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Temporal and within-plant distribution of the parasitoid and predator complexes associated with *Acyrthosiphon pisum* and *A. kondoi* (Homoptera: Aphididae) on alfalfa in Japan

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Abstract

Seasonal abundance and within-plant distribution of *Acyrthosiphon* spp. and their natural enemies were investigated on alfalfa in Obihiro, Hokkaido, Japan in 2001. Densities of *A. pisum* and *A. kondoi* peaked in early June. Coccinellid and syrphid predators appeared to respond to aphid numbers and were synchronized with aphid populations. Two species of braconid (aphidiine) parasitoids (*Aphidius ervi* and *Praon barbatum*) were collected. Parasitism rates for each aphid species were variable through the season, but the estimated number of parasitized aphids reached a peak at the same time as overall aphid densities. After the first harvesting of alfalfa (14 June) aphid densities became very low, and coccinellid and syrphid predators were not found. In contrast to the coccinellid and syrphid predators, the heteropteran predators, *Nabis stenoferus* and *Orius* spp. became common during summer. Aphids, immature parasitoids (live parasitized aphids and mummies), *Coccinella septempunctata* and syrphids occurred more on the upper parts of alfalfa stems than on the lower parts, but this trend was not found for *Hippodamia tredecimpunctata*. The temporal and spatial distributions of members of the aphidphagous guild are likely to influence natural enemy–aphid relationships and intraguild interactions.

Key words: Aphidius ervi; Coccinella septempunctata; hyperparasitoid; intraguild interaction; Praon barbatum

INTRODUCTION

The pea aphid, Acyrthosiphon pisum (Harris), and the blue alfalfa aphid, A. kondoi Shinji (Homoptera: Aphididae), are serious pests of alfalfa, Medicago sativa L., and other leguminous crops in many parts of the world (Stern et al., 1980; Summers and Coviello, 1984; Lenssen et al., 1991; Lane and Walters, 1991). Their infestation causes deformation of leaves and stunting of the plants (Schepers, 1989; Cuperus et al., 1982), and they are also vectors of plant viruses (Sylvester, 1989; Bosque Perez and Buddenhagen, 1990). These aphids are associated with a large assemblage of insect natural enemies such as Hymenopteran parasitoids, and Coccinellid, Syrphid, Nabid, Anthocorid, and Chrysopid predators (Mackauer and Finlayson, 1967; Takada, 1968; Wheeler, 1977; Ekbom, 1994; Brodeur and Rosenheim, 2000).

The ladybird beetle Coccinella septempunctata

L. (Ekbom, 1994) and the braconid parasitoids *Aphidius ervi* Haliday and *Praon barbatum* Mackauer are reported as important natural enemies of *Acyrthosiphon* species on alfalfa (Carver, 1989; Hughes, 1989; Takahashi, 1996; Milne, 1999; Senoo et al., 2002). However, the temporal and spatial distribution of the whole natural enemy complex in relation to the aphids is not clearly understood at present.

The degrees of temporal and spatial associations between the target pest and each natural enemy species are likely to influence the ability of a natural enemy to suppress the pest population. Furthermore, similarities in the temporal and spatial distributions of different members of the natural enemy guild are important for inferring intraguild relationships, which may be useful for understanding the indirect effects of intratrophic predator interactions on aphid population dynamics. For instance, intraguild predation (IGP), trophic interactions

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between natural enemies sharing the same resource, may be common in agroecosystems and may influence the level of herbivore population suppression (Rosenheim et al., 1993, 1995; Rosenheim, 1998; Raymond et al., 2000). Interactions between various combinations of intraguild natural enemies associated with aphids are known, such as IGP between parasitoids (Mackauer, 1990), between predators (Wheeler, 1977; Atim and Graham, 1984), between predators and parasitoids (Wheeler et al., 1968; Colfer and Rosenheim, 1995; Wells et al., 2001; Nakashima and Senoo, 2003; Nakashima et al., 2004) and between entomopathogens and parasitoids (Powell et al., 1986; Brooks, 1993). Thus, identifying the spatio-temporal patterns of each member of an aphidophagous guild is important to assess the functional role of the guild in controlling aphid abundance.

To determine temporal and spatial relationships among aphids and their natural enemy complex, we measured aphid and predator abundance and levels of parasitism of two *Acyrthosiphon* species in an alfalfa field. We also investigated the patterns of within-plant distribution of aphids, immature parasitoids (live parasitized aphids and mummies) and the major predator species. The roles of parasitoid and predator complexes in the suppression of aphid populations and possible intraguild interactions are discussed.

MATERIALS AND METHODS

Field site. Field studies were conducted throughout the 2001 growing season in an alfalfa field (4 ha) at Obihiro University of Agriculture and Veterinary Medicine in Obihiro (42°92′N, 143°22′E), Hokkaido, Japan. No insecticides were used in the field. The alfalfa was harvested on 13 June, 29 July and 19 September in 2001.

Seasonal abundance. Samples were obtained approximately every 10 d from 12 May to 17 November 2001. Ten sweeps within a 1 m×1 m quadrat were taken with a 40 cm diameter sweepnet at 15 random locations within the alfalfa field on each sampling date. The heights of five selected stems of alfalfa in each quadrat were also measured to monitor the growth rate of the alfalfa crop. Sweep samples were kept in plastic boxes (28×13×7 cm), placed in coolers with ice, and transported to the laboratory for sorting and counting the num-

bers of aphids and their natural enemies present. Potential insect predators of *A. pisum* and *A. kondoi*, such as coccinellid larvae and adults and hemipteran (Nabidae and Anthocoridae) adults were identified. Neuropteran larvae were not identified to species. To identify the species of syrphid larvae, those collected from the field on 5 June were reared until adult emergence in a growth room at $20\pm1^{\circ}\text{C}$ and a photoperiod of 16L:8D. These larvae were reared in 9 cm diameter Petri dishes, and provided with 100-200~A.~pisum as food, which was replaced every 2 d until pupation.

Parasitism rates of *A. pisum* and *A. kondoi* were evaluated by rearing live aphid samples in the laboratory. The aphids were reared on broad bean seedlings, *Vicia fabae* L. (Fabaceae) (var. minor), at the 4 to 6-leaf stage, in plant pots measuring 7.0 cm in height and 5.5 cm in diameter at the bottom and 7.5 cm in diameter at the top. Each pot contained 3–4 seedlings, grown in vermiculite, and was kept in a cylindrical acrylic tube (diameter 8.5 cm, height 25.0 cm) with the top covered by mesh to enclose the infested seedlings securely.

For each sampling unit, each aphid species (A. pisum or A. kondoi) was separately transferred to the plants with a fine paintbrush. In general, 30 randomly selected aphids of each species were transferred per sampling unit, whilst all aphids were transferred when the number of aphids was less than 30. The rate of parasitism for each sample unit was estimated by dividing the number of mummified aphids that developed by the number of reared aphids. Plants with aphids were maintained for 10–14 d in a growth room at 20±1°C and a photoperiod of 16L:8D.

Within-plant distribution. To evaluate the vertical distribution of predators and unparasitized and parasitzed aphids, insects were collected from two strata of alfalfa stems in 20, randomly chosen, quadrats $(1 \text{ m} \times 1 \text{ m})$ in the field. In each quadrat, 15, randomly selected, alfalfa stems were cut 25 cm from the base of the stem and at the base (the alfalfa stem heights were 50.1 cm on average). The upper and lower stems taken from each quadrat were placed separately in boxes $(15\times10\times5 \text{ cm})$ and transported to the laboratory. Aphids and predators on each stratum were counted, and aphids collected from each part of the plant were reared on broad bean seedlings to estimate rates of parasitism as described above. Sampling was done

on 4 and 5 June 2001 when aphids were most abundant.

To characterize the vertical distribution of mummies that contained either primary parasitoids or hyperparasitoids, 100 alfalfa stems were randomly selected and cut at the base. The height of each alfalfa stem and the height at which mummies were present were recorded. Collected mummies were labeled and kept individually in gelatine capsules at 20±1°C and a photoperiod of 16L:8D until adult emergence, when emerged primary parasitoids and hyperparasitoids were identified. The sampling was conducted on 12 June.

Data analysis. Insect densities and rates of parasitism were analyzed using the Mann-Whitney *U* test and Wilcoxon's signed rank test. Densities of predator species on upper and lower halves of alfalfa stems were compared using the Mann-Whitney *U* test with correction (the sequential Bonferroni method; Rice, 1989). The heights of primary parasitoid and hyperparasitoid mummies on the plants were compared using the Kruskal Wallis test. All tests were conducted with a 5% level of significance.

RESULTS

Densities of *A. pisum* and *A. kondoi* increased during May and early June and peaked on 4 June. After the first harvest, the population density decreased and remained low until the end of the season $(0-4.4 \text{ and } 0-3.3/\text{m}^2 \text{ for } A. \text{ pisum} \text{ and } A. \text{ kondoi}$, respectively) (Fig. 1). On the first three sampling dates, densities of *A. kondoi* were significantly higher than those of *A. pisum* (Mann-Whitney U test, p < 0.05), but no difference was found on subsequent dates.

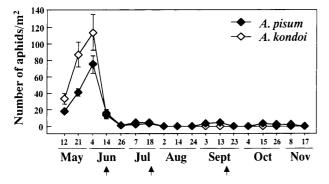


Fig. 1. Changes in densities of *Acyrthosiphon pisum* and *A. kondoi*. Arrows below calendar and bars with plots indicate days of harvesting and SE, respectively.

Natural enemies collected in the survey are shown in Table 1. Parasitism rates of *A. pisum* by *A. ervi* at the beginning of the season (early May) reached 31% (Fig. 2). The rates due to *A. ervi* were higher than those caused by *P. barbatum* throughout the season, for both *A. pisum* and *A. kondoi*

Table 1. Natural enemies of *Acyrthosiphon pisum* and *A. kondoi* collected in the alfalfa field in Hokkaido, Japan

Family of predators and parasitoids ^a	Species of predators or parasitoids
Parasitoids:	
Aphididae	Aphidius ervi Haliday
	Praon barbatum Mackauer
Predators:	
Coccinelidae	Coccinella septempunctata L.
	Hippodamia tredecimpunctata Capra
	Propylea quatuordecimpunctata (L.) ^b
Syrphidae ^c	Metasvrphus (Eupeodes)
	corollae (Fabricius)
	Episyrphus balteatus (de Geer)
	Metasyrphus ferquens Matsumura
	Syrphus vitripennis Meigen
Anthocoridae	Orius sauteri Poppius
	Orius minutus L.
Nabidae	Nabis stenoferus Hisao

^a Five Neuropteran larvae were sampled in July and September, but not identified.

^c Adults were reared from field collected larvae.

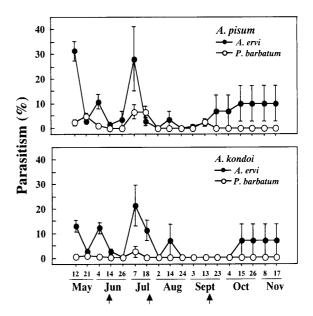


Fig. 2. Changes in parasitism rates of *Acyrthosiphon pisum* and *A. kondoi* by *Aphidius ervi* and *Praon barbatum*. Arrows and bars indicate days of harvesting and SE, respectively.

^b Two adults were sampled in June.

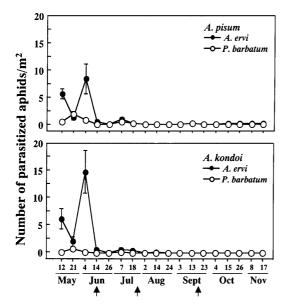


Fig. 3. Changes in estimated densities of Acyrthosiphon pisum and A. kondoi parasitized by Aphidius ervi and Praon barbatum. Arrows and bars indicate days of harvesting and SE, respectively.

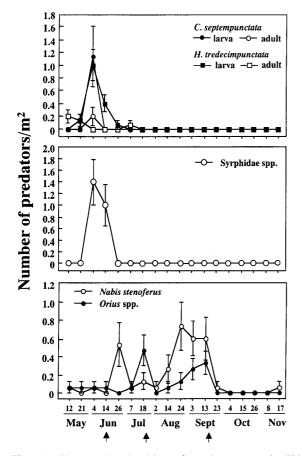


Fig. 4. Changes in densities of predators, coccinellids (top), syrphids (middle), and heteropterans (bottom). Arrows and bars indicate days of harvesting and SE, respectively.

(Mann-Whitney U test, p < 0.05). Estimated numbers of aphids parasitized by A. ervi and P. barbatum were also restricted in this period (Fig. 3). After the first harvest, parasitism rates due to A. ervi varied over the season (Fig. 2). Occurrence of coccinellids (C. septempunctata and $Hippodamia\ tredecimpunctata$ Capra) and syrphids (larvae) were limited during late May and early June (Fig. 4). In contrast to coccinellids and syrphids, heteropteran predators were abundant from late June to late September when the aphid population was small (Figs. 1 and 4).

Within-plant distributions of aphids and immature parasitoids (live parasitized aphids) were concentrated in the upper half of alfalfa stems (Fig. 5). Aphidius ervi was the only parasitoid species found in this part of the investigation. Both the number of parasitized aphids and rates of parasitism were also greater in the upper half of the plant for each aphid species (Wilcoxon's signed rank test, p < 0.01) (Fig. 5). Also, all mummies of A. ervi were on the upper half of the stem. The average height of alfalfa stems was 64.3 ± 1.2 cm (mean \pm SE, n=100) on 12 June. Dendrocerus carpenteri (Curtis)

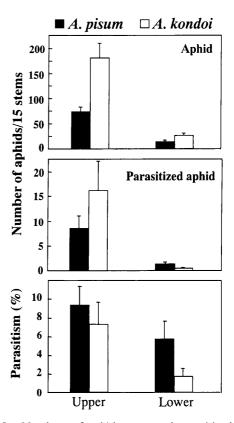


Fig. 5. Numbers of aphids (top) and parasitized aphids (middle), and rates of parasitism (bottom) in lower (<25 cm) and upper (>25 cm) parts of alfalfa stems. Bars indicate SE.

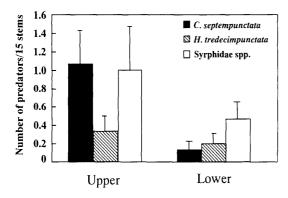


Fig. 6. Number of predators in lower (<25 cm) and upper (>25 cm) parts of alfalfa stems. Bars indicate SE.

(Megaspilidae) and Asaphes suspensus (Nees) (Pteromalidae) emerged from A. ervi mummies. Average heights on the plant of mummies containing A. ervi, D. carpenteri and A. suspensus were $50.86\pm2.0 \,\mathrm{cm}$ (n=62), $54.5\pm7.4 \,\mathrm{cm}$ (n=9) and $48.6\pm3.0 \,\mathrm{cm}$ (n=21), respectively. These heights for the different species were not significantly different (Kruskal Wallis test, p>0.05).

Within-plant distributions were variable amongst predator species (Fig. 6). Numbers of C. septempunctata and syrphid species (larvae) were significantly greater on the upper parts of alfalfa stems (Wilcoxon's signed rank test, p < 0.01). On the other hand, no significant difference between upper and lower parts of the stem in the number of *H. tre*decimpunctata was found (Wilcoxon's signed rank test, p > 0.05). Within the upper part of the stems, there were significantly more C. septempunctata than H. tredecimpunctata (Mann-Whitney U test with correction of the sequential Bonferroni method, p < 0.05), but there were no significant differences between the numbers of C. septempunctata and syrphidae species or between numbers of H. tredecimpunctata and syrphidae species. Within the lower half of the plant, statistically similar numbers of ladybirds and syrphids were recorded.

DISCUSSION

Numbers of *A. pisum* and *A. kondoi* were restricted during May and early June (Fig. 1). The low aphid densities during summer were probably because of decreased reproduction due to high temperatures (Siddiqui et al., 1973; Takahashi, 1996). The aphid population trend up until summer was roughly consistent with that recorded in an

earlier survey in 2000 (Senoo et al., 2002). An increase in aphid densities during autumn, however, was not found in this survey, although previous studies showed that another peak normally appears from October to November (Takahashi, 1996; Senoo et al., 2002). We are unsure of the reason for this difference, but we note that higher rates of parasitism by *A. ervi* during summer and autumn, 2001 (20–30% in July and 8–10% in autumn) may have prevented a second increase in aphid numbers (Fig. 2). In contrast, parasitization by *A. ervi* was scarce from summer to the end of the season in the earlier survey (Senoo et al., 2002).

Parasitism rates of *A. pisum* by *A. ervi* at the beginning of the season were relatively high (Fig. 2). This parasitization, probably due to adults of the overwintering generation, would have had a significant impact on the initial aphid density. The parasitism during this part of the season may contribute to decreasing the initial number of aphids colonising crops the following spring. Comparison with data from our previous survey suggests that the dominant parasitoid species can change from year to year. The parasitism due to *P. barbatum* was three times higher than that caused by *A. ervi* at the peak in June 2000 (Senoo et al., 2002).

Seasonal distributions of predator species were divided into two distinct phases. Number of coccinelids and syrphids (larvae) were restricted during May and June. Ladybird species were also dominated by C. septempunctata and H. tredecimpunctata in the previous year, but syrphid populations appear to be more variable from year to year because few were collected in 2000 compared with 2001 (Nakashima, unpublished data). *Nabis* and Orius predators are common in alfalfa fields, and appear during summer when aphid densities are very low (Nielson and Henderson, 1959; Wheeler, 1977), as shown in this study. These groups were considered to be less effective predators on the basis of their feeding capacities against aphids (Nielson and Henderson, 1959), and are regarded as polyphagous predators (Wheeler, 1977; Nagai, 1993). Nagai (1993) suggested that aphids are sub-optimal prey for Orius species. Although N. stenoferus and the two Orius species may help to delay a new build up of the aphid population, relationships with other prey species in alfalfa need be determined.

Within-plant distributions of aphids and imma-

ture parasitoids (live parasitized aphids) were concentrated in the upper half of alfalfa stems (Fig. 5). This distribution pattern was probably due to the preference of these aphids for young alfalfa leaves or buds because the upper part of the plant is where new buds arise, and younger leaves usually have a higher concentration of primary metabolites (e.g. Merritt, 1996). Also, all mummies of *A. ervi* were on the upper half of the stems. These results suggest that adult parasitoids allocate more foraging time in the upper parts of alfalfa plants and parasitized aphids predominantly mummified there.

Differences in vertical distribution amongst aphidophagous coccinellids may occur due to microhabitat preferences (Ewert and Chiang, 1966; Hemptinne et al., 1988) or interference between predator species (Schellhorn and Andow, 1999). It is not clear, however, whether the different patterns of within-plant distribution between the two ladybird species in our study were due to their microhabitat preferences or mutual interference.

In this crop system, aphid densities are high over a limited period (late May and early June) (Fig. 1 and Senoo et al., 2002). Thus, members of the aphidophagous guild occurring during this economically important period seem to play an important role in aphid suppression. Our data suggest that A. ervi, C. septempunctata and syrphids contribute to suppression of the aphid population significantly more than other species because of their abundance and the similarity of their temporal and spatial distribution patterns to those of the aphids. The temporal aggregation of some species of natural enemies may be caused by attraction to induced plant volatiles emitted after aphids have damaged the alfalfa. Aphidius ervi (Du et al., 1998; Powell et al., 1998) and C. septempunctata (Ninkovic et al., 2001) were reported to be attracted to herbivore-induced volatiles. The aggregation of the parasitoid and predator guild may play an important role in the population dynamics of aphids before the first harvest.

A high degree of temporal and spatial overlap in the distributions of a number of natural enemies may increase intraguild interactions. Such interactions could be critical events in the population dynamics of aphids, because they may indirectly affect suppression of aphid populations by aphidophagous guilds. For instance, it is reported that intraguild predation negatively affects biological control of aphid populations (Colfer and Rosenheim, 1995; Ferguson and Stiling, 1996; Yasuda and Shinya, 1997). However, intraguild interactions sometimes facilitate aphid suppression (Losey and Denno, 1998, 1999). From the results of our survey, these types of interaction may occur among A. ervi, C. septempunctata and syrphids. Acyrthosiphon pisum parasitized by A. ervi are also consumed by C. septempunctata (Nakashima, unpublished data) as in other parasitoid-predator systems (Wheeler et al., 1968; Takizawa et al., 2000; Wells et al., 2001). The direction and intensity of intraguild interactions in limited time and space need to be determined for further understanding of the role of the whole aphidophagous guild in biological aphid control.

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