging range and nest density. *Oikos* **2004**, *107*, 471–478. [[CrossRef](#)]
57. van Rijn, P.C.J.; Wäckers, F.L. Nectar accessibility determines fitness, flower choice and abundance of hoverflies that provide natural pest control. *J. Appl. Ecol.* **2016**, *53*, 925–933. [[CrossRef](#)]
58. Inouye, D.W. The effect of proboscis and corolla tube lengths on patterns and rates of flower visitation by bumblebees. *Oecologia* **1980**, *45*, 197–201. [[CrossRef](#)]
59. Inoue, M.N.; Yokoyama, J. Competition for flower resources and nest sites between *Bombus terrestris* (L.) and Japanese native bumblebees. *Appl. Entomol. Zool.* **2010**, *45*, 29–35. [[CrossRef](#)]

60. Inoue, T.; Kato, M. *Hana ni Hikiyose Rareru Dobutsu—Hana to Doubutsu no Kyoshinka*; Heibonsya: Tokyo, Japan, 1993. (In Japanese)
61. van Rijn, P.C.J.; Kooijman, J.; Wäckers, F.L. The contribution of floral resources and honeydew to the performance of predatory hoverflies (Diptera: Syrphidae). *Biol. Control* **2013**, *67*, 32–38. [[CrossRef](#)]
62. Rostás, M.; Bollmann, F.; Saville, D.; Riedel, M. Ants contribute to pollination but not to reproduction in a rare calcareous grassland forb. *PeerJ* **2018**, *6*, e4369. [[CrossRef](#)]
63. Rostás, M.; Tautz, J. Ants as pollinators of plants and the role of floral scents. In *All Flesh Is Grass: Plant-Animal Interrelationships*; Dubinsky, Z., Seckbach, J., Eds.; Springer: Dordrecht, The Netherlands, 2011; pp. 149–161. [[CrossRef](#)]
64. de Vega, C.; Herrera, C.M.; Dötterl, S. Floral volatiles play a key role in specialized ant pollination. *Perspect. Plant Ecol. Evol. Syst.* **2014**, *16*, 32–42. [[CrossRef](#)]
65. Collevatti, R.G.; Amara, M.E.C.; Lopes, F.S. Role of pollinators in seed set and a test of pollen limitation hypothesis in the tropical weed *Triumfetta semitriloba* (Tiliaceae). *Rev. Biol. Trop.* **1997**, *45*, 1401–1407.