

**Title**

Genetic characterization of *Neospora caninum* from aborted bovine fetuses in Hokkaido, Japan

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**List of abbreviations:** JPN, Japan; MS, microsatellite; MLGs, multilocus genotypes; BIC, Bayesian information criterion; DAPC, discriminant analysis of principal components; PCs, posterior clusters

## Abstract

*Neospora caninum* is one of the main causes of bovine abortions worldwide, including Japan. Nothing is known about the *N. caninum* population substructures in Japan, and only one isolate from a pregnant sheep has been studied to date. This study describes, for the first time, the genetic characterization of isolates of *N. caninum* implicated in cattle abortions in Japan. Brains from five aborted fetuses were successfully genotyped based on multilocus microsatellite markers. Assigned genotypes showed high frequencies of mixed alleles in the sequenced markers MS7 and MS10, raising concerns about the subpopulation structures of *N. caninum* infecting animals in Japan. Clustering analysis of the genotypes, together with those from a previous dataset, showed that five of the six genotypes were distinct from other clusters. Meanwhile, the remaining genotype, together with the sheep isolate from Japan, was grouped with those from Mexico and Spain. These preliminary data may indicate a complex transmission pattern of *N. caninum* in Japan via clonal spreading by vertical and horizontal transmission and geographically related population substructuring.

**Keywords:** *Neospora caninum*, Abortion, Cattle, Microsatellite, Japan

## 1. Introduction

Neosporosis is considered a serious disease in cattle, causing abortions and neonatal mortality (Dubey et al., 2007; Dubey and Schares, 2011) and subsequent economic losses (Reichel et al., 2013). Evidence suggests that *Neospora caninum* isolates differ in their virulence, which may explain variations in disease outcomes and epidemiology (Al-Qassab et al., 2010; Dubey and Schares 2011; Goodswen et al., 2013). The clonal model of *N. caninum* expansion has been recently suggested, with a highly inbred genome (Khan et al., 2019). Nevertheless, multilocus microsatellite genotyping revealed extensive genetic diversity among *N. caninum* isolates, indicating predominant clonal propagation in cattle and geographically related population substructures (Regidor-Cerrillo et al., 2020).

*Neospora caninum* has been implicated in bovine abortions in Japan, with nationwide distribution and high economic losses (Koiwai et al., 2005a; Koiwai et al., 2006; Yamane et al., 2000). However, genetic analysis has not been conducted on the isolates from cattle abortions in Japan, and to date, only one isolate has been analyzed from a pregnant sheep without clinical disease (Koyama et al., 2001). This study, for the first time, aimed to determine the genetic relationship between *N. caninum* isolates causing cattle abortions in Japan and other worldwide isolates.

## 2. Material and methods

### 2.1. Ethics statement

All animal experiments strictly followed the recommendations of the Guide for the Care and Use of Laboratory Animals of the Ministry of Education, Culture, Sports, Science and Technology, Japan. The study protocol was approved by the Committee on the Ethics of Animal

1 Experiments at the Obihiro University of Agriculture and Veterinary Medicine (permission  
2 numbers: 18-15, 19-3, 20-3).  
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## 7 2.2. *Neospora caninum* infected tissues 8 9

10 Fetal tissues from aborted dams were collected by the Tokachi Livestock Hygiene  
11 Service Center and tested for *N. caninum* antigens using immunohistochemical assays.  
12 Additionally, their mother's sera were tested for seropositivity to *N. caninum* by  
13 immunofluorescence antibody test (VMRD, Pullman, WA, USA). Four samples (JPN-10-H10,  
14 JPN-10-H11, JPN-10-H12, JPN-10-H13) were obtained from aborted bovine fetuses upon  
15 outbreak of an abortion epidemic in a dairy cattle farm in Shihoro, Hokkaido, Japan (2010).  
16 Additionally, one sample (JPN-10-H218) was obtained from an aborted fetus in a farm located  
17 in the same area (2018).  
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## 31 2.3. Genetic characterization of *N. caninum* 32 33

34 Brain tissues (0.5 grams) were lysed by incubation with 10 volumes (5 ml) of extraction  
35 buffer (0.1 M Tris-HCl pH 9.0, 1% SDS, 0.1 M NaCl, 1 mM EDTA) and 1 mg/ml of proteinase  
36 K at 50°C. DNA was extracted from the tissue's lysate using phenol-chloroform extraction and  
37 precipitated with ethanol. Confirmation of *N. caninum* DNA in the infected brain tissues was  
38 achieved by amplifying a fragment of the specific sequence Nc5 using nested PCR, with the  
39 primers described by Yamage et al.,1996, and PCR cycling was performed as previously  
40 described (Du et al., 2015).  
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51 Genetic characterization of *N. caninum* from brain samples DNA (200 ng) was  
52 performed by microsatellite analysis employing 12 MS markers: MS1A, MS1B, MS2, MS4,  
53 MS5, MS6A, MS6B, MS7, MS8, MS10, MS12, and MS21. All PCR conditions, including the  
54 external and internal primers as well as the 6-FAM-labeled reverse primers, were adopted as  
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previously described (Pedraza-Díaz et al., 2009; Regidor-Cerrillo et al., 2013). Negative controls were used, including Vero-cell lysates and milliQ water, and the DNA of the Nc1 strain of *N. caninum* tachyzoites was used as the positive control in all PCR reactions. The sizes of the 6-FAM-labeled PCR products for all of the MSs were determined using an 8-capillary 3500 DNA analyzer (Applied Biosystems, Foster City, CA, USA) with Gene Scan-500 (LIZ) Size Standards (Applied Biosystems) and the GeneMapper™ Software 5 (Applied Biosystems).

Amplicons from the non-labeled MS7 and MS10 primers were separated by electrophoresis on 1% agarose gels. The bands of interest were excised from the gel and purified with NucleoSpin® Gel and a PCR Clean-up Kit (Macherey-Nagel, Diiren, Germany). The quantity and purity of the purified amplicons were measured using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA) and cloned using pCR™4-TOPO® TA Cloning Kits for Sequencing (Thermo Fisher Scientific, St. Louis, MO, USA). The plasmid construct was extracted using the QIA prep®Spin Miniprep Kit (Qiagen), and four positive clones for each isolate were bidirectionally sequenced using the M13 forward and reverse primers and Bigdye Terminator Cycle Sequencing Kit V 3.1 (Applied Biosystems) on a 3500 Genetic Analyzer (Applied Biosystems).

The representative MS7 and MS10 sequences obtained in this study were deposited in the GenBank database under the accession numbers MT782054 to MT782062. The multilocus genotype (MLG) of each isolate was determined based on the combined results of the number of repeats deduced from automated allele sizing and cloning-sequencing in the nine MS markers following the procedure described by Regidor-Cerrillo et al. (2013).

## 2.4. Data analysis

Nine loci were used for clustering analysis (MS4, MS5, MS6A, MS6B, MS7, MS8, MS10, MS12, and MS21). A previous *N. caninum* MLGs dataset (Medina-Esparza et al., 2016),

1 together with the new sheep genotypes (García-Sánchez et al., 2020), was used to identify  
2 genetic relationships; all analyses were conducted using R software with the adegenete (Jombart  
3 2008) and poppr (Kamvar et al. 2013) packages. The geographical locality from where the  
4 isolates in the present study and previous studies were collected was defined as the priori  
5 population. The optimum number of posterior clusters that all isolates were divided into without  
6 priori population information was determined by *k*-means clustering with the smallest Bayesian  
7 information criterion (BIC). The *k* value sequentially increased from *k* = 1 to *k* = 12 in the  
8 *find.clusters* function (Jombart et al. 2010). This function was performed for 10 independent  
9 trials with 1,000 starting points and 1,000,000 iterations. Subsequently, discriminant analysis  
10 of principal components (DAPC) was conducted to group the isolates into the optimum number  
11 of posterior clusters with 20 PCs, implying >80% of the cumulative proportion of variance.  
12 Next, PCs were applied to discriminant analyses for the detection of genetic variability among  
13 the posterior clusters. The probability of each isolate belonging to each posterior cluster was  
14 also obtained from the results of the DAPC.

### 38 **Theory/calculation**

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41 Multilocus microsatellite genotyping was proved to be a valid tool for discrimination of  
42 *N. caninum* isolates into subtypes related to geographical locations, so it can be helpful for  
43 understanding the clonal expansion and diversity of this parasite worldwide. Thus, in this study,  
44 with cloning-sequencing approach, we applied this system in accurate way to investigate the  
45 genetic relationship of Japanese isolates with worldwide genotypes. In addition, we searched  
46 for mixed alleles which may be underestimated by automated allele sizing and direct  
47 sequencing especially for alleles similar in sizes with different repeat motifs.

### 3. Results and Discussion

Parasite DNA was successfully amplified from the five brain samples for the 12 MS markers. Non-amplified markers from some samples were not considered after three successive nested PCR reactions. Automated allele sizing data of fluorescent PCR products were considered after performing several repetitions of the experiments and obtaining constant data. The alleles were successfully assigned following the protocols described by Pedraza-Díaz et al. (2009) and Regidor-Cerrillo et al. (2013), except for the marker MS1A (Table S1). Because MS10 is a highly informative marker, all alleles identified in the MS10 locus were given equal opportunities to appear in the multilocus genotypes considering the lack of dominance after fragment analysis. Additionally, the MS10 amplified products showed the same PCR product sizes with different allelic repeat motifs after cloning-based sequencing. Similarly, one sample (JPN-10-H 12) showed two alleles in the locus MS5 with nearly equal peaks, which were assigned into two genotypes. In contrast, the locus MS7 showed a high frequency of mixed alleles after clone sequencing, wherein TA (13) was the dominant allele corresponding to the number of positive sequenced clones and fragment analysis prediction (Table S1).

From the five investigated samples, six multilocus genotypes were identified based on similarity and difference between samples as well as the existence of mixed alleles without dominance (Table 1). Mixed two alleles were previously reported from *N. caninum* isolates and only the dominant ones were considered for further analysis (Regidor-Cerrillo et al., 2013). Notably, one investigated sample (JPN-10-H10) showed three mixed alleles from the sequenced clones of MS7 and MS10. Consequently, this difference was attributed to the sequencing method. We referred to cloning-based sequencing with the highest resolution and best discriminative capabilities over the direct DNA sequencing methods used in other studies. Minor mixed subpopulations can be missed by microsatellite genotyping if they represent <20%

1 of the overall parasite population (Juliano et al., 2007; Figan et al., 2018). Although the minor  
2 subpopulations are not important to the geographic distribution, they are extremely important  
3 when linking them to the disease outcome or biological differences.  
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7 Genotypes were divided into 12 posterior clusters ( $k = 12$ ), with the lowest BIC in the  
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9  $k$ -mean clustering (Fig. S1). The scatter plot of DAPC based on the first two linear discriminant  
10 functions showed that five of the seven genotypes from Japan were separated from other  
11 clusters (Fig. 1). These five genotypes were assigned from samples (JPN-10-H10, JPN-10-H11,  
12 JPN-10-H12, JPN-10-H13) obtained during outbreak of an abortion epidemic in a dairy cattle  
13 farm in Shihoro, Hokkaido, Japan (2010). The existence of different *N. caninum* subpopulations  
14 may be attributed to clonal spreading by vertical and horizontal transmission and geographically  
15 related population substructures. Nationwide surveys in Japan have revealed that dogs harbor  
16 antibodies against *N. caninum* (Sawada et al., 1998; Hara et al., 2006; Kubota et al., 2008).  
17 Horizontal transmission between cattle and dogs was proposed in Japan, wherein dogs reared  
18 in dairy farms experienced bovine neosporosis that was seropositive (31.3%) for *N. caninum*  
19 (Sawada et al., 1998). However, the existence of a cluster separate from other *Neospora*  
20 populations has been detected in Japan, showing that there is not only a worldwide population  
21 of *Neospora* but also an endemic population in Japan.  
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41 Another sample obtained from the same town in 2018 (JPN-10-H218) was found to be  
42 genetically related to the previously identified sheep isolate from Japan. The isolate Nc-Sheep,  
43 which has been extensively used as a reference strain for *N. caninum* genetic studies, was  
44 isolated from naturally infected pregnant sheep with no history of abortion (Koyama et al.,  
45 2001). The authors of this study do not mention the location of this sheep isolate, but it may be  
46 the Tokachi area, which neighbors Shihoro in Hokkaido, indicating a predominant clonal  
47 propagation in the country and geographically related population substructures. These two  
48 Japanese isolates were grouped with those from Mexico and Spain (Fig. 1). This result, together  
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1 with other subpopulation clustering of isolates from different countries, suggests that there is  
2 no relationship between the genetic variation of most *Neospora* populations and their  
3 geographical characteristics and that their populations have been mixed. Whole genome  
4 sequencing suggested the existence of a global single lineage, which recently evolved from a  
5 common ancestor and expanded worldwide owing to the movement of cattle (Khan et al., 2019).  
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11 Contrastingly, a close relative of *N. caninum*, *Toxoplasma gondii* possessed a different  
12 population genetic structure with a high degree of genetic diversity compared to the single  
13 genome of *N. caninum* (Khan et al., 2019). The three predominate lineages of *T. gondii* are  
14 classified as either type I, II, or III single nucleotide polymorphisms (SNPs) whereas, types I  
15 and III are second- and first-generation offspring, respectively, resulted from a cross between a  
16 type II strain and one of two ancestral strains (Boyle et al. 2006). However, subsequent studies  
17 incorporating multilocus genotyping techniques for a wider range of isolates, proved the  
18 existence of a more complex population structures (Galal et al. 2019). Frequent genetic  
19 exchanges among the three lineages via sexual recombination in the definitive felid host give  
20 rise to new genotypes (Grigg and Sundar, 2009). In contrast to *T. gondii* which utilizes both the  
21 sexual and asexual transmission to shape its population structure (Grigg and Sundar, 2009), the  
22 vertical transmission is a primary mode for the expansion of *N. caninum* genome (Goodswen  
23 et al. 2013; Khan et al., 2019; Calarco and Ellis, 2020). More recently, comparison of genome-  
24 wide SNPs and insertions/deletions between 7 *N. caninum* isolates was lower than that was  
25 identified between 2 *T. gondii* type I strains, RH and GT1 (Yang et al., 2013; Khan et al., 2019).  
26 Taken together, these data suggest that *Neospora caninum* population genetics is composed of  
27 one species, that have evolved from a common ancestor and expanded as a single clonal lineage  
28 throughout the world.  
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56 *Neospora caninum* was first identified in aborted fetuses in Japan in 1992 (Ogino et al.,  
57 1992). A nationwide survey was conducted to investigate the seroprevalence of *N. caninum*  
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1 among cattle with reproductive disorders (Koiwai et al., 2005a) and among clinically healthy  
2 cattle (Koiwai et al., 2006) during the years 1996 and 1997, respectively. The seropositive and  
3 seronegative cattle determined to be clinically healthy (Koiwai et al., 2006) were tracked for  
4 their reproductive performance during a retrospective 1-year study. The findings revealed that  
5 seropositive animals were 6.1 times more likely to abort compared with seronegative animals,  
6 and 83.6% of seropositive abortions could be attributed to *N. caninum* (Koiwai et al., 2005b).  
7 Furthermore, 1,031 *Neospora*-seropositive cattle and 2,030 aborted fetuses were estimated  
8 during a nationwide survey conducted in 1998, with an estimated total economic loss of  
9 JPY1,362,000,000 (Yamane et al., 2000). Although these data are alarming and indicate a  
10 considerable economic loss due to neosporosis in Japan, there is a gap in the epidemiological  
11 surveys of neosporosis during the last 20 years in Japan (Takashima et al., 2013; Masatani et  
12 al., 2018; Abdelbaky et al., 2020).

#### 33 **4. Conclusions**

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36 Seven Japanese genotypes of *N. caninum* are clustered into two subpopulations, one of  
37 which was unique and distinct from European and American subpopulations. Meanwhile,  
38 another cluster was genetically related to the Mexican and Spanish genotypes. These findings  
39 may be attributed to clonal spreading by vertical and horizontal transmission as well as  
40 geographically related population substructuring. More isolates are needed to determine the  
41 transmission patterns based on genetic relationships, and sero-epidemiological studies are also  
42 required for both cattle and dogs.

#### 56 **Declarations of interest**

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59 The authors declare that they have no competing interests.

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26 **References**  
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- 28  
29 Abdelbaky, H.H., Nishimura, M., Shimoda, N., Hiasa, J., Fereig, R.M., Tokimitsu, H.,  
30  
31 Inokuma, H. and Nishikawa, Y., 2020. Evaluation of *Neospora caninum* serodiagnostic  
32  
33 antigens for bovine neosporosis. *Parasitol. Int.* 75, p.102045.  
34  
35 <https://doi.org/10.1016/j.parint.2019.102045>.  
36  
37  
38  
39 Al-Qassab, S.E., Reichel, M.P. and Ellis, J.T., 2010. On the biological and genetic diversity in  
40  
41 *Neospora caninum*. *Diversity*, 2, 411-438.  
42  
43  
44 Boyle, J.P., Rajasekar, B., Saeij, J.P., Ajioka, J.W., Berriman, M., Paulsen, I., Roos, D.S.,  
45  
46 Sibley, L.D., White, M.W. and Boothroyd, J.C., 2006. Just one cross appears capable  
47  
48 of dramatically altering the population biology of a eukaryotic pathogen like  
49  
50 *Toxoplasma gondii*. *Proc. Natl. Acad. Sci.* 103, 10514-10519.  
51  
52  
53  
54 Calarco, L. and Ellis, J., 2020. Species diversity and genome evolution of the pathogenic  
55  
56 protozoan parasite, *Neospora caninum*. *Infect. Genet. Evol.* 84, 104444.  
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- Du, L., Yang, D., Zhai, T., Gong, P., Zhang, X. and Li, J., 2015. Detection of *Neospora caninum*-DNA in brain tissues from pigeons in Changchun, Jilin (China). *Vet. Parasitol.* 214, 171-173.
- Dubey, J.P. and Schares, G., 2011. Neosporosis in animals—the last five years. *Vet. Parasitol.* 180, 90-108.
- Dubey, J.P., Schares, G. and Ortega-Mora, L.M., 2007. Epidemiology and control of neosporosis and *Neospora caninum*. *Clin. Microbiol. Rev.* 20, 323-367.
- Figan, C.E., Sa, J.M., Mu, J., Melendez-Muniz, V.A., Liu, C.H. and Wellems, T.E., 2018. A set of microsatellite markers to differentiate *Plasmodium falciparum* progeny of four genetic crosses. *Malar. J.* 17, 1-6.
- Galal, L., Hamidović, A., Dardé, M.L. and Mercier, M., 2019. Diversity of *Toxoplasma gondii* strains at the global level and its determinants. *Food Waterborne Parasitol.* 15, p.e00052.
- García-Sánchez, M., Moreno-Gonzalo, J., González-Warleta, M., Mezo, M., Ortega-Mora, L.M. and Regidor-Cerrillo, J., 2020. Isolation and genetic characterization of *Neospora caninum* from naturally infected sheep. *Vet. Parasitol.*, p.109091. <https://doi.org/10.1016/j.vetpar.2020.109091>.
- Goodswen, S.J., Kennedy, P.J. and Ellis, J.T., 2013. A review of the infection, genetics, and evolution of *Neospora caninum*: from the past to the present. *Infect. Genet. Evol.* 13, 133-150.
- Grigg, M.E. and Sundar, N., 2009. Sexual recombination punctuated by outbreaks and clonal expansions predicts *Toxoplasma gondii* population genetics. *Int. J. Parasitol.* 39, 925-933.
- Hara, O.A., Liao, M., Baticados, W., Bannai, H., Zhang, G., Zhang, S., Lee, E., Nishikawa, Y., Claveria, F., Igarashi, M. and Nagasawa, H., 2006. Expression of recombinant dense

1 granule protein 7 of *Neospora caninum* and evaluation of its diagnostic potential for  
2 canine neosporosis. J. Protozool. Res. 16, 34-41.  
3

4 Jombart, T., 2008. Adegenet: a R package for the multivariate analysis of genetic markers.  
5  
6 Bioinformatics 24, 1403-1405. doi: 10.1093/bioinformatics/btn129  
7

8  
9 Jombart, T., Devillard, S., Balloux, F., 2010. Discriminant analysis of principal components: a  
10 new method for the analysis of genetically structured populations. BMC Genet 11, 94.  
11  
12 <https://doi.org/10.1186/1471-2156-11-94>  
13  
14  
15

16  
17 Juliano, J. J., Kwiek, J. J., Cappell, K., Mwapasa, V., & Meshnick, S. (2007). Minority-Variant  
18 pfprt K76T Mutations and Chloroquine Resistance, Malawi. Emerg. Infect. Dis. 13,  
19 873-877.  
20  
21  
22

23  
24 Kamvar, Z.N., Tabima, J.F., Grünwald, N.J., 2014. Poppr: an R package for genetic analysis of  
25 populations with clonal, partially clonal, and/or sexual reproduction. PeerJ 2, e281.  
26  
27 <https://doi.org/10.7717/peerj.281>  
28  
29  
30

31  
32 Khan, A., Fujita, A.W., Randle, N., Regidor-Cerrillo, J., Shaik, J.S., Shen, K., Oler, A.J.,  
33  
34 Quinones, M., Latham, S.M., Akanmori, B.D. and Cleaveland, S., 2019. Global  
35 selective sweep of a highly inbred genome of the cattle parasite *Neospora caninum*.  
36 Proc. Natl. Acad. Sci., 116, 22764-22773.  
37  
38  
39

40  
41 Koiwai, M., Hamaoka, T., Haritani, M., Shimizu, S., Tsutsui, T., Eto, M. and Yamane, I., 2005<sup>a</sup>.  
42 Seroprevalence of *Neospora caninum* in dairy and beef cattle with reproductive  
43 disorders in Japan. Vet. Parasitol. 130, 15-18.  
44  
45  
46

47  
48 Koiwai, M., Hamaoka, T., Haritani, M., Shimizu, S., Kimura, K. and Yamane, I., 2005<sup>b</sup>.  
49 Proportion of abortions due to neosporosis among dairy cattle in Japan. J. Vet. Med.  
50 Sci. 67, 1173-1175.  
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- Koiwai, M., Hamaoka, T., Haritani, M., Shimizu, S., Zeniya, Y., Eto, M., Yokoyama, R., Tsutsui, T., Kimura, K. and Yamane, I., 2006. Nationwide seroprevalence of *Neospora caninum* among dairy cattle in Japan. *Vet. Parasitol.* 135, pp.175-179.
- Koyama, T., Kobayashi, Y., Omata, Y., Yamada, M., Furuoka, H., Maeda, R., Matsui, T., Saito, A. and Mikami, T., 2001. Isolation of *Neospora caninum* from the brain of a pregnant sheep. *J. Parasitol.* 87, 1486-1488.
- Kubota, N., Sakata, Y., Miyazaki, N., Itamoto, K., Bannai, H., Nishikawa, Y., Xuan, X. and Inokuma, H., 2008. Serological survey of *Neospora caninum* infection among dogs in Japan through species-specific ELISA. *J. Vet. Med. Sci.* 70, 869-872.
- Masatani, T., Fereig, R.M., Otomaru, K., Ishikawa, S., Kojima, I., Hobo, S. and Nishikawa, Y., 2018. Seroprevalence of *Cryptosporidium parvum* and *Neospora caninum* in cattle in the southern Kyushu region of Japan. *Parasitol. Int.* 67, 763-767.
- Medina-Esparza, L., Regidor-Cerrillo, J., García-Ramos, D., Álvarez-García, G., Benavides, J., Ortega-Mora, L.M. and Cruz-Vázquez, C., 2016. Genetic characterization of *Neospora caninum* from aborted bovine foetuses in Aguascalientes, Mexico. *Vet. Parasitol.* 228, 183-187.
- Ogino, H., Watanabe, E., Watanabe, S., Agawa, H., Narita, M., Haritani, M. and Kawashima, K., 1992. Neosporosis in the aborted fetus and newborn calf. *J. Comp. Pathol.* 107, 231-237.
- Pedraza-Díaz, S., Marugán-Hernández, V., Collantes-Fernández, E., Regidor-Cerrillo, J., Rojo-Montejo, S., Gómez-Bautista, M. and Ortega-Mora, L.M., 2009. Microsatellite markers for the molecular characterization of *Neospora caninum*: application to clinical samples. *Vet. Parasitol.* 166, 38-46.
- Regidor-Cerrillo, J., Díez-Fuertes, F., García-Culebras, A., Moore, D.P., González-Warleta, M., Cuevas, C., Schares, G., Katzer, F., Pedraza-Díaz, S., Mezo, M. and Ortega-Mora,

- 1 L.M., 2013. Genetic diversity and geographic population structure of bovine *Neospora*  
2 *caninum* determined by microsatellite genotyping analysis. PLoS One, 8, p.e72678.  
3  
4 Regidor-Cerrillo, J., Horcajo, P., Ceglie, L., Schiavon, E., Ortega-Mora, L.M. and Natale, A.,  
5  
6  
7 2020. Genetic characterization of *Neospora caninum* from Northern Italian cattle  
8  
9 reveals high diversity in European *N. caninum* populations. Parasitol. Res. 1-10.  
10  
11 <https://doi.org/10.1007/s00436-020-06642-2>.  
12  
13  
14 Reichel, M.P., Ayanegui-Alcérreca, M.A., Gondim, L.F. and Ellis, J.T., 2013. What is the  
15  
16 global economic impact of *Neospora caninum* in cattle—the billion dollar question. Int.  
17  
18 J. Parasitol. 43, 133-142.  
19  
20  
21 Sawada, M., Park, C.H., Kondo, H., Morita, T., Shimada, A., Yamane, I. and Umemura, T.,  
22  
23  
24 1998. Serological survey of antibody to *Neospora caninum* in Japanese dogs. J. Vet.  
25  
26 Med. Sci. 60, 853-854.  
27  
28  
29 Takashima, Y., Takasu, M., Yanagimoto, I., Hattori, N., Batanova, T., Nishikawa, Y. and Kitoh,  
30  
31 K., 2013. Prevalence and dynamics of antibodies against NcSAG1 and NcGRA7  
32  
33 antigens of *Neospora caninum* in cattle during the gestation period. J. Vet. Med. Sci.  
34  
35 13-0198.  
36  
37  
38 Yamane, I., Koiwai, M., Haritani, M. and Hamaoka, T., 2000. Economic losses from *Neospora*  
39  
40 *caninum* infection in dairy cattle in Japan. J. Jpn. Vet. Med. Assoc. 53, 67-69.  
41  
42 <https://doi.org/10.12935/jvma1951.53.67>  
43  
44  
45 Yamage, M., Flechtner, O. and Gottstein, B., 1996. *Neospora caninum*: specific oligonucleotide  
46  
47 primers for the detection of brain" cyst" DNA of experimentally infected nude mice by  
48  
49 the polymerase chain reaction (PCR). J. Parasitol. 82, 272-279.  
50  
51  
52 Yang, N., Farrell, A., Niedelman, W., Melo, M., Lu, D., Julien, L., Marth, G.T., Gubbels, M.J.  
53  
54 and Saeij, J.P., 2013. Genetic basis for phenotypic differences between different  
55  
56 *Toxoplasma gondii* type I strains. BMC genomics 14, 1-19.  
57  
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6 **Figure legends**  
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8 **Figure 1** (A) Scatter plot of discriminant analysis of principal components based on the first  
9 two linear discriminant functions in  $k = 12$ . The 12 posterior clusters are shown in different  
10 colors. (B) Bar plot of posterior cluster membership probability for the 114 isolates. The bar  
11 plots show the probability of the isolate belonging to each posterior cluster. The colors are the  
12 same as those of the posterior clusters in (A). The priori population of the isolates is shown at  
13 the top of the bar plot, and the name of the isolate is indicated under the bar plot.  
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**Table 1:** Multilocus microsatellite genotypes obtained from fetal brain tissues included in this study

Genotypes ID <sup>a</sup>	Geographic Origin (Country)	Host sampled	Microsatellite Genotype <sup>b</sup>								
			MS4	MS5	MS6A	MS6B	MS7 <sup>d</sup>	MS8	MS10	MS12	MS21
JPN-10-H10.1 <sup>e</sup>	Japan	Cattle	10	15	12	11	12	13	6.14.8	14	6
JPN-10-H10.2 <sup>e</sup>	Japan	Cattle	10	15	12	11	12	13	7.12.9	14	6
JPN-10-H10.3 <sup>e</sup>	Japan	Cattle	10	15	12	11	12	13	7.13.9*	14	6
JPN-10-H11											
JPN-10-H13											
JPN-10-H12.1 <sup>e</sup>	Japan	Cattle	10	15	NA	NA	12	13	7.14.9*	14	6
JPN-10-H12.2 <sup>e</sup>	Japan	Cattle	10	16	NA	NA	12	13	7.14.9*	14	6
JPN-10-H218	Japan	Cattle	13	9	NA	NA	12	NA	NA	15	6

<sup>a</sup> Sample identification: JPN (Japan) - **genotype** number (ID).

<sup>b</sup> Allele assignment of MS markers are expressed as the number of repeats following the criteria described by Regidor-Cerrillo et al. (2013).

<sup>d</sup> Only dominant alleles from cloning and automated allele sizing were included for this marker.

<sup>e</sup> Mixed alleles were identified, and dominant alleles could not be assigned.

\*Newly described MS10 alleles obtained in this study.

NA: not amplified

Figure 1

