Investigating Enteric Methane Mitigation: Assessing the Impact of Various Materials on Ruminants' Fermentation, Intake, and Digestibility

2024

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消化管からのメタン発生抑制:

種々の素材が反芻動物の胃内発酵、飼料摂取量および消化率に及ぼす影響の検討

令和6年

(2024)

带広畜産大学大学院畜産学研究科畜産科学専攻博士後期課程

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General Introduction

Farming animals, particularly within the ruminant sector, stands as a linchpin in the landscape of regenerative agriculture and global food security (Adegbeye et al., 2020). Ruminants, exemplified by cows, play an irreplaceable role in providing substantial quantities of milk and meat globally, thereby fulfilling a fundamental role in addressing the world's dietary needs (Sekaran et al., 2021). These animals underscore their significance by contributing a staggering 51% of the total protein derived from the global livestock industry. Within this notable contribution, 67% is attributed to milk, while the remaining 33% is allocated to meat, emphasizing the crucial role of ruminants in shaping the nutritional landscape (*OECD-FAO Agricultural Outlook 2023-2032*, 2023).

However, amidst this indispensable contribution, the global population is undergoing rapid expansion. Projections indicate that the world's populace will reach around 9.7 billion by 2050, with further estimates soaring to an estimated 10.9 billion by 2100. This burgeoning population poses a consequential increase in the demand for meat and milk, projected to surge by 14% for meat and over 30% for milk by 2030 (*OECD-FAO Agricultural Outlook 2023-2032*, 2023). This surge in demand places additional pressure on the livestock sector, particularly ruminant farming, to meet the nutritional needs of the growing global population.

Yet, as ruminants strive to meet this escalating demand, they present a substantial challenge concerning environmental sustainability. These animals emerge as significant emitters of greenhouse gases (GHGs), with 44% composed of methane (CH₄) and the remaining part almost evenly divided between 27% carbon dioxide (CO₂) and 29% Nitrous oxide (N₂O). In 2019, the agriculture sector contributed 22% of the total anthropogenic GHG emissions produced (*FAO Publications Catalogue 2023*, 2023;

Intergovernmental Panel on Climate Change (IPCC), 2023). The notable contribution of ruminants to GHG emissions makes them primary drivers of climate change, a phenomenon that poses severe threats to the well-being of both current and future generations (Althor et al., 2016).

The rapid escalation of GHG levels raises profound concerns, prompting warnings from institutions like the World Bank (*Reality Check: Lessons from 25 Policies Advancing a Low-Carbon Future*, 2023). If current trends persist, the Earth is on a trajectory to become 4 °C warmer, surpassing a critical threshold. This alarming temperature rise is associated with various devastating consequences, including recurrent and severe heatwaves, rampant wildfires, prolonged droughts, catastrophic floods, and an array of social disruptions such as famines and mass migrations (*Reality Check: Lessons from 25 Policies Advancing a Low-Carbon Future*, 2023; UN Climate Change, 2021).

In the complex interplay of addressing the rising global demand for nutrition while mitigating the environmental impacts of GHG emissions, the livestock industry, particularly in the ruminant sector, faces a pivotal challenge. Ruminant animals, exemplified by cows, produce GHGs such as CO₂ and CH₄ as natural by-products of digestion, a process called enteric fermentation. This intricate process involves methanogenic archaea, small organisms in their stomachs, producing CH₄, which is then released mostly when the animal burps. Unfortunately, this release signifies a substantial loss, amounting to up to 12% of the energy they consume, making it a notable inefficiency in the industry (Pickering et al., 2015).

In light of increasing public apprehension regarding the detrimental impacts of enteric CH₄ emissions on the efficacy and sustainability of livestock production, the scientific community, particularly in the realm of animal husbandry, is actively exploring strategies

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for effective management. This quest involves a multifaceted approach, integrating scientific research, technological innovations, and conscientious agricultural practices. The primary goal is to mitigate GHG emissions from ruminants without compromising the environment, farmers' profits, or the animals' welfare (Adegbeye et al., 2020; Honan et al., 2021).

By fostering innovation and adopting sustainable practices, the industry strives to transition toward more efficient production systems. This approach not only ensures the fulfillment of essential nutritional needs for a growing global population but also contributes significantly to safeguarding the future of our planet. Embracing these solutions not only meets the immediate demands for food but also establishes a harmonious balance, ensuring the well-being of both humanity and the environment in the long run.

The efforts of research groups worldwide to combat CH₄ emissions and optimize production methods are noteworthy (Beauchemin et al., 2020; Hopkins et al., 2016). Among the most effective methods is dietary intervention through feed additives and by-products. Historically focused on improving animal health, dietary changes are now seen as promising for mitigating CH₄ emissions due to their impact on rumen microbes (Beauchemin et al., 2020; Hopkins et al., 2016). This approach offers a dual advantage, enhancing livestock production efficiency while preserving the environment (Beauchemin et al., 2020).

Numerous feed additives have undergone extensive research to mitigate CH₄ emissions. However, current studies, such as those available at Beauchemin et al. (2022) and Ahmed et al. (2022), indicate that there are currently no fully effective and sustainable options. A significant challenge faced by animal husbandry researchers is achieving 'clean,

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green, and ethical' production. This necessitates that livestock products be free from chemical or synthetic compounds in ruminants' feed, aligning with animal welfare guidelines to minimize environmental impact and ensure sustainable and responsible livestock farming (Guyomard et al., 2021).

Due to the prohibition of synthetic compounds, such as antibiotics, in livestock feed across several countries, there's a growing global interest in utilizing bioactive components from plants and their by-products as alternative feed additives (Guyomard et al., 2021; Sandström et al., 2022). These include natural compounds like polyphenols, flavonoids, saponins, tannins, and organic acids, which have the potential to influence animal behavior, alter rumen fermentation, and inhibit various rumen microorganisms. While there are promising results in inhibiting rumen methanogens, challenges emerge when transitioning from *in vitro* studies to *in vivo* applications (Beauchemin et al., 2020; Kobayashi et al., 2016).

Utilizing human consumption by-products as food for ruminants is considered a potential way to replace conventional feed in the agriculture sector. Coffee, the second-most consumed beverage globally after water, generates various by-products during its processing stages. Spent coffee waste (SCW) is a major by-product containing significant amounts of organic compounds, lipids, phenolic compounds, and polysaccharides (Batbekh et al., 2023; Otálora et al., 2020). SCW, with its bioactive compounds, finds application in various industries, including biofuel, beauty products, construction, and animal feed (Carta et al., 2022; Kang et al., 2022; Wong et al., 2022; Xu et al., 2007). However, for SCW to serve as an alternative feed source for the livestock sector, consideration must be given to dosages, methods, and effects on animal health and

performance (Batbekh et al., 2023; Xu et al., 2007). The chemical composition of coffee by-products from previous studies described in Table 1.

Previous research extensively explored SCW as a potential ruminant feed, revealing various advantages and disadvantages, dependent on the applied dosage (Badarina et al., 2013; Batbekh et al., 2023; Campos-Vega et al., 2015a; Seo et al., 2015; Xu et al., 2007). High doses of SCW might impact nutrient digestibility, potentially affecting animal health and production (Batbekh et al., 2023; Xu et al., 2007). Understanding optimal dosage effects is crucial when considering SCW in ruminant diets, considering factors like cost, availability, processing, and compatibility with other dietary components (Xu et al., 2007). Implementing appropriate processing methods, such as ensiling, is vital to mitigating negative effects. Technologies for creating superior animal feed using SCW are crucial, especially in countries like Japan, enabling long-term storage. Silage production, combining wet and dry by-products, gains interest due to reduced effluent risk, improved rumen function, and longer storage potential. The lactic acid and soybean curd in silage enhance fermentation quality and digestibility and reduce CH4 production.

Table1

Chemical composition of a	coffee by-products.
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Parameters (%)	Coffee pulp	Coffee husk	Silver skin	Coffee spent
Cellulose	63.0 ± 2.5	43.0 ± 8.0	17.8 ± 6.0	8.6 ± 1.8
Hemicellulose	2.3 ± 1.0	7.0 ± 3.0	13.1 ± 9.0	36.7 ± 5.0
Protein	11.5 ± 2.0	8.0 ± 5.0	18.6 ± 4.0	13.6 ± 3.8
Fat	2.0 ± 2.6	0.5 ± 5.0	2.2 ± 1.9	ND
Total fiber	60.5 ± 2.9	24 ± 5.9	62.4 ± 2.5	ND
Total polyphenols	1.5 ± 1.5	0.8 ± 5.0	1.0 ± 2.0	1.5 ± 1.0
Total sugars	14.4 ± 09	58.0 ± 20.0	6.65 ± 10	8.5 ± 1.2
Pectic substance	6.5 ± 1.0	1.6 ± 1.2	0.02 ± 1.0	0.01 ± 0.005
Lignin	17.5 ± 2.2	9.0 ± 1.6	1.0 ± 2.0	0.05 ± 0.05
Tannins	3.0 ± 5.0	5.0 ± 2.0	0.02 ± 0.1	0.02 ± 0.1
Chlorogenic acid	2.4 ± 1.0	2.5 ± 0.6	3.0 ± 0.5	2.3 ± 1.0
Caffeine	1.5 ± 1.0	1.0 ± 0.5	0.03 ± 0.6	0.02 ± 0.1

From Mussatto et al. (2011b), Franca et al. (2009) and Murthy and Madhava Naidu (2010).

ND: not determined.

To address these challenges, researchers explore anti-methanogenic materials. In past studies, organic acids such as fumarate demonstrated the capability to reduce CH₄ in both *in vitro* and *in vivo* (Bayaru et al., 2001; Carro & Ranilla, 2003; Gheller et al., 2020; Islam & Lee, 2019; Palangi & Macit, 2021). Decreasing fumarate in the rumen alters how hydrogen is processed, leading to more propionate production and less CH₄ (Islam & Lee, 2019). Other organic acids, like citrate and itaconate, common in the swine and poultry industries, may also regulate rumen microbiota and metabolic activity, influencing CH₄ production. Additionally, acrylate, a substance in rumen metabolic pathways, is being explored as a potential additive to reduce CH₄ production in the rumen (Bampidis & Robinson, 2006; Oryza,S. et al., 2021; Oryza,S. et al., 2021b).

This growing interest in natural feed additives aims to enhance rumen fermentation efficiency, reduce GHGs, and ensure the sustainability and ethical practices of livestock production (Ku-Vera et al., 2020; Ponnampalam et al., 2023; Thornton et al., 2010). Exploring these innovative solutions seeks to strike a balance between effective CH₄ reduction and maintaining optimal animal performance and welfare. The availability of multiple additives is key to the widespread and sustainable implementation of methanereducing strategies globally.

This integrated study comprises two parts. The first part explores the use of by-products from human consumption as feed and additives for ruminants, examining both raw and ensiled forms under *in vitro* conditions. The latter part concentrates on *in vivo* conditions, exploring the application of novel CH₄ suppressors.

Reference

- Adegbeye, M. J., Ravi Kanth Reddy, P., Obaisi, A. I., Elghandour, M. M. M. Y., Oyebamiji, K. J., Salem, A. Z. M., Morakinyo-Fasipe, O. T., Cipriano-Salazar, M., & Camacho-Díaz, L. M. (2020). Sustainable agriculture options for production, greenhouse gasses and pollution alleviation, and nutrient recycling in emerging and transitional nations An overview. *Journal of Cleaner Production*, 242, 118319. https://doi.org/10.1016/j.jclepro.2019.118319
- Ahmed, E., Batbekh, B., Fukuma, N., Hanada, M., & Nishida, T. (2022). Evaluation of Different Brown Seaweeds as Feed and Feed Additives Regarding Rumen Fermentation and Methane Mitigation. *Fermentation*, 8(10), 504. https://doi.org/10.3390/fermentation8100504
- Althor, G., Watson, J. E. M., & Fuller, R. A. (2016). Global mismatch between greenhouse gas emissions and the burden of climate change. *Scientific Reports*, 6(1), 20281. https://doi.org/10.1038/srep20281
- Badarina, I., Evvyernie, D., Toharmat, T., Herliyana, E. N., & Darusman, L. K. (2013).
 Nutritive Value of Coffee Husk Fermented with Pleurotus ostreatus as Ruminant
 Feed. *Media Peternakan*, 36(1), 58–63.
 https://doi.org/10.5398/medpet.2013.36.1.58
- Bampidis, V. A., & Robinson, P. H. (2006). Citrus by-products as ruminant feeds: A review. Animal Feed Science and Technology, 128(3–4), 175–217. https://doi.org/10.1016/j.anifeedsci.2005.12.002
- Batbekh, B., Ahmed, E., Hanada, M., Fukuma, N., & Nishida, T. (2023). Assessment of the Impact of Coffee Waste as an Alternative Feed Supplementation on Rumen

Fermentation and Methane Emissions in an In Vitro Study. *Fermentation*, 9(9), 858. https://doi.org/10.3390/fermentation9090858

- BAYARU, E., KANDA, S., KAMADA, T., ITABASHI, H., ANDOH, S., NISHIDA, T., ISHIDA, M., ITOH, T., NAGARA, K., & ISOBE, Y. (2001). Effect of Fumaric Acid on Methane Production, Rumen Fermentation and Digestibility of Cattle Fed Roughage Alone. *Nihon Chikusan Gakkaiho*, 72(2), 139–146. https://doi.org/10.2508/chikusan.72.139
- Beauchemin, K. A., Ungerfeld, E. M., Abdalla, A. L., Alvarez, C., Arndt, C., Becquet, P.,
 Benchaar, C., Berndt, A., Mauricio, R. M., McAllister, T. A., Oyhantçabal, W.,
 Salami, S. A., Shalloo, L., Sun, Y., Tricarico, J., Uwizeye, A., De Camillis, C.,
 Bernoux, M., Robinson, T., & Kebreab, E. (2022). Invited review: Current enteric
 methane mitigation options. *Journal of Dairy Science*, *105*(12), 9297–9326.
 https://doi.org/10.3168/jds.2022-22091
- Beauchemin, K. A., Ungerfeld, E. M., Eckard, R. J., & Wang, M. (2020). Review: Fifty years of research on rumen methanogenesis: lessons learned and future challenges for mitigation. *Animal*, 14, s2–s16. https://doi.org/10.1017/S1751731119003100
- Campos-Vega, R., Loarca-Piña, G., Vergara-Castañeda, H. A., & Oomah, B. D. (2015).
 Spent coffee grounds: A review on current research and future prospects. *Trends in Food Science & Technology*, 45(1), 24–36.
 https://doi.org/10.1016/j.tifs.2015.04.012
- Carro, M. D., & Ranilla, M. J. (2003). Effect of the addition of malate on *in vitro* rumen fermentation of cereal grains. *British Journal of Nutrition*, 89(2), 181–188. https://doi.org/10.1079/BJN2002759

- Carta, S., Tsiplakou, E., Nicolussi, P., Pulina, G., & Nudda, A. (2022). Effects of spent coffee grounds on production traits, haematological parameters, and antioxidant activity of blood and milk in dairy goats. *Animal*, 16(4), 100501. https://doi.org/10.1016/j.animal.2022.100501
- de Otálora, X. D., Ruiz, R., Goiri, I., Rey, J., Atxaerandio, R., San Martin, D., Orive, M., Iñarra, B., Zufia, J., Urkiza, J., & García-Rodríguez, A. (2020). valorisation of spent coffee grounds as functional feed ingredient improves productive performance of Latxa dairy ewes. *Animal Feed Science and Technology*, 264, 114461. https://doi.org/10.1016/j.anifeedsci.2020.114461

FAO publications catalogue 2023. (2023). FAO. https://doi.org/10.4060/cc7285en

- Gheller, L. S., Ghizzi, L. G., Marques, J. A., Takiya, C. S., Grigoletto, N. T. S., Dias, M. S. S., Silva, T. B. P., Nunes, A. T., da Silva, G. G., Fernandes, L. G. X., Rennó, L. N., & Rennó, F. P. (2020). Effects of organic acid-based products added to total mixed ration on performance and ruminal fermentation of dairy cows. *Animal Feed Science and Technology*, 261, 114406. https://doi.org/10.1016/j.anifeedsci.2020.114406
- Guyomard, H., Bouamra-Mechemache, Z., Chatellier, V., Delaby, L., Détang-Dessendre, C., Peyraud, J.-L., & Réquillart, V. (2021). Review: Why and how to regulate animal production and consumption: The case of the European Union. *Animal*, 15, 100283. https://doi.org/10.1016/j.animal.2021.100283
- Honan, M., Feng, X., Tricarico, J. M., & Kebreab, E. (2021). Feed additives as a strategic approach to reduce enteric methane production in cattle: modes of action, effectiveness and safety. *Animal Production Science*, 62(14), 1303–1317. https://doi.org/10.1071/AN20295

- Hopkins, F. M., Ehleringer, J. R., Bush, S. E., Duren, R. M., Miller, C. E., Lai, C., Hsu,
 Y., Carranza, V., & Randerson, J. T. (2016). Mitigation of methane emissions in cities: How new measurements and partnerships can contribute to emissions reduction strategies. *Earth's Future*, 4(9), 408–425. https://doi.org/10.1002/2016EF000381
- Intergovernmental Panel on Climate Change (IPCC). (2023). *Climate Change 2022 Impacts, Adaptation and Vulnerability*. Cambridge University Press. https://doi.org/10.1017/9781009325844
- Islam, M., & Lee, S.-S. (2019). Advanced estimation and mitigation strategies: a cumulative approach to enteric methane abatement from ruminants. *Journal of Animal Science and Technology*, 61(3), 122–137. https://doi.org/10.5187/jast.2019.61.3.122
- Kang, L.-L., Zeng, Y.-N., Wang, Y.-T., Li, J.-G., Wang, F.-P., Wang, Y.-J., Yu, Q., Wang, X.-M., Ji, R., Gao, D., & Fang, Z. (2022). Removal of pollutants from wastewater using coffee waste as adsorbent: A review. *Journal of Water Process Engineering*, 49, 103178. https://doi.org/10.1016/j.jwpe.2022.103178
- Kobayashi, Y., Oh, S., Myint, H., & Koike, S. (2016). Use of Asian selected agricultural byproducts to modulate rumen microbes and fermentation. *Journal of Animal Science and Biotechnology*, 7(1), 70. https://doi.org/10.1186/s40104-016-0126-4
- Ku-Vera, J. C., Castelán-Ortega, O. A., Galindo-Maldonado, F. A., Arango, J., Chirinda, N., Jiménez-Ocampo, R., Valencia-Salazar, S. S., Flores-Santiago, E. J., Montoya-Flores, M. D., Molina-Botero, I. C., Piñeiro-Vázquez, A. T., Arceo-Castillo, J. I., Aguilar-Pérez, C. F., Ramírez-Avilés, L., & Solorio-Sánchez, F. J. (2020). Review:

Strategies for enteric methane mitigation in cattle fed tropical forages. *Animal*, *14*, s453–s463. https://doi.org/10.1017/S1751731120001780

- *OECD-FAO* Agricultural Outlook 2023-2032. (2023). OECD. https://doi.org/10.1787/08801ab7-en
- Oryza.S, M., Wongtangtintharn, S., Tengjaroenkul, B., Cherdthong, A., Tanpong, S., Bunchalee, P., Pootthachaya, P., Reungsang, A., & Polyorach, S. (2021). Physico-Chemical Characteristics and Amino Acid Content Evaluation of Citric Acid by-Product Produced by Microbial Fermentation as a Potential Use in Animal Feed. *Fermentation*, 7(3), 149. https://doi.org/10.3390/fermentation7030149
- Oryza.S, M., Wongtangtintharn, S., Tengjaroenkul, B., Cherdthong, A., Tanpong, S., Pootthachaya, P., Boonkum, W., & Pintaphrom, N. (2021). Investigation of Citric Acid By-Products from Rice Produced by Microbial Fermentation on Growth Performance and Villi Histology of Thai Broiler Chicken (KKU 1). *Veterinary Sciences*, 8(11), 284. https://doi.org/10.3390/vetsci8110284
- Palangi, V., & Macit, M. (2021). Indictable Mitigation of Methane Emission Using Some Organic Acids as Additives Towards a Cleaner Ecosystem. *Waste and Biomass Valorization*, 12(9), 4825–4834. https://doi.org/10.1007/s12649-021-01347-8
- Pickering, N. K., Oddy, V. H., Basarab, J., Cammack, K., Hayes, B., Hegarty, R. S., Lassen, J., McEwan, J. C., Miller, S., Pinares-Patiño, C. S., & de Haas, Y. (2015).
 Animal board invited review: genetic possibilities to reduce enteric methane emissions from ruminants. *Animal*, 9(9), 1431–1440. https://doi.org/10.1017/S1751731115000968

- Ponnampalam, E. N., & Holman, B. W. B. (2023). Sustainability II: Sustainable animal production and meat processing. In *Lawrie's Meat Science* (pp. 727–798). Elsevier. https://doi.org/10.1016/B978-0-323-85408-5.00001-7
- Reality Check: Lessons from 25 Policies Advancing a Low-Carbon Future. (2023). Washington, DC: World Bank. https://doi.org/10.1596/40262
- Sandström, V., Chrysafi, A., Lamminen, M., Troell, M., Jalava, M., Piipponen, J., Siebert, S., van Hal, O., Virkki, V., & Kummu, M. (2022). Food system by-products upcycled in livestock and aquaculture feeds can increase global food supply. *Nature Food*, 3(9), 729–740. https://doi.org/10.1038/s43016-022-00589-6
- Sekaran, U., Lai, L., Ussiri, D. A. N., Kumar, S., & Clay, S. (2021). Role of integrated crop-livestock systems in improving agriculture production and addressing food security – A review. *Journal of Agriculture and Food Research*, 5, 100190. https://doi.org/10.1016/j.jafr.2021.100190
- Seo, J., Jung, J. K., & Seo, S. (2015). Evaluation of nutritional and economic feed values of spent coffee grounds and *Artemisia princeps* residues as a ruminant feed using *in vitro* ruminal fermentation. *PeerJ*, *3*, e1343. https://doi.org/10.7717/peerj.1343
- Thornton, P. K. (2010). Livestock production: recent trends, future prospects. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 365(1554), 2853–2867. https://doi.org/10.1098/rstb.2010.0134
- UN Climate Change. (2021). The Glasgow Climate Pact Key Outcomes from COP26.
- Wong, P. K., Ghadikolaei, M. A., Chen, S. H., Fadairo, A. A., Ng, K. W., Lee, S. M. Y., Xu, J. C., Lian, Z. D., Li, S., Wong, H. C., Zhao, J., Ning, Z., & Gali, N. K. (2022).
 Physicochemical and cell toxicity properties of particulate matter (PM) from a diesel vehicle fueled with diesel, spent coffee ground biodiesel, and ethanol.

Science of The Total Environment, 824, 153873. https://doi.org/10.1016/j.scitotenv.2022.153873

Xu, C. C., Cai, Y., Zhang, J. G., & Ogawa, M. (2007). Fermentation quality and nutritive value of a total mixed ration silage containing coffee grounds at ten or twenty percent of dry matter1. *Journal of Animal Science*, 85(4), 1024–1029. https://doi.org/10.2527/jas.2005-628 Chapter I

Assessment of the Impact of Coffee Waste as an Alternative Feed Supplementation on Rumen Fermentation and Methane Emissions in an *In Vitro* Study

Abstract

Spent coffee waste (SCW), a common by-product of the coffee industry, has shown potential as a dietary source for ruminants. This study aimed to determine the optimal inclusion dosage and application method of SCW in ruminant feed, investigating its impact on the rumen ecosystem and gas emission production. The *in vitro* study consisted of three trials with distinct experimental designs. In the first trial and experimental design (TRIAL. 1), a control diet with 500 mg of fresh matter basal diet (60% hay/40% concentrate) was used, and SCW was used as a feed additive at 1%, 10%, and 20% of the substrate. The second experimental design involved the same control diet, with SCW replacing either some of the hay (TRIAL. 2) or some of the concentrate mixture (TRIAL. 3) at four different dosages (30:70, 50:50, 70:30, and 100). When SCW was included as a feed additive, an increase in the production of volatile fatty acids (VFA) and gas was observed, but no suppressive effects on CH₄ production were noted. Conversely, when SCW replaced hav or concentrate, there were significant reductions in CH₄ production with increasing SCW inclusion levels. However, these reductions in CH₄ production were associated with negative effects on nutrient digestibility and total VFA production. These findings suggest that SCW could serve as a potential prebiotic feed additive. Moreover, the use of SCW as a substitute for silage at dosages of 70:30 and 50:50 appears feasible in animal feed (hay and concentrate).

I.1. Introduction

The world population is expected to reach approximately 9.7 billion people by 2050 and 10.4 billion people by 2100 (United Nations, n.d.). This growing population needs to be fed, so the prevalence of animal products, such as meat and milk, in human diets needs to be considered (Billen et al., 2015). Ruminants play an important role in animal production and contribute significantly to the overall quantity of animal products on the market. In the future, there will be an increased demand for ruminants in order to meet the food requirements of the growing global population. However, ruminants are also the most significant contributors to greenhouse gas (GHG) emissions, particularly CH₄. When their numbers increase, this has a significant effect on global warming (JIRCAS, 2010). In addition, the production of CH₄ during the fermentation process in the rumen is correlated with the loss of energy from the consumed feed. Moreover, CH₄ is 25 times more potent than CO₂ in terms of trapping heat from the sun (Hook et al., 2010). Therefore, reducing CH₄ emissions from ruminants would significantly decrease the associated environmental impacts, as long as energy utilization efficiency is not affected (Hristov et al., 2015). Thus, there is a need to find strategies that improve feed efficiency, balance the supply of nutrients to meet animal requirements, reduce environmental impacts and achieve economic benefits (JIRCAS, 2010). So, it is important for animal nutrition researchers to focus on finding alternative options to replace conventional resources and feed additives (Kobayashi et al., 2010).

Accordingly, using by-products from human food as feed for livestock is considered a possible solution. Coffee is one of the most popular beverages in the world and several by-products are generated throughout its processing stages. One of the major by-products of coffee is SCW, which contains large amounts of organic compounds, particularly lipids, polyphenols and polysaccharides (Campos-Vega et al., 2015). Due to the presence of these bioactive compounds, SCW is used in several industries, including biodiesel, cosmetics, construction

and animal feed (Carta et al., 2022; Kang et al., 2022; Wong et al., 2022; Xu et al., 2007; Yun et al., 2020). Moreover, the utilization of SCW as an alternative feed source for ruminants could help to mitigate CH₄ emissions, reduce waste and improve the environmental sustainability of livestock production; however, the dosages, processing methods and effects on animal health and performance need to be carefully considered.

Previously, researchers (Badarina et al., 2013; Campos-Vega et al., 2015; Seo et al., 2015; Xu et al., 2007) have focused on the potential of raw or ensiled SCW as a feed source for ruminants and have observed that SCW has some disadvantages, depending on the dosage. High levels of certain compounds, such as polyphenols, can hinder nutrient digestibility and affect animal health and production. Additionally, the use of SCW as a feed source for ruminants depends on a number of factors, such as cost, availability, processing and compatibility with other dietary components. Furthermore, appropriate processing methods (such as ensiling) need to be determined to mitigate any negative effects of compounds on animal performance and nutrient utilization. Large quantities of high-moisture by-products are produced in many countries, including Japan; therefore, there is a need to develop technologies to design superior animal feed using SCW and enable the long-term storage of the resulting silage (Cao et al., 2011). In Japan, there is an increasing interest in making silage by mixing wet and dry by-products, which offers a number of advantages, such as reduced risk of effluent production, stabilized rumen function and extended storage periods (Cao et al., 2011). The addition of lactic acid and soybean curd to silage when ensiling it with fresh grass or certain vegetable residues can improve fermentation quality. Moreover, when mixed with silage, these additives can also increase dry matter digestibility and reduce ruminal CH₄ production (Cao et al., 2011; Wang et al., 2022). Therefore, the objective of this *in vitro* study was to assess the impact of using raw or ensiled SCW as a feed additive or a partial replacement for the basal components (hay or concentrate) in ruminant diets on rumen fermentation profiles and CH4

production. Moreover, it is also important to establish the optimal level of SCW in animal diets (Seo et al., 2015; Xu et al., 2007). However, there are still limitations to the potential use of SCW as a feed additive or replacement for conventional feed and the exact optimal dosages and methods remain unclear.

I.2. Materials and Method

I.2.1. Basal diet and SCW

The basal diet consisted of ground Kleingrass (*Panicum coloratum*) hay with a particle size of 1mm and a concentrate mixture. The SCW, both raw and ensiled, was provided in powder form by Sanyu Group Co., Ltd., Kanagawa, Japan. The chemical compositions of the SCW and basal diet components are detailed in Table 1.

(Dry Matter	1/1 •	C ((((((((((Coffee	Coffee
g/kg)	Klein grass	Concentrate	(Raw)	(Silage)
Dry Matter				
(g/kg in fresh	908.6	884.4	961.3	956.6
matter)				
Organic Matter	909.1	943.6	978.2	961.8
Crude Protein	140.8	180.0	126.0	154.9
Ether Extract	20.6	35.0	140.0	146.3
NDF ¹	696.2	524.4	708.0	792.9
ADF ²	366.7	97.0	442.4	422.8
ADL ³	89.6	20.0	232.2	234.0

Table 1. Chemical composition of the feed used in this study

 1 NDF = neutral detergent fiber.

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 2 ADF = acid detergent fiber.

 3 ADL = acid detergent lignin.

I.2.2. Preparation of the silage

The silage preparation was conducted at Sanyu Group Co., Ltd., Kanagawa, Japan. SCW was obtained from Starbucks coffee stores across Japan. After being stored at the Customer Futures Distribution Center by Starbucks' chilled logistics, samples were collected at the factory by Sanyu Group logistics. After being drained at the stores, the SCW was packed in plastic bags, sprayed with vinegar spray and sealed for storage. The collected substrates were mixed with dried bean curd, bran, soy sauce dregs, vinegar and lactic acid bacteria. Then, the mixture was put in the polyethylene bags and placed into a stainless steel container for incubation. The entire ensiling process lasted for 14 days and was performed in the Sanyu Group factory.

I.2.3. Collection of Rumen fluid

In this experiment, the used animals were kept and cared for by the Field Science Center, Obihiro University of Agriculture and Veterinary Medicine, Japan. The animal management and sampling procedures were approved by the Obihiro University of Agriculture and Veterinary Medicine, Animal Care and Use Committee (approval number, 20-119).

In this study, two rumen-fistulated, non-lactating Holstein cows, which were about 9 years old, were used as rumen fluid donor animals. The cows were fed at maintenance level on a diet of orchard grass hay (organic matter, 980 g/kg; crude protein, 132 g/kg; neutral detergent fiber, 701 g/kg; acid detergent fiber, 354 g/kg; acid detergent lignin, 40 g/kg; dry matter base), with access to free drinking water and mineral blocks. Approximately 650 milliliters of rumen fluid were collected from each cow using a vacuum tube, strained through four layers of surgical gauze. Subsequently, approximately 1.3 liters of this strained fluid were placed into a pre-warmed Thermos flask with hot water. The collected rumen fluid was immediately transferred to the laboratory.

I.2.4. Experimental Design and Incubation Procedure

In the present study, 500 mg of the substrate was added to pre-weighed and nylon bags that have a fixed size and a pore size of $53 \pm 10 \,\mu$ m (BG1020, Sanshin Industrial Co., Ltd., Kanagawa, Japan), which were sealed using a heat-sealer before being placed into fermentation bottles.

This study was conducted using three experimental designs. The first experimental design (TRIAL. 1) was performed using a control diet of 500 mg of fresh matter basal diet (60% hay/40% concentrate), with SCW (both raw and ensiled) being used as a feed additive at 1%, 10%, and 20% of the substrate. In TRIAL. 1, the raw and ensiled SCW were added directly to the bottle and the experiments were conducted separately. The second experimental design was conducted using the same control diet as TRIAL. 1 but the SCW was added to the feed as a substrate in a nylon bag. The TRIAL. 2 and TRIAL. 3 were carried out on different days to assess the effects of replacing a portion of the hay or concentrate with SCW. TRIAL. 2 focused on replacing part of the hay with SCW, while TRIAL. 3 examined the replacement of a proportion of the concentrate with SCW. In the TRIAL. 2, four different dosages of SCW (raw and ensiled) were added to the basal diet to replace the hay 70:30 (42% hay/18% SCW/40% concentrate); 50:50 (30% hay/30% SCW/40% concentrate); 30:70 (18% hay/42% SCW/40% concentrate); and 100 (60% SCW/40% concentrate). In the TRIAL. 3, another four dosages of SCW (raw and ensiled) were used to replace a proportion of the concentrate 70:30 (60% hay/28% concentrate/12% SCW); 50:50 (60% hay/20% concentrate/20% SCW); 30:70 (60% hay/12% concentrate/28% SCW); 100 (60% hay/40% SCW). In TRIAL. 1, each group had four replicates and the experiment was repeated on four separate days. In TRIAL. 2 and TRIAL. 3, each group had three replicates and the experiments were repeated on three different days. In all of the trials, each run included two bottles of blank.

Via continuous CO₂ flushing, 40 mL of artificial saliva (McDougall, 1948) and 20 mL of rumen fluid were added to each fermentation bottle. The bottles were then reinjected with CO₂

before being sealed with rubber and aluminum caps (Maruemu Co., Ltd., Osaka, Japan). The incubation procedure was as described by (Ahmed et al., 2022).

I.2.5. In Vitro Incubation Procedure

After 24 h of incubation, total gas production was measured using a gas-tight syringe and headspace gas was collected from each bottle and stored in a vacuum tube (BD Vacutainer, Becton Drive, NJ, USA). Then, the gas composition was analyzed via gas chromatography (GC-8A, Shimadzu Corp., Kyoto, Japan), as described previously by Ahmed et al. (2022). Next, the bottles were opened, the pH was measured immediately using a pH meter (LAQUA F-72, HORIBA Scientific, Kyoto, Japan) and 1 mL of the culture medium was collected in an Eppendorf tube (Eppendorf AG, Hamburg, Germany) and centrifuged at 16,000×g at 4°C for 5 min. Following the centrifugation, the supernatant was gathered for further VFA analysis, which was measured via high-performance liquid chromatography (Shimadzu LC-20 HPLC, Shimadzu Corp., Kyoto, Japan). To determine the *in vitro* dry matter digestibility (IVDMD), the nylon bags containing the substrate were rinsed with tap water until the effluent became clear. They were then dried at 60°C for 48 h to enable us to measure the IVDMD, which was calculated as the percentage of DM that disappeared from the initial DM weight that was input into the bags.

I.2.6. Chemical analysis

The chemical composition analyses of the SCW, hay and concentrate mixture were performed according to the Association of Official Analytical Chemists procedures (AOAC, 1995). The DM content was determined by drying the matter in an oven at 135°C for 2 h (930.15). The OM and ash contents were measured by placing the samples in a muffle furnace at 500°C for 3 h (942.05). Nitrogen (N) content was measured according to the method of Kjeldahl (984.13) using an electrical heating digester (DK 20, VELP Scientifica, Usmate (MB), Monza, Italy) and an automatic distillation apparatus (UDK 129 VELP Scientifica, Usmate

(MB), Monza, Italy), CP was then estimated as N × 6.25. The NDF and ADF contents were estimated and expressed as the inclusive residual ash values using an ANKOM200 fiber analyzer (Ankom Technology, Methods 6 and 5, respectively; ANKOM Technology Corp., Macedon, NY, USA). The NDF content was measured using sodium sulfite without heat-stable α -amylase (FSS, ANKOM Technology).

I.2.7. Statistical analysis

All data were analyzed using SAS version 9.4 (SAS Institute Inc., Cary, NC, USA). For all experiments, the data were analyzed using PROC MIXED models, including the treatments as fixed effects, whereas the experimental runs were considered random effects. The values are presented as the means with the pooled standard errors of the means. Any differences in means between the experimental groups were estimated using Tukey's test. The statistical significance difference was declared at p < 0.05.

I.3 Results

I.3.1 TRIAL. 1

The inclusion of both raw and ensiled SCW as an additive at 1%, 10% and 20% levels resulted in increased total gas production (8.7%-11.8%) compared to the control diet (Tables 2 and 4). Similarly, CH₄ and CO₂ production per digestible DM also increased, except for CH₄ production per digestible DM at the 1% addition level of raw SCW by 4% (Table 2) and total gas production at the 1% addition level of ensiled SCW, which decreased by 3.5% (Table 4). The IVDMD was significantly lower with the addition of raw SCW at the 10% and 20% levels compared to the control diet (3.4%-12.06%; Table 3), while ensiled SCW did not show any significant effects on digestibility (Table 5). The results regarding rumen fermentation showed mostly increased parameters when both raw and ensiled SCW were used as an additive (Tables 3 and 5). The total VFA production was significantly increased in the raw SCW groups and there was also an observable increase in the ensiled SCW groups (Tables 3 and 5). Additionally, the raw SCW groups showed increases in propionate production, while the ensiled SCW groups demonstrated increased butyrate production (Tables 3 and 5). Notably, none of the groups showed any significant effects on pH or the acetate-to-propionate (A:P) ratio (Tables 3 and 5). However, more significant effects on rumen parameters were observed when raw SCW was used as a feed additive.

I.3.2 TRIAL. 2 and TRIAL. 3

The inclusion of both the raw and ensiled SCW resulted in decreases in CH_4 production and total gas production compared to the control group (Tables 6 and 8). When hay was replaced with SCW, there were significant reductions in total gas production (5.4%-27.2%) and CO_2 production (5.5%-26.7%) in both the raw and ensiled SCW groups (Table 8). However, the

reductions in CH₄ production (ml/day) were more pronounced when the concentrate was replaced by either raw or ensiled SCW, with reductions ranging from 6% to 59%, compared to the grass diet replacement groups, which showed reductions of 1.9%-38% (Tables 6 and 8). The IVDMD was also significantly reduced by the inclusion of SCW, with the hay replacement groups showing reductions of 3%-27% and the concentrate replacement groups showing reductions of 3.8%-33.5% (Tables 7 and 9). However, the lower dosage of SCW as a concentrate replacement did not show significant reductions in IVDMD in any groups (Table 6). The VFA production was significantly reduced in all tested groups when raw SCW was used to replace both hay and concentrate (Tables 7 and 9), but there were no significant effects on VFA production in some of the ensiled SCW groups (Tables 7 and 9).

Additionally, butyrate production was unchanged when SCW was used to replace hay, although the percentage of production increased (Table 9). In contrast, when SCW was used to replace the concentrate, all groups showed significant reductions in butyrate production (Table 7). The A:P increased in all groups when raw SCW was used to replace hay, except for the 70:30 raw SCW group (Table 9), while none of the ensiled SCW groups showed any significant effects. In contrast, when the concentrate was replaced with SCW, most groups showed significant increases in the A:P ratio, except for the 70:30 ensiled group (Table 7).

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Item	Control	1%	10%	20%	SEM	Р-
item	Control	1 /0	10 /0	20 /0	SEIVI	Value
рН	6.69	6.69	6.68	6.69	0.006	0.090
IVDMD%	48.83 ^{ab}	50.29ª	47.16 ^{a-c}	42.94°	1.21	0.008
Acetate (<i>mM</i>)	164.62	166.82	166.77	168.19	3.51	0.080
Propionate (<i>mM</i>)	53.88 ^b	55.37 ^{ab}	56.01 ^{ab}	56.37ª	1.54	0.020
Butyrate (<i>mM</i>)	20.03	20.52	20.35	20.57	0.58	0.300
TVFA ¹ (<i>mM</i>)	238.55 ^b	242.71 ^{ab}	243.13 ^{ab}	245.14ª	4.45	0.040
Acetate (%)	68.98	68.80	68.63	68.64	0.67	0.210
Propionate (%)	22.57	22.74	22.98	22.95	0.46	0.110
Butyrate (%)	8.44	8.46	8.39	8.41	0.21	0.820
A:P ²	3.13	3.11	3.08	3.06	0.08	0.200

Table 2: Effect of raw SCW as a feed additive on rumen fermentation characteristics

¹TVFA: Total Volatile Fatty Acids. ²A/P: Acetate/Propionate. SEM: Standard Error of the Mean. ^{a, b, c} in the same row with different superscript means differ significantly (p < 0.05).

Item	Control	1%	10%	20%	SEM	Р-
Item	Control	170	1070	2070		Value
Total gas (ml/day)	52.91°	56.00 ^{ab}	57.50 ^{ab}	58.66ª	1.26	<.0001
Total gas /DDM ¹ (ml/g)	108.05 ^b	111.51 ^b	124.97 ^b	153.80ª	6.08	0.0001
CH4(%)	5.16	4.81	4.79	4.94	0.07	0.0600
CO ₂ (%)	94.84	95.19	95.21	95.06	0.07	0.0600
CH ₄ /DDM (ml/g)	5.54 ^{bc}	5.32 ^{bc}	5.92 ^b	7.46 ^a	0.26	0.0004
CH4 (ml/day)	2.72	2.69	2.75	2.91	0.07	0.1500
CO ₂ /DDM (ml/g)	102.51 ^{bc}	106.19 ^{bc}	119.04 ^b	146.34 ª	5.83	0.0001
CO2 (ml/day)	50.19°	53.30 ^{ab}	54.75 ^{ab}	55.75ª	1.21	<.0001

Table 3: Effect of raw SCW as a feed additive on gas profile

¹ DDM, Digestible Dry Matter. SEM: Standard Error of the Mean. ^{a, b, c} in the same row with different superscript means differ significantly (p < 0.05).

Item	Control	1%	10%	20%	SEM	Р-
						Value
Total gas (ml/day)	56.50 ^{bc}	54.50°	61.70 ^{ab}	63.20ª	1.14	0.003
Total gas /DDM ¹ (ml/g)	103.23 ^{ab}	97.30 ^b	112.12 ^{ab}	118.28ª	2.80	0.010
CH4(%)	3.95°	6.97 ^a	5.73 ^{a-c}	6.04 ^{ab}	0.35	0.005
CO ₂ (%)	96.04ª	93.02 ^{bc}	94.26 ^{ab}	93.95 ^{bc}	0.35	0.005
CH ₄ /DDM (ml/g)	4.09°	6.81 ^{ab}	6.42 ^{a-c}	7.15 ^a	0.40	0.011
CH4 (ml/day)	2.24 ^b	3.82 ^a	3.53 ^{ab}	3.82 ^a	0.21	0.009
CO ₂ /DDM (ml/g)	99.14 ^{ab}	90.48 ^b	105.69 ^{ab}	111.13ª	2.65	0.010
CO ₂ (ml/day)	54.25 ^{a-c}	50.67°	58.21 ^{ab}	59.42ª	1.06	0.002

Table 4: Effect of silage SCW as a feed additive on gas profile

¹ DDM, Digestible Dry Matter. SEM: Standard Error of the Mean. ^{a, b, c} in the same row with different superscript means differ significantly (p < 0.05).

Item	Control	1%	10%	20%	SEM	Р-
						Value
рН	6.66	6.64	6.66	6.65	0.005	0.460
IVDMD%	54.72	56.01	55.34	53.81	0.84	0.850
Acetate (<i>mM</i>)	151.50	148.07	152.28	156.86	1.42	0.180
Propionate (<i>mM</i>)	66.04	64.72	63.67	67.68	1.28	0.750
Butyrate (<i>mM</i>)	25.23 ^{ab}	24.70 ^b	27.31ª	27.37ª	0.39	0.009
TVFA ¹ (<i>mM</i>)	242.77	237.51	243.28	251.92	2.72	0.320
Acetate (%)	62.41	62.34	62.65	62.27	0.17	0.890
Propionate (%)	27.19	27.25	26.07	26.84	0.31	0.550
Butyrate (%)	10.38	10.40	11.26	10.87	0.16	0.150
A:P ²	2.29	2.28	2.41	2.32	0.03	0.600

Table 5: Effect of silage SCW as a feed additive on rumen fermentation characteristics

¹TVFA: Total Volatile Fatty Acids. ²A/P: Acetate/Propionate. SEM: Standard Error of the Mean. ^{a, b} means in the same row with different superscript differ significantly (p < 0.05).

14	C		Rí	Raw			Sil	Silage			Ъ.
пеш	Control	30:70	50:50	70:30	100	30:70	50:50	70:30	100	SEM	Value
Total gas (ml/day)	48.22 ^a	45.67 ^{bc}	42.11 ^d	38.44 ^d	31.00€	47.56 ^b	45.72 ^{bc}	41.11 ^d	36.56 ^d	0.65	<.0001
Total gas /DDM ¹ (ml/g)	204.98 ^{ab}	201.71 ^{bc}	198.31 ^{bcd}	197.23 ^{bcde} 183.86 ^{befg}	183.86 ^{befg}	206.99ª	204.77 ^{ab}	194.16 ^{bcef}	180.05 ^g	1.54	0.004
CH4 (%)	4.78ª	4.40^{a}	4.19ª	3.93 ^b	3.07°	4.57 ^{ab}	3.85 ^b	4.08 ^b	3.60 ^b	0.09	<.0001
$CO_{2}(\%)$	95.22 ^a	95.60ª	95.80ª	96.07 ^b	96.93 ^b	95.43 ^b	96.15 ^{ab}	95.92 ^{ab}	96.40ª	0.09	<.0001
CH4/DDM (ml/g)	9.69 ^a	8.70 ^{ab}	8.20 ^{bc}	7.63 ^{bc}	5.72 ^d	9.33 ^{ab}	8.70 ^{abc}	7.94°	6.66 ^d	0.19	<.0001
CH4 (ml/day)	2.32 ^a	2.02 ^{ab}	$1.77^{ m bc}$	1.52°	0.95 ^d	2.18 ^{ab}	1.98^{b}	1.68°	1.32 ^d	0.05	<.0001
CO ₂ /DDM (ml/g)	#	187.8	187.02	186.31	181.79	194.38	192.74	186.19	178.09	1.42	0.15
CO2 (ml/day)	45.91 ^a	43.64 ^a	40.34 ^b	36.93°	30.05 ^d	45.37 ^{ab}	43.74 ^{ab}	39.43°	35.23 ^d	0.61	<.0001

Table 6: Effect of SCW as a replacing concentrate on gas profile

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7 1	C		R	Raw			Sil	Silage			
Item	Control	30:70	50:50	70:30	100	30:70	50:50	70:30	100	SEM	P-Value
ЬН	6.64 ^d	6.68°	6.71 ^{ab}	6.72 ^{ab}	6.73ª	6.66 ^{a-d}	6.66 ^{abc}	6.68^{ab}	6.70 ^a	0.01	<.0001
IVDMD%	53.53 ^a	51.19ª	47.24 ^b	43.17°	35.60 ^d	51.52 ^{ab}	49.81 ^b	46.19°	42.69 ^d	0.65	<.0001
Acetate (<i>mM</i>)	39.27 ^a	38.03 ^b	52.24°	49.58 ^d	45.36°	55.75 ^{ab}	54.28 ^{bc}	52.59°	50.32 ^d	0.94	<.0001
Propionate (<i>mM</i>)	14.24^{a}	13.18 ^b	36.95 ^b	35.65°	33.46 ^d	38.73 ^b	37.97 ^{bc}	37.24 ^{cd}	36.14 ^d	0.27	<.0001
Butyrate (<i>mM</i>)	3.74ª	3.35 ^b	12.20°	11.17 ^d	9.69∘	13.66 ^{ab}	13.14 ^{bc}	12.50°	11.66 ^d	0.18	<.0001
TVFA ¹ (<i>mM</i>)	57.24 ^a	54.56 ^b	3.09 ^{bc}	2.76°	2.21 ^d	3.38 ^b	3.16 ^{bc}	2.85 ^{cd}	2.53 ^d	1.13	<.0001
Acetate (%)	68.14°	69.22 ^d	70.24°	71.36 ^b	73.23 ^a	68.92°	69.46^{b}	70.30 ^b	71.27 ^a	0.41	<.0001
Propionate (%)	25.04 ^a	24.31 ^a	23.47 ^b	22.64°	21.38 ^d	24.67 ^{ab}	24.36 ^{abc}	23.85 ^{cd}	23.17 ^d	0.16	<.0001
Butyrate (%)	6.82 ^a	6.47 ^{ab}	6.29 ^{bc}	6.00 ^{cd}	5.40°	6.41 ^{ab}	6.19 ^{bc}	5.85 ^{cd}	5.56 ^d	0.36	<.0001
A:P ²	2.73°	2.86 ^d	3.00°	3.16^{b}	3.43^{a}	2.80^{cd}	2.86^{bc}	2.95 ^b	3.08 ^a	0.03	<.0001

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			R	Raw			Silage	ıge			
Item	Control	30:70	50:50	70:30	100	30:70	50:50	70:30	100	SEM	P-Value
Total gas (ml/day)	53.89ª	50.44 ^b	47.44°	46.06°	39.22 ^d	50.89 ^b	51 ^{bc}	49 ^{bc}	44.89 ^d	0.5	<.0001
Total gas /DDM ¹ (ml/g)	207.58 ^{ab}	211.89 ^{bc}	215.41 ^{bcd}	208.43 ^{bcde} 199.53 ^{befg}	199.53 ^{befg}	199.41 ^{abc}	199.96 ^{ab}	201.20 ^{bcef}	188.91 ^g	1.4	<.0001
CH4 (%)	4.83	4.61	4.75	4.72	4.11	4.73	4.99	4.57	4.28	0.1	0.3
$CO_2(\%)$	95.2	95.4	95.25	95.28	95.9	95.3	95	95.43	95.7	0.1	0.3
CH4/DDM (ml/g)	10	9.76	10.2	9.82	8.21	9.4	10.02	9.16	8.14	0.2	0.03
CH4 (ml/day)	2.60 ^a	2.32 ^a	2.25 ^a	2.17 ^a	1.61 ^b	$2.40^{\rm abc}$	2.55 ^{ab}	2.23 ^{abc}	1.92°	0.1	<.0001
CO ₂ /DDM (ml/g)	197.58 ^{ab}	202.13 ^{ab}	205.20 ^a	198.61 ^{ab}	191.32 ^b	190.00 ^{abc}	189.94 ^{abc}	192.04 ^{ab}	180.77°	1.3	<.0001
CO2 (ml/day)	51.29 ^a	48.12 ^b	45.20°	43.89°	37.61 ^d	48.49 ^b	48.45 ^{bc}	46.77 ^{bc}	42.96 ^d	0.5	<.0001

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ì	- - (R	Raw			Sil	Silage			
Item	Control	30:70	50:50	70:30	100	30:70	50:50	70:30	100	SEM	P-Value
рН	6.59 ^b	6.61 ^{ab}	6.61 ^{ab}	6.61 ^{ab}	6.63 ^a	6.60 ^b	6.58 ^b	6.60 ^b	6.59 ^b	0.004	0.0001
IVDMD%	58.14 ^a	52.60 ^b	48.27°	48.05°	42.30 ^d	56.41 ^{ab}	56.12 ^{abc}	53.19 ^{cd}	51.57 ^d	0.57	<.0001
Acetate (<i>mM</i>)	37.15 ^a	35.67 ^b	52.04°	51.36°	47.67 ^d	55.80 ^{abc}	55.99 ^{ab}	54.75 ^{bcd}	34.16 ^d	0.68	<.0001
Propionate (<i>mM</i>)	13.95 ^a	12.99 ^b	34.21°	33.79°	31.50 ^d	36.15 ^{abc}	36.21^{ab}	35.26 ^{bc}	12.84 ^d	0.24	<.0001
Butyrate (<i>mM</i>)	5.98 ^{abcde}	5.95 abcde	12.06°	11.63°	10.43^{d}	13.61 ^{ab}	13.62 ^{abc}	13.42 ^{abcd}	6.19ª	0.07	<.0001
TVFA ¹ (mM)	57.08ª	54.62 ^b	5.77 ^{de}	5.94 abcde	5.75°	6.04 ^{abcd}	6.17 ^{ab}	6.07 ^{abc}	53.07 ^d	0.86	<.0001
Acetate (%)	64.78 ^d	64.98 ^{bcd}	65.41 ^{abc}	65.46 ^{ab}	65.71 ^a	64.45 ^{ab}	64.33 ^{ab}	64.07 ^b	63.93 ^b	0.30	<.0001
Propionate (%)	24.36 ^a	23.73 ^{ab}	23.14 ^{bc}	22.61°	21.87 ^d	24.34ª	24.27 ª	24.47	24.2	0.11	<.0001
Butyrate (%)	10.87 ^d	11.29°	11.44 ^b	11.93 ^b	12.43 ^a	11.21 ^{bc}	11.40 ^{bc}	11.46^{ab}	11.88 ^a	0.32	<.0001
\mathbf{A} : \mathbf{P}^2	2.66^{d}	2.74 ^{cd}	2.83 ^{bc}	2.9 ^b	3.01 ^a	2.7	2.65	2.62	2.65	0.02	<.0001

Table 9: Effect of SCW as a replacing grass on rumen fermentation characteristics

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I.4 Discussion

I.4.1 Nutritive value

In the present study, the proximate analysis showed that SCW has a high nutritive value (NDF > 650g/kg; ADF > 350g/kg; CP > 120g/kg). Previously, coffee grounds have been shown to contain high protein, fat and fiber levels, meaning that they can be considered a feed source for ruminant (Campbell et al., 1976). Several studies have used coffee pulp and husk as feed for ruminants (Badarina et al., 2013; Maxiselly et al., 2022; Mazzafera et al., 2002; Murthy & Madhava Naidu, 2012). Additionally, some previous studies have reported that coffee residue contains high levels of organic compounds and is an appropriate substrate for fermentation processes (Badarina et al., 2013; Seo et al., 2015; Xu et al., 2007).

I.4.2 SCW a feed additive at the 1%, 10%, 20% levels of DM

In the present study, the addition of raw SCW resulted in increased gas production and improved rumen fermentation parameters. The addition of raw SCW also led to increases in total VFA production and propionate production, with no changes in acetate production. These findings could be attributed to the presence of polyphenols and fatty acids in SCW, which have been shown to exhibit anti-methanogenic effects in the rumen (Beauchemin & McGinn, 2006; Bodas et al., 2012; Cieslak et al., 2012). However, the decrease in IVDMD with higher dosages of raw SCW was consistent with previous studies, which have indicated that plant secondary metabolites, including phenolic compounds and fatty acids, can slow intake degradation while improving ruminant production. These products can reduce the nutritive value of SCW at increased dosages, but they can also exhibit other beneficial rumen modulation effects, such as reducing protein and starch degradation and inhibiting amino acid degradation, via selective actions on certain rumen microorganisms. Some *in vivo* studies have reported that these

compounds can improve live weight, milk production, and ovulation rate in ruminants (A. K. Patra et al., 2013; Patra & Saxena, 2010; Waghorn & McNabb, 2003).

The addition of ensiled SCW resulted in increased gas production without any significant effects on the IVDMD or rumen fermentation parameters. However, there were increases in butyrate production with higher dosages of ensiled SCW. This could be attributed to the presence of certain compounds in the silage, which can affect the diversity of ruminal bacteria and the ability of certain bacterial taxa to degrade lignocellulosic material, ultimately leading to increased butyrate concentrations (Otálora et al., 2020; Goiri et al., 2020). The results from both the raw and ensiled SCW groups were almost the same, but the addition of raw SCW had a stronger impact on rumen parameters. In addition, both raw and ensiled SCW can be considered good sources of energy and protein, as previously described by Senevirathne et al. (2012). Therefore, they could potentially be used as prebiotic feed additives to enhance the health status of animals.

I.4.3 SCW as a feed replacement

In the present study, significant reductions in production were observed in almost all groups in TRAIL. 3 (p < 0.05). Previous studies have reported that coffee grounds contain significant amounts of lipids, particularly palmitic acid (C16:0) and linoleic acid (C18:2), which can contribute to reductions in CH₄ production (Jenkins et al., 2014; Somnuk et al., 2017). Dietary lipids, especially medium-chain fatty acids (MCFAs) and long-chain unsaturated fatty acids (UFAs), have been shown to decrease CH₄ production in ruminants (Beauchemin & McGinn, 2006; Castagnino et al., 2015; Dong et al., 1997; Machmüller et al., 1998). The addition of 1% fat, the most common source of MCFAs, can reduce CH₄ production by 3.1% to 9.1% (Aemiro et al., 2016; Grainger et al., 2008). Similarly, McGinn et al. (2004) found that the addition of sunflower oil decreased CH₄ production by 22% per 5% of DM. The results of the present study confirmed that SCW, with its high fat content (up to 140 g fat/kg DM), increased dietary fat concentrations by 1.8% and suppressed CH₄ emissions by 12.9%.

In our findings, the use of SCW to replace hay or concentrate in the diet of ruminants resulted in significant decreases in IVDMD in almost all groups (p < 0.05). High lipid contents in ruminant diets have been shown to reduce DM, OM and fiber digestibility (Beauchemin & McGinn, 2006; Castagnino et al., 2015; McGinn et al., 2004; Patra et al., 2013). Previous studies have also reported that SCW contains a maximum of 14.7 wt% oil and high concentrations of tri- and monoglycerides (wt%), which can hinder feed particle adhesion and reduce nutrient availability for ruminal bacteria (Somnuk et al., 2017). Furthermore, the decreases in total gas and VFA production could be attributed to the interference of glycerol in SCW, which leads to slow feed particle adhesion and reduced degradability (Abo El-Nor et al., 2010; Castagnino et al., 2015).

Contrary to expectations, the addition of SCW resulted in reduced VFA production and increased pH compared to control, particularly in the concentrate replacement groups. This finding contradicted the results of some previous studies that showed little or no impact of SCW on ruminal pH or VFA production (Senevirathne et al., 2012; Seo et al., 2015). The reductions in VFA production and increases in pH could be attributed to certain polyphenolic compounds, such as tannins, lignans and caffeic acids, that are present in SCW. These compounds have been shown to reduce total VFA concentrations and alter ruminal microbial diversity (Badarina et al., 2013; Hassanat & Benchaar, 2013). These reductions in total VFA could also be related to the structures of tannin carbohydrate and protein compounds that cannot be degraded by rumen microbes or are toxic to ruminal microbes (Hassanat & Benchaar, 2013). The changes in the rumen microbial community caused by the SCW could also have led to altered rumen fermentation.

In previous studies, researchers have explored the use of coffee grounds as silage as a strategy to increase the nutritional value of animal diets and address environmental concerns (Badarina et al., 2013; Seo et al., 2015; Xu et al., 2007). The ensiling process involves fermenting and preserving the coffee grounds, potentially enhancing their digestibility, nutrient profile and storage time (Xu et al., 2007). However, in the present study, no significant effects of ensiled SCW were observed on rumen fermentation parameters. This suggested that ensiled SCW did not have any notable impacts on rumen microbial activity, gas production or volatile fatty acid production under the specific experimental conditions of this study.

Additionally, the use of SCW (raw or ensiled) as a replacement for hay or concentrate led to significant decreases in IVDMD and total VFA production, mostly at higher dosages. However, at lower dosages of ensiled SCW (specifically 70:30 and 50:50), there were no significant reductions in some ruminal fermentation parameters. Thus, these dosages could be feasible as a replacement for traditional animal feed.

I.5 Conclusions

The findings of this study indicated that SCW could serve as a feasible alternative to conventional ruminant feed ingredients due to its nutrient composition. The inclusion of SCW in ruminant diets has demonstrated promising results in terms of reducing CH₄ emissions, which is advantageous for minimizing the environmental impacts of ruminant agriculture. However, it is important to note that higher dosages of SCW may have adverse effects on animal production, although some lower dosages could be considered for use in animal feed. Therefore, further research is required to comprehensively evaluate the implications of using SCW in ruminant diets, as well as to determine the most suitable administration methods and optimal dosage levels. By conducting additional studies, we could gain deeper insights into the potential benefits and limitations of using SCW as a feed ingredient for ruminants. This knowledge could aid in the development of sustainable and efficient feeding strategies that could optimize animal health, productivity and environmental sustainability. It is through continuous investigation and scientific exploration that we can unlock the full potential of SCW as a valuable resource in ruminant nutrition and contribute to a more sustainable agricultural future.

I.6 References

- Abo El-Nor, S., AbuGhazaleh, A. A., Potu, R. B., Hastings, D., & Khattab, M. S. A. (2010). Effects of differing levels of glycerol on rumen fermentation and bacteria. *Animal Feed Science and Technology*, 162(3–4), 99–105. https://doi.org/10.1016/j.anifeedsci.2010.09.012
- Aemiro, A., Watanabe, S., Suzuki, K., Hanada, M., Umetsu, K., & Nishida, T. (2016).
 Effects of Euglena (Euglena gracilis) supplemented to diet (forage: concentrate ratios of 60:40) on the basic ruminal fermentation and methane emissions in in vitro condition. *Animal Feed Science and Technology*, 212, 129–135. https://doi.org/10.1016/j.anifeedsci.2015.12.010
- Ahmed, E., Batbekh, B., Fukuma, N., Hanada, M., & Nishida, T. (2022). Evaluation of
 Different Brown Seaweeds as Feed and Feed Additives Regarding Rumen
 Fermentation and Methane Mitigation. *Fermentation*, 8(10), 504.
 https://doi.org/10.3390/fermentation8100504

AOAC. (1995). Official Methods of Analysis of AOAC International: 16th Edition.

- Badarina, I., Evvyernie, D., Toharmat, T., Herliyana, E. N., & Darusman, L. K. (2013).
 Nutritive Value of Coffee Husk Fermented with Pleurotus ostreatus as Ruminant
 Feed. *Media Peternakan*, 36(1), 58–63.
 https://doi.org/10.5398/medpet.2013.36.1.58
- Beauchemin, K. A., & McGinn, S. M. (2006). Methane emissions from beef cattle: Effects of fumaric acid, essential oil, and canola oil1. *Journal of Animal Science*, 84(6), 1489–1496. https://doi.org/10.2527/2006.8461489x
- Billen, G., Lassaletta, L., & Garnier, J. (2015). A vast range of opportunities for feeding the world in 2050: trade-off between diet, N contamination and international trade.

Environmental Research Letters, 10(2), 025001. https://doi.org/10.1088/1748-9326/10/2/025001

- Bodas, R., Prieto, N., García-González, R., Andrés, S., Giráldez, F. J., & López, S. (2012). Manipulation of rumen fermentation and methane production with plant secondary metabolites. *Animal Feed Science and Technology*, 176(1–4), 78–93. https://doi.org/10.1016/j.anifeedsci.2012.07.010
- Campbell, T. W., Bartley, E. E., Bechtle, R. M., & Dayton, A. D. (1976). Coffee Grounds. I. Effects of Coffee Grounds on Ration Digestibility and Diuresis in Cattle, on In Vitro Rumen Fermentation, and on Rat Growth. *Journal of Dairy Science*, 59(8), 1452–1460. https://doi.org/10.3168/jds.S0022-0302(76)84384-6
- Campos-Vega, R., Loarca-Piña, G., Vergara-Castañeda, H. A., & Oomah, B. D. (2015).
 Spent coffee grounds: A review on current research and future prospects. *Trends* in Food Science & Technology, 45(1), 24–36. https://doi.org/10.1016/j.tifs.2015.04.012
- Cao, Y., Cai, Y., Takahashi, T., Yoshida, N., Tohno, M., Uegaki, R., Nonaka, K., & Terada, F. (2011). Effect of lactic acid bacteria inoculant and beet pulp addition on fermentation characteristics and in vitro ruminal digestion of vegetable residue silage. *Journal of Dairy Science*, 94(8), 3902–3912. https://doi.org/10.3168/jds.2010-3623
- Carta, S., Tsiplakou, E., Nicolussi, P., Pulina, G., & Nudda, A. (2022). Effects of spent coffee grounds on production traits, haematological parameters, and antioxidant activity of blood and milk in dairy goats. *Animal*, 16(4), 100501. https://doi.org/10.1016/j.animal.2022.100501

- Castagnino, P. S., Messana, J. D., Fiorentini, G., de Jesus, R. B., San Vito, E., Carvalho,
 I. P. C., & Berchielli, T. T. (2015). Glycerol combined with oils did not limit biohydrogenation of unsaturated fatty acid but reduced methane production in vitro. *Animal Feed Science and Technology*, 201, 14–24. https://doi.org/10.1016/j.anifeedsci.2014.12.004
- Cieslak, A., Zmora, P., Pers-Kamczyc, E., & Szumacher-Strabel, M. (2012). Effects of tannins source (Vaccinium vitis idaea L.) on rumen microbial fermentation in vivo. *Animal Feed Science and Technology*, 176(1–4), 102–106. https://doi.org/10.1016/j.anifeedsci.2012.07.012
- de Otálora, X. D., Ruiz, R., Goiri, I., Rey, J., Atxaerandio, R., San Martin, D., Orive, M., Iñarra, B., Zufia, J., Urkiza, J., & García-Rodríguez, A. (2020). valorisation of spent coffee grounds as functional feed ingredient improves productive performance of Latxa dairy ewes. *Animal Feed Science and Technology*, 264, 114461. https://doi.org/10.1016/j.anifeedsci.2020.114461
- Dong, Y., Bae, H. D., McAllister, T. A., Mathison, G. W., & Cheng, K.-J. (1997). Lipidinduced depression of methane production and digestibility in the artificial rumen system (RUSITEC). *Canadian Journal of Animal Science*, 77(2), 269–278. https://doi.org/10.4141/A96-078
- Goiri, I., Díaz de Otálora, X., Ruiz, R., Rey, J., Atxaerandio, R., Lavín, J. L., San Martin,
 D., Orive, M., Iñarra, B., Zufia, J., Urkiza, J., & García-Rodríguez, A. (2020).
 Spent Coffee Grounds Alter Bacterial Communities in Latxa Dairy Ewes. *Microorganisms*, 8(12), 1961. https://doi.org/10.3390/microorganisms8121961
- Grainger, C., Clarke, T., Beauchemin, K. A., McGinn, S. M., & Eckard, R. J. (2008). Supplementation with whole cottonseed reduces methane emissions and can

profitably increase milk production of dairy cows offered a forage and cereal grain diet. *Australian Journal of Experimental Agriculture*, 48(2), 73. https://doi.org/10.1071/EA07224

- Hassanat, F., & Benchaar, C. (2013). Assessment of the effect of condensed (acacia and quebracho) and hydrolysable (chestnut and valonea) tannins on rumen fermentation and methane production *in vitro*. *Journal of the Science of Food and Agriculture*, 93(2), 332–339. https://doi.org/10.1002/jsfa.5763
- Hook, S. E., Wright, A.-D. G., & McBride, B. W. (2010). Methanogens: Methane Producers of the Rumen and Mitigation Strategies. *Archaea*, 2010, 1–11. https://doi.org/10.1155/2010/945785
- Hristov, A. N., Oh, J., Giallongo, F., Frederick, T., Weeks, H., Zimmerman, P. R., Harper, M. T., Hristova, R. A., Zimmerman, R. S., & Branco, A. F. (2015). The Use of an Automated System (GreenFeed) to Monitor Enteric Methane and Carbon Dioxide Emissions from Ruminant Animals. *Journal of Visualized Experiments*, 103. https://doi.org/10.3791/52904
- Jenkins, R. W., Stageman, N. E., Fortune, C. M., & Chuck, C. J. (2014). Effect of the Type of Bean, Processing, and Geographical Location on the Biodiesel Produced from Waste Coffee Grounds. *Energy & Fuels*, 28(2), 1166–1174. https://doi.org/10.1021/ef4022976
- JIRCAS. (2010). Nutrient Requirements of Beef Cattle in Indochinese Peninsula The Working Committee of Thai Feeding Standard for Ruminant (WTSR) Department of Livestock Development, Ministry of Agriculture and Cooperatives, Thailand Nutrient Requirements of Beef Cattle in Indochinese Peninsula.

- Kang, L.-L., Zeng, Y.-N., Wang, Y.-T., Li, J.-G., Wang, F.-P., Wang, Y.-J., Yu, Q., Wang, X.-M., Ji, R., Gao, D., & Fang, Z. (2022). Removal of pollutants from wastewater using coffee waste as adsorbent: A review. *Journal of Water Process Engineering*, 49, 103178. https://doi.org/10.1016/j.jwpe.2022.103178
- Kobayashi, Y. (2010). Abatement of Methane Production from Ruminants: Trends in the Manipulation of Rumen Fermentation. Asian-Australasian Journal of Animal Sciences, 23(3), 410–416. https://doi.org/10.5713/ajas.2010.r.01
- Machmüller, A., Ossowski, D. A., Wanner, M., & Kreuzer, M. (1998). Potential of various fatty feeds to reduce methane release from rumen fermentation in vitro (Rusitec). *Animal Feed Science and Technology*, 71(1–2), 117–130. https://doi.org/10.1016/S0377-8401(97)00126-0
- Maxiselly, Y., Chiarawipa, R., Somnuk, K., Hamchara, P., Cherdthong, A., Suntara, C., Prachumchai, R., & Chanjula, P. (2022). Digestibility, Blood Parameters, Rumen Fermentation, Hematology, and Nitrogen Balance of Goats after Receiving Supplemental Coffee Cherry Pulp as a Source of Phytochemical Nutrients. *Veterinary Sciences*, 9(10), 532. https://doi.org/10.3390/vetsci9100532
- Mazzafera, P. (2002). Degradation of caffeine by microorganisms and potential use of decaffeinated coffee husk and pulp in animal feeding. *Scientia Agricola*, 59(4), 815–821. https://doi.org/10.1590/S0103-90162002000400030
- McDougall, E. I. (1948). Studies on ruminant saliva. 1. The composition and output of sheep's saliva. *The Biochemical Journal*, *43*(1), 99–109.
- McGinn, S. M., Beauchemin, K. A., Coates, T., & Colombatto, D. (2004). Methane emissions from beef cattle: Effects of monensin, sunflower oil, enzymes, yeast,

and fumaric acid1. *Journal of Animal Science*, 82(11), 3346–3356. https://doi.org/10.2527/2004.82113346x

- Murthy, P. S., & Madhava Naidu, M. (2012). Sustainable management of coffee industry by-products and value addition—A review. *Resources, Conservation and Recycling*, 66, 45–58. https://doi.org/10.1016/j.resconrec.2012.06.005
- Patra, A. K. (2013). The effect of dietary fats on methane emissions, and its other effects on digestibility, rumen fermentation and lactation performance in cattle: A metaanalysis. *Livestock Science*, 155(2–3), 244–254. https://doi.org/10.1016/j.livsci.2013.05.023
- Patra, A. K., & Saxena, J. (2010). A new perspective on the use of plant secondary metabolites to inhibit methanogenesis in the rumen. *Phytochemistry*, 71(11–12), 1198–1222. https://doi.org/10.1016/j.phytochem.2010.05.010
- Senevirathne, N. D., Okamoto, T., Takahashi, J., Umetsu, K., & Nishida, T. (2012).
 Effect of Mixed Microbial Culture Treatment on the Nutritive Value of Coffee,
 Green Tea and Oolong Tea Residues and the Effect of the Fermented Residues on
 in Vitro Rumen Fermentation. *APCBEE Procedia*, 4, 66–72.
 https://doi.org/10.1016/j.apcbee.2012.11.012
- Seo, J., Jung, J. K., & Seo, S. (2015). Evaluation of nutritional and economic feed values of spent coffee grounds and *Artemisia princeps* residues as a ruminant feed using *in vitro* ruminal fermentation. *PeerJ*, *3*, e1343. https://doi.org/10.7717/peerj.1343
- Somnuk, K., Eawlex, P., & Prateepchaikul, G. (2017). Optimization of coffee oil extraction from spent coffee grounds using four solvents and prototype-scale

extraction using circulation process. *Agriculture and Natural Resources*, 51(3), 181–189. https://doi.org/10.1016/j.anres.2017.01.003

- United Nations, 2022. (n.d.). United Nations Department of Economic and Social Affairs, Population Division (2022). World Population Prospects 2022: Summary of Results. UN DESA/POP/2022/TR/NO. 3.
- Waghorn, G. C., & McNabb, W. C. (2003). Consequences of plant phenolic compounds for productivity and health of ruminants. *Proceedings of the Nutrition Society*, 62(2), 383–392. https://doi.org/10.1079/PNS2003245
- Wang, X., Song, J., Liu, Z., Zhang, G., & Zhang, Y. (2022). Fermentation Quality and Microbial Community of Corn Stover or Rice Straw Silage Mixed with Soybean Curd Residue. *Animals*, 12(7), 919. https://doi.org/10.3390/ani12070919
- Wong, P. K., Ghadikolaei, M. A., Chen, S. H., Fadairo, A. A., Ng, K. W., Lee, S. M. Y., Xu, J. C., Lian, Z. D., Li, S., Wong, H. C., Zhao, J., Ning, Z., & Gali, N. K. (2022).
 Physicochemical and cell toxicity properties of particulate matter (PM) from a diesel vehicle fueled with diesel, spent coffee ground biodiesel, and ethanol. *Science of The Total Environment*, 824, 153873. https://doi.org/10.1016/j.scitotenv.2022.153873
- Xu, C. C., Cai, Y., Zhang, J. G., & Ogawa, M. (2007). Fermentation quality and nutritive value of a total mixed ration silage containing coffee grounds at ten or twenty percent of dry matter1. *Journal of Animal Science*, 85(4), 1024–1029. https://doi.org/10.2527/jas.2005-628
- Yun, B. Y., Cho, H. M., Kim, Y. U., Lee, S. C., Berardi, U., & Kim, S. (2020). Circular reutilization of coffee waste for sound absorbing panels: A perspective on material

recycling.	Environmental	Research,	184,	109281.
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https://doi.org/10.1016/j.envres.2020.109281

Chapter II

Citric acid: Impacts on Nutrient Feed Intake, Digestibility and

Rumen Fermentation in Sheep

Abstract

The objective of this study was to explore the potential of citric acid as a feed additive for mitigating enteric methane (CH₄) emissions in sheep. In the current study, we measured nutrient intake, digestibility, ruminal fermentation (at 0, 3, 6, 9, and 12 hours after feeding), and CH₄ emissions with citric acid-supplemented Corriedale sheep. The experimental design involved a replicated 2×2 crossover with 15-day periods and included four Corriedale wether sheep (initial BW of 77.5 ± 2.75 kg). The treatments consisted of a control group (no additive) and citric acid (40 *mM*). The basal diet comprised Klein grass hay and a commercial concentrate mixture at a ratio of 2:1. Four respiration chambers for experimental treatments (animals fed the same diet two by two) were equipped to measure CH₄ emissions for two days in each period. The results of this study indicated that citric acid, when added as an additive to the sheep diet at high dosages, led to a reduction in animal feed intake and digestibility without suppressing CH₄ production.

II.1 Introduction

CH₄ emissions are an unavoidable consequence of the natural digestive process occurring in both wild and domesticated ruminant animals, including cattle, goats, sheep, and buffalos (Kelly & Kebreab, 2023). Within the intricate ecosystem of the ruminant's digestive tract, microbes break down food and fibers, providing essential energy and nutrients to the animal while simultaneously releasing CH₄ as a by-product of digestible energy loss. This biological phenomenon, known as enteric fermentation, stands as a significant contributor to CH₄ emissions arising from livestock. Approximately 12 percent of a ruminant's energy intake typically dissipates as CH₄ through this enteric fermentation process (Hill et al., 2016). However, the specific amount of CH₄ expelled by the animal is contingent upon several factors, such as the type and quality of the animal's diet, its health condition, reproductive status, and the surrounding environmental factors (Broucek, 2014).

Consequently, substantial efforts have been dedicated to mitigating CH₄ emissions from the rumen by regulating rumen fermentation through dietary manipulation. This approach primarily involves the utilization of feed or feed additives (Ahmed et al., 2022). Currently, feed additives are undergoing approval processes for practical application in numerous countries globally (Palangi & Lackner, 2022). Moreover, ongoing research is actively exploring additional functional additive candidates to broaden the array of options for alternative materials. The exploration of alternative additive candidates is crucial due to limited availability, particularly when locally or seasonally produced, and the persistent concern regarding potential diminishing effectiveness over time, as seen with antibiotic additives. Hence, having multiple readily available additives could significantly enhance the sustainable application of methane-reducing solutions on a

global scale. Therefore, some of the alternative feed additives, such as organic acids, are suppressing CH₄ production in the rumen indirectly. For example, fumaric acid, malic acid, formic acid, and other organic acids have been shown to reduce the amount of CH₄ in ruminants *in vitro* (Newbold & Rode, 2006; Palangi & Macit, 2021; Yamada et al., 2023) and *in vivo* conditions (Bayaru et al., 2001; Beauchemin & McGinn, 2006; Foley et al., 2009). However, not all studies have confirmed that organic acids suppress CH₄ in the field (Beauchemin & McGinn, 2006). Additionally, nowadays, animal nutritionists are interested in citrus-based products for ruminants or non-ruminants as a CH₄ suppressor or protein source (Yamada et al., 2023). Although Tanpong et al. (2021) used the by-product of citric acid as an energy source for quails, and their team also described citric acid by-products as an alternative energy source in animal feed (Tanpong et al., 2019). Thus, Suriyapha et al. (2022) described that citric acid waste fermented yeast waste did not have any significant effect on the ruminant's intake or digestibility. Furthermore, they described citric acid waste fermented yeast waste that could be used as an alternative source of additive without any negative impact on tropical lactating cows.

The aims of this study were to investigate the impact of citric acid, a feed additive approved for sheep, on enteric CH₄ production. Our investigation aimed to validate specific *in vitro* findings, as reported by Yamada et al. (2023), through *in vivo* analysis. This approach allowed us to bridge the gap between laboratory *in vitro* results and real-life *in vivo* conditions, providing a comprehensive understanding of the potential effects of citric acid as a CH₄ suppressor in the context of live sheep.

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II.2 Material and method

II.2.1 Animals, treatments, and experimental procedures

The experimental design, animal handling, and sample collection processes were supervised by the Obihiro University of Agriculture and Veterinary Medicine, Animal Care, and Use Committee (approval number 22-182). The selected four sheep were matched in age and body weight, with an average weight of 77.5±2.75 kg, for a crossover design. The experiment spanned two periods of 15 days each, totaling 30 days, including 10 days for adaptation and 5 days for sampling. All animals were housed in a metabolic cage equipped with ventilated respiratory collection hoods individually, providing a sufficient place for standing or lying down comfortably.

The animals were fed twice a day at 8:30 and 16:30 h with a basal diet at a rate of 55 g dry matter (DM)/kg of BW ^{0.75}/day, following the recommendation of Pen et al. (2007). The diet consisted of Kleingrass (*Panicum coloratum*) hay and a commercial concentrate mixture (Sky Dairy, Chubu Feed Co., Ltd., Aichi, Japan) at a ratio of 2:1. All four sheep had unrestricted access to clean fresh water and a lick mineral block containing iron oxide (1742 mg), ferric oxide (196 mg), copper sulfate (377 mg), cobalt sulfate (66 mg), zinc sulfate (1235 mg), manganese carbonate (1046 mg), calcium iodate (77 mg), sodium selenite (33 mg), and sodium chloride (971 g/kg) (KOEN® SELENICS TZ, Nippon Zenyaku Kogyo Co., Fukushima, Japan).

The treatment divided into two groups: a control (0 mM citric acid) and a treatment group (40 mM citric acid). The additional amount of citric acid per day is mixed through concentrates with the morning feed only. Animals were kept in the same room, keeping lights on, from 8 h to 17 h. Samples of the offered diet, refusal, fecal and urine samples

were gathered throughout the collection period and subjected to analysis for nutrient content using established protocols.

II.2.2. Chemical analyses and apparent total-tract digestibility assay

All the refused feed, feces, and urine were sampled over a 5-day period. The collection period was adjusted from day 10 to day 14, and the samples were kept frozen until the next analysis. When the collection period ended, the sampled feces were thawed at temperatures between 18-24°C for approximately 24 hours. After the thawing process, a 200g subsample from each treatment was dried at 60°C for 48 hours in the oven. The dried samples were then ground for subsequent laboratory analysis to determine apparent total tract digestibility. Daily urine collection was done using buckets with 100 mL of 10% (v/v) H₂SO₄ to maintain a pH below 3, with approximately 150 mL of the collected urine stored daily in the freezer for subsequent nitrogen (N) analysis. Following the AOAC International (1995) procedures offered and refused feed, feces, and urine samples were analyzed in triplicate for each treatment and period. Dry matter (DM) content was determined by drying samples in an oven at 135°C for 2 hours (method 930.15). Organic matter (OM) and ash were assessed by subjecting samples to a muffle furnace at 500°C for 3 hours (method 942.05). N content in samples and urine was determined using the Kjeldahl method (method 984.13), and crude protein (CP) was estimated as N \times 6.25. An ANKOM200 fiber analyzer (Ankom Technology Methods 6, 5, and 8; ANKOM Technology Corp., Macedon, NY, USA) was used to measure neutral detergent fiber (NDF), acid detergent fiber (ADF), and acid detergent lignin. The chemical composition of the basal diet is detailed in Table 1.

Item ¹	Klein Grass	Concentrate
DM (g/kg in	908.6	884.4
fresh matter)		
OM	909.1	943.6
СР	140.8	180.0
EE	20.6	35.0
NDF	696.2	524.4
ADF	366.7	97.0
ADL	89.6	20.0

Table 1. Chemical composition of basal diet (g/kg Dry

¹DM: Dry Matter; OM: Organic Matter; CP: Crude Protein; EE: Either Extract; NDF: Neutral Detergent Fiber; ADF: Acid Detergent Fiber; ADL: Acid Detergent Lignin

II.2.3. Daily monitoring and health condition

Matter)

The health status of the animals was monitored twice a day, checking for cough, diarrhea, nasal and ocular discharges. Body weight and average daily gain (ADG) were recorded weekly, and feed intake (FI) was documented daily.

II.2.4. Gas measurement and formula

The measurement of CH₄ and CO₂ occurred over a two-day period (days 13 and 14) using open-circuit respiration chambers. Each sheep was positioned in a gas collection chamber designed to maintain the sheep's head inside for the systematic collection of gas

emissions. CH₄ and CO₂ emissions from each chamber were recorded using an infrared gas analyzer. The machine of the gas analyzer was linked to a personal computer (PC) and programmed for data collection from each respiration cage, following a previously described protocol (Takahashi et al., 1999).

Before commencing gas measurements every single period on the 13th day morning, the analyzer underwent calibration using a calibrating gas containing 2.04% CO₂ and 907 ppm CH₄. The calculation of CH₄ energy involved determining the gas volume obtained from the respiratory chamber, following the formula: CH₄ energy (kJ) = CH₄ (L) × 39.57 (BROUWER E., 1965).

II.2.5. Rumen fluid sampling and rumen fermentation parameter analysis

On day 15, rumen fluid was gathered at five distinct intervals: prior to morning feeding (0 h), and then at 3, 6, 9, and 12 hours after morning feeding. The collection of rumen fluid samples involved inserting a stomach tube through the esophagus and utilizing a vacuum pump to extract the fluid. The initial portion of rumen fluid was not used as a sample, and the subsequent portion was promptly filtered through two layers of medical gauze. Around 45 mL of rumen fluid from each treatment was amassed in 50 mL tubes. A quick electronic pH meter measurement of the rumen fluid's pH took place. Upon reaching the laboratory, 3 mL from each tube was further transferred into two 2 mL Eppendorf tubes[®]. Volatile fatty acids (VFA) were analyzed following the procedures previously outlined by Ahmed et al. (2021).

II.2.6. Statistical analysis

In the analytical process, a replicated 2 × 2 design was deployed, utilizing the MIXED model procedure within SAS 9.4 (SAS Institute Inc., Cary, NC, USA). This model systematically factors in treatment and period as fixed effects while considering animals as random effects. For the examination of CH₄ and CO₂, a comprehensive analysis spanning 2 days was specifically integrated as fixed effects. Furthermore, the model applied to rumen fermentation data embraced treatment interactions as fixed effects. Significance was attributed to differences at p < 0.05, with tendencies acknowledged within the range of 0.05 . The elucidation of results adopted the presentation of least squares means, complemented by the standard error of the mean (SEM). To unravel the correlation coefficients among variables, the implementation of PROC CORR within SAS was executed, employing Pearson's method and a two-tailed test to gauge significance.

II.3 Results

II.3.1 Citric acid supplementation on nutrient intake in sheep

The treatments, whether supplemented with or without citric acid, did not display any signs of illness or disease throughout the entire duration of the experiment.

The intricate examination of nutrient intake in sheep, as delineated in Table 2, reveals the nuanced impact of including citric acid in the basal diet. Notably, the incorporation of citric acid resulted in a discernible decrease in fresh matter intake (FI), DM and CP (p > 0.05). Moreover, trends in reduced OM intake were observed among the sheep, albeit not reaching statistical significance (p > 0.05). The intake of NDF, while not demonstrating statistically significant outcomes in the current study (p < 0.09), reflects the complex interplay of dietary components and the potential influence of citric acid on the nutritional dynamics of the basal diet. This multifaceted exploration underscores the need for a comprehensive understanding of the intricate relationships between dietary additives and nutrient intake in ruminants.

Item ¹				
(g/day)	Control	Citric acid	SEM	<i>P</i> -value
FI	1564.19	909.34	165.49	0.040
DM	1353.03	796.19	141.04	0.040
ОМ	1241.25	685.27	133.80	0.052
СР	189.02	101.77	22.00	0.040
NDF	292.54	192.30	57.26	0.090

Table 2. Citric acid supplementation on nutrient intake in sheep

¹FI: Fresh matter intake; DM: Dry matter; OM: Organic matter; CP: Crude protein; NDF: Neutral

detergent fiber

II.3.2 Citric acid on apparent total tract digestibility

Table 3 serves as a detailed repository, offering a thorough examination of apparent nutrient digestibility concerning various dietary interventions, with a particular emphasis on the incorporation of citric acid into the dietary regimen of sheep. This comprehensive analysis delves into the nuanced responses of the sheep's digestive system to the varying nutritional treatments explored in the study. Notably, the specific focus on citric acid inclusion in the sheep's diet unveils insightful observations.

Within this context, it is noteworthy to highlight that the augmentation of the diet with citric acid yielded results indicative of a lack of substantial influence on apparent ruminal digestibility. Despite the intricate interplay of dietary components and the physiological intricacies of the ruminal environment, the addition of citric acid emerged as a factor with no discernible impact on the sheep's digestive processes, particularly in the rumen.

This nuanced exploration not only contributes to the understanding of the dynamic relationship between dietary composition and nutrient assimilation but also sheds light on the specific role, or lack thereof, that citric acid plays in the intricate tapestry of the sheep's digestive physiology. The findings presented in Table 3 thus provide a valuable resource for researchers and practitioners seeking a deeper comprehension of the effects of citric acid supplementation on apparent nutrient digestibility in the context of sheep nutrition.

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Item ¹				Р-
(g/kg)	Control	Citric acid	SEM	value
DM	665.56	475.80	64.39	0.163
ОМ	648.34	402.27	76.10	0.167
СР	722.94	521.41	63.81	0.120
NDF	576.87	266.60	106.64	0.192

Table 3. Addition of citric acid on apparent total tract digestibility of

nutrients in sheep

¹DM: Dry matter; OM: Organic matter; CP: Crude protein; NDF: Neutral detergent fiber

II.3.3 Citric acid supplementation on CH4 emissions

Table 5 provides a comprehensive overview of the influence of incorporating citric acid on CH₄ emissions. The various metrics used to measure CH₄ emission, including L/day, g/day, L/kg DM intake, g/kg DM intake, L/kg OM, g/kg OM, g/kg DDMI, and g/kg DOMI, did not show any effects that were statistically significant. Despite observing a numerical reduction in CH₄ production in ml/min/MBW, L/day and g/day, it is noteworthy that the intakes for CH₄ production displayed a numerical increase. These findings suggest a nuanced impact of citric acid on CH₄ emission metrics, warranting further exploration and consideration in the broader context of ruminant dietary management.

CH4 emission ¹	Control	Citric acid	SEM	Р-
	Control	Chine actu	SEIVE	value
ml/min/MBW	0.51	0.48	0.02	0.579
L/day	19.81	18.53	0.99	0.064
g/day	14.15	13.24	0.71	0.064
L/kg DMI	14.14	28.65	4.48	0.155
g/kg DMI	10.15	20.46	3.19	0.157
L/kg OM	14.23	31.05	5.41	0.175
g/kg OM	10.16	22.18	3.86	0.175
g/kg DDMI	15.75	26.75	3.00	0.203
g/kg DOMI	17.67	36.14	5.07	0.200

*Table 5. Citric acid supplementation on CH*⁴ *emissions in sheep.*

¹MBW: Metabolic Body Weight, DMI: Dry Matter Intake, OMI: Organic Matter Intake, DDMI:

Digestible Dry Matter Intake, DOMI: Digestible Organic Matter Intake.

II.3.4 Citric acid supplementation on CO₂ emissions

Additionally, the result of CO₂ production is shown in Table 6. CO₂ production did not change with the inclusion of citric acid in any parameters, such as L/day, g/day, L/kg DM intake, and g/kg DM intake.

CO ₂	Control		CEM	D l
emissions ¹	Control	Citric acid	SEM	<i>P</i> -value
ml/min/MBW	6.21	5.77	0.55	0.539
L/day	241.66	223.15	11.64	0.106
g/day	172.61	159.39	8.31	0.106
L/kg DMI	173.59	336.42	47.68	0.128
g/kg DMI	123.99	240.30	34.06	0.128

Table 6. Addition of citric acid on Carbon dioxide (CO₂) emissions in sheep.

¹MBW: Metabolic Body Weight, DMI: Dry Matter Intake

II.3.5 Citric acid supplementation on Rumen Fermentation Profile

The basal diet treated with citric acid exhibited no significant effect on TVFA, showing a reduction as a tendency (p < 0.08). Acetate production, when compared to the control, did not yield any significant results (p > 0.1). However, concentrations of propionate and butyrate (mmol/L) showed a notable decrease, with a significant reduction of 18.92% and 25.82%, respectively (p < 0.05), when compared to the control group. Furthermore, the addition of citric acid to the diet did not result in a significant effect on the A/P ratio (p = 0.3). Exploring the proportions of the three main VFAs, acetate percentage demonstrated an increase of 4.57% (p = 0.004), while propionate and butyrate percentage production experienced a decrease with the inclusion of citric acid, showing reductions of 7.96% and 17.83%, respectively (p < 0.01). These nuanced shifts in VFAs emphasize the impact of citric acid on rumen fermentation patterns and underline the intricate nature of dietary interventions in influencing microbial processes in the rumen. Although there was no significant increase in pH with the citric acid supplement.

x7 · 11 1			GEN	Р-
Variable ¹	Control	Citric acid	SEM	value
TVFA (<i>mM</i>)	81.70	72.19	3.06	0.085
Acetate (<i>mM</i>)	58.77	54.20	1.80	0.164
Propionate (<i>mM</i>)	14.11	11.44	0.64	0.025
Butyrate (<i>mM</i>)	8.83	6.55	0.78	0.034
Acetate (%)	72.02	75.31	0.91	0.302
Propionate (%)	17.21	15.84	0.29	0.004
Butyrate (%)	10.77	8.85	0.74	0.005
A/P ratio	4.34	4.65	0.12	0.014
рН	6.63	6.78	0.09	0.112

Table 3. Citric acid supplementation on rumen fermentation profile in sheep.

¹ TVFA: Total Volatile Fatty Acids, A/P ratio: Acetate/Propionate ratio.

II.4 Discussion

Mitigating CH₄ emissions not only serves to reduce environmental impact but also holds the potential to increase ruminant feed efficiency. Methanogenesis, the process by which CH₄ is made in the rumen, needs energy. Finding ways to decrease CH₄ without negatively affecting rumen fermentative processes could result in both environmental and economic benefits for the cattle industry. Numerous synthesized and organic compounds have shown anti-methanogenic effects in *in vitro* conditions (Cattani et al., 2016; Chagas et al., 2019; Ku-Vera, Jiménez-Ocampo, et al., 2020). However, many of these compounds sometimes have negative impacts on animal intake, digestibility, and fermentation production (Patra et al., 2017).

In this study, citric acid was used as a feed additive to suppress products for ruminants, but it did not have any significant effect on CH₄ output. The high-forage diet used in this study consisted of Klein grass hay and a commercial concentrate mixture at a ratio of 2:1. Previously, Yamada et al. (2023) reported that citric acid, used as a feed additive for *in vitro* batch culture conditions, suppresses CH₄ production using the same ratio as the basal diet. Additionally, they also suggested that a high-forage basal diet treated with citric acid suppresses CH₄ production by 9%-10%. They reported that citric acid reduced CH₄ production without the accumulation of hydrogen and increased propionate but increased short-chain fatty acids (SCFA), especially acetate production in the rumen (Yamada et al., 2023). They also suggested that CH₄ production might occur through the acrylate pathway and discussed the necessity of testing this material on ruminant animals. However, *in vitro* and *in vivo* studies are incomparable, as *in vivo* processes are sometimes not accurately represented. Previous studies mostly describe that the supplementation of organic acids has a positive or no effect on feed intake and digestibility of ruminants (Bampidis &

Robinson, 2006; Clemmons et al., 2021; Gheller et al., 2020). However, in the current study, the addition of citric acid to feed significantly reduced nutrient intake, although the ruminal fermentation process, specifically total VFA and acetate, propionate, and butyrate, remained unaffected. In previous studies, feed intake decreased due to some feed additives, such as essential oils, ionophores, and other additives, and these materials also led to a lack of digestibility in the rumen (Azzaz et al., 2015; Beauchemin & McGinn, 2006; Hou et al., 2023). However, these materials decreased utilization and simultaneously suppressed CH₄ production too (Azzaz et al., 2015). In the current study, addition of citric acid did not affect the ruminal digestibility. Although, there was no significant reduction in CH₄ production or the fermentation process, but utilization was reduced, possibly due to the higher dosage of citric acid and potentially lower palatability for the animals.

II.5 Conclusions

This study demonstrates that citric acid reduces feed intake when added in high amounts. However, these materials did not affect the digestibility and fermentation process of the rumen. Although some *in vitro* studies have shown promising results in suppressing CH₄ emission, the translation to lower CH₄ emission *in vivo* is influenced by many other confounding factors. Further research is still required to identify optimal levels and usage methods for these products in commercial ruminant feed.

II.6 References

- Ahmed, E., Batbekh, B., Fukuma, N., Hanada, M., & Nishida, T. (2022). Evaluation of Different Brown Seaweeds as Feed and Feed Additives Regarding Rumen Fermentation and Methane Mitigation. *Fermentation*, 8(10), 504. https://doi.org/10.3390/fermentation8100504
- Ahmed, E., Batbekh, B., Fukuma, N., Kand, D., Hanada, M., & Nishida, T. (2021). A garlic and citrus extract: Impacts on behavior, feed intake, rumen fermentation, and digestibility in sheep. *Animal Feed Science and Technology*, 278, 115007. https://doi.org/10.1016/j.anifeedsci.2021.115007
- Azzaz, H. H., Murad, H. A., & Morsy, T. A. (2015). Utility of Ionophores for Ruminant Animals: A Review. Asian Journal of Animal Sciences, 9(6), 254–265. https://doi.org/10.3923/ajas.2015.254.265
- Bampidis, V. A., & Robinson, P. H. (2006). Citrus by-products as ruminant feeds: A review. Animal Feed Science and Technology, 128(3–4), 175–217. https://doi.org/10.1016/j.anifeedsci.2005.12.002
- BAYARU, E., KANDA, S., KAMADA, T., ITABASHI, H., ANDOH, S., NISHIDA, T., ISHIDA, M., ITOH, T., NAGARA, K., & ISOBE, Y. (2001). Effect of Fumaric Acid on Methane Production, Rumen Fermentation and Digestibility of Cattle Fed Roughage Alone. *Nihon Chikusan Gakkaiho*, 72(2), 139–146. https://doi.org/10.2508/chikusan.72.139
- Beauchemin, K. A., & McGinn, S. M. (2006). Methane emissions from beef cattle: Effects of fumaric acid, essential oil, and canola oil1. *Journal of Animal Science*, 84(6), 1489–1496. https://doi.org/10.2527/2006.8461489x

- Broucek, J. (2014). Production of Methane Emissions from Ruminant Husbandry: A Review. Journal of Environmental Protection, 05(15), 1482–1493. https://doi.org/10.4236/jep.2014.515141
- Cattani, M., Maccarana, L., Rossi, G., Tagliapietra, F., Schiavon, S., & Bailoni, L. (2016). Dose-response and inclusion effects of pure natural extracts and synthetic compounds on in vitro methane production. *Animal Feed Science and Technology*, 218, 100–109. https://doi.org/10.1016/j.anifeedsci.2016.05.014
- Chagas, J. C., Ramin, M., & Krizsan, S. J. (2019). In Vitro Evaluation of Different Dietary Methane Mitigation Strategies. *Animals*, 9(12), 1120. https://doi.org/10.3390/ani9121120
- Clemmons, B. A., Schneider, L. G., Melchior, E. A., Lindholm-Perry, A. K., Hales, K. E., Wells, J. E., Freetly, H. C., Hansen, S. L., Drewnoski, M. E., Hartman, S. J., & Myer, P. R. (2021). The effects of feeding ferric citrate on ruminal bacteria, methanogenic archaea and methane production in growing beef steers. *Access Microbiology*, 3(1). https://doi.org/10.1099/acmi.0.000180
- Foley, P. A., Kenny, D. A., Lovett, D. K., Callan, J. J., Boland, T. M., & O'Mara, F. P. (2009). Effect of dl-malic acid supplementation on feed intake, methane emissions, and performance of lactating dairy cows at pasture. *Journal of Dairy Science*, 92(7), 3258–3264. https://doi.org/10.3168/jds.2008-1633
- Gheller, L. S., Ghizzi, L. G., Marques, J. A., Takiya, C. S., Grigoletto, N. T. S., Dias,
 M. S. S., Silva, T. B. P., Nunes, A. T., da Silva, G. G., Fernandes, L. G. X., Rennó,
 L. N., & Rennó, F. P. (2020). Effects of organic acid-based products added to total mixed ration on performance and ruminal fermentation of dairy cows. *Animal*

FeedScienceandTechnology,261,114406.https://doi.org/10.1016/j.anifeedsci.2020.114406

- Hill, J., McSweeney, C., Wright, A.-D. G., Bishop-Hurley, G., & Kalantar-zadeh, K.
 (2016). Measuring Methane Production from Ruminants. *Trends in Biotechnology*, 34(1), 26–35. https://doi.org/10.1016/j.tibtech.2015.10.004
- Hou, Z., Liu, J., Cai, M., Liu, Y., Zhang, M., Wang, L., Yang, W., & Huang, B. (2023).
 The volatile organic compounds and palatability of mixed ensilage of marigold (Tagetes erecta L.) crop residues. *Scientific Reports*, 13(1), 2080. https://doi.org/10.1038/s41598-023-28511-5
- Kelly, L., & Kebreab, E. (2023). Recent advances in feed additives with the potential to mitigate enteric methane emissions from ruminant livestock. *Journal of Soil and Water Conservation*, 78(2), 111–123. https://doi.org/10.2489/jswc.2023.00070
- Ku-Vera, J. C., Jiménez-Ocampo, R., Valencia-Salazar, S. S., Montoya-Flores, M. D., Molina-Botero, I. C., Arango, J., Gómez-Bravo, C. A., Aguilar-Pérez, C. F., & Solorio-Sánchez, F. J. (2020). Role of Secondary Plant Metabolites on Enteric Methane Mitigation in Ruminants. *Frontiers in Veterinary Science*, 7. https://doi.org/10.3389/fvets.2020.00584
- Newbold, C. J., & Rode, L. M. (2006). Dietary additives to control methanogenesis in the rumen. *International Congress Series*, 1293, 138–147. https://doi.org/10.1016/j.ics.2006.03.047
- Palangi, V., & Lackner, M. (2022). Management of Enteric Methane Emissions in Ruminants Using Feed Additives: A Review. Animals, 12(24), 3452. https://doi.org/10.3390/ani12243452

- Palangi, V., & Macit, M. (2021). Indictable Mitigation of Methane Emission Using Some Organic Acids as Additives Towards a Cleaner Ecosystem. *Waste and Biomass Valorization*, 12(9), 4825–4834. https://doi.org/10.1007/s12649-021-01347-8
- Patra, A., Park, T., Kim, M., & Yu, Z. (2017). Rumen methanogens and mitigation of methane emission by anti-methanogenic compounds and substances. *Journal of Animal Science and Biotechnology*, 8(1), 13. https://doi.org/10.1186/s40104-017-0145-9
- Pen, B., Takaura, K., Yamaguchi, S., Asa, R., & Takahashi, J. (2007). Effects of Yucca schidigera and Quillaja saponaria with or without β 1–4 galacto-oligosaccharides on ruminal fermentation, methane production and nitrogen utilization in sheep. *Animal Feed Science and Technology*, 138(1), 75–88. https://doi.org/10.1016/j.anifeedsci.2006.11.018
- Suriyapha, C., Supapong, C., So, S., Wanapat, M., & Cherdthong, A. (2022). Bioconversion of agro-industrial residues as a protein source supplementation for multiparous Holstein Thai crossbreed cows. *PLOS ONE*, *17*(9), e0273916. https://doi.org/10.1371/journal.pone.0273916
- Tanpong, S., Cherdthong, A., Tengjaroenkul, B., Reungsang, A., Sutthibak, N., & Wongtangtintharn, S. (2021). A study on citric acid by-product as an energy source for Japanese quail. *Tropical Animal Health and Production*, 53(5), 474. https://doi.org/10.1007/s11250-021-02920-y
- Tanpong, S., Cherdthong, A., Tengjaroenkul, B., Tengjaroenkul, U., & Wongtangtintharn, S. (2019). Evaluation of physical and chemical properties of

citric acid industrial waste. *Tropical Animal Health and Production*, 51(8), 2167–2174. https://doi.org/10.1007/s11250-019-01917-y

Yamada, K., Iwamae, K., Suzuki, Y., Koike, S., & Kobayashi, Y. (2023). Batch culture analysis to identify potent organic acids for suppressing ruminal methane production. *Animal Science Journal*, 94(1). https://doi.org/10.1111/asj.13873 Chapter III

Itaconic acid: Impacts on Feed Intake, Digestibility and

Rumen Fermentation in Sheep

Abstract

Methane (CH₄) is a major contributor to GHG emissions globally, and its increasing production poses significant threats to humans, animals, and the environment. The livestock sector, particularly ruminant animals, is responsible for approximately 40% of agricultural CH₄ emissions.

The aim of this study was to identify organic acids that can effectively reduce enteric CH₄ emissions from ruminants. The research assessed the impact of an organic acid, namely itaconic acid, on feed intake, digestibility, CH₄ emissions, and ruminal fermentation in Corriedale sheep following a specific diet. The experimental design involved replicated 2×2 and 15-day periods. The present study utilized four Corriedale wether sheep (initial BW of 76.5±5.1 kg) as a treatment.

The treatments consisted of two animals for the control group (no additive) and another two animals for itaconic acid (40 m). The basal diet included Klein grass and a commercial concentrate mixture at a ratio of 2:1. Four chambers were equipped to measure CH₄ emissions over a 2-day period in each cycle. Feces, urine, and refusal samples were collected throughout the collection period. Rumen fluid was collected at five different times, including before morning feed, at 3-hour intervals, with the final collection 12 hours after morning feed.

The present study did not show any significant results in CH₄ production for all collected data except CH₄ emissions (L/kg DMI). CH₄ emissions (L/kg DMI) increased significantly in the study, and there were also some significant changes in rumen fermentation results. All fermentation parameters, whether statistically or non-statistically, showed a reduction in this study, and due to this result, total VFA was also significantly

reduced (p > 0.04). Furthermore, itaconic acid is potentially a feed additive for the livestock sector, but there is still limited information on its advantages and disadvantages. Further research is needed to clarify the potential of this product.

III.1 Introduction

CH₄, the second GHG following carbon dioxide CO₂, possesses a higher energyabsorbing capacity than CO₂. Moreover, CH₄ boasts a 28-fold greater greenhouse potential than CO₂ (Intergovernmental Panel on Climate Change (IPCC), 2023). The surge in atmospheric CH₄ concentrations is attributed to burgeoning human populations, intensified feed production, and anthropogenic sources driving CH₄ production (Philip Thornton & Mario Herrero, 2010).

Ruminant animals, including cattle, sheep, and goats, inherently emit CH₄ during their physiological digestive processes (Soren et al., 2017). Notably, cattle and dairy cows emerged as the predominant contributors to CH₄ emissions in 2020, constituting 72% of the total sector emissions (Reisinger et al., 2021).

Throughout anaerobic bacterial fermentation, CH₄ emission arises in the rumen and the hindgut and is subsequently released into the environment (la Fuente et al., 2019). Microorganisms break down feed components into smaller molecules like amino acids, simple sugars, and volatile compounds (Terry et al., 2019). Bacteria and protozoa constitute 80% of the microbial mass inside the rumen, generating CH₄ (Newbold & Ramos-Morales, 2020). This process, however, results in a loss of 2–12% of the host's metabolic energy intake. Hence, a global endeavor is imperative to curtail ruminant emissions (Günal et al., 2019).

Feed additives assume the role of methanogen inhibitors, both directly and indirectly. In recent years, researchers have considered strategies for mitigating enteric CH₄, particularly focusing on organic acids such as malic and fumaric acids. These acids, intermediates of the citric acid cycle in the propionate-succinate pathway, provide an alternative electron-sink pathway, diverting hydrogen that might otherwise contribute to

methanogenesis. Studies by Bayaru et al. (2001) and Carro & Ranilla (2003) extensively explored the *in vitro* effects of fumaric and malic acids, reporting dose-dependent reductions in CH₄ production. Nonetheless, certain *in vitro* studies, exemplified by Czerkawski & Breckenridge (1972), suggested that specific organic acids might stimulate CH₄ production.

The methane-suppressing mechanism is indirect, involving the reduction of fumarate in the rumen, thereby altering the hydrogen-metabolizing pathway. This shift induces a transition from CH₄ to propionate production, a pivotal step in reducing CH₄ emissions. Furthermore, organic acids like citrate, gluconate, and itaconate, prevalent in the swine and poultry industries, await thorough evaluation for their impact on rumen metabolism. It is possible that these acids may regulate the rumen microbiota and metabolic activity, contributing to CH₄ suppression. Additionally, potential candidates for suppressing ruminal CH₄ production include known acids used in synthesizing fumarate and malate, potentially enhancing propionate production. For example, the addition of fumarate to rumen batch cultures has demonstrated a decrease in CH₄ production and an increase in propionate production (Newbold et al., 2005). Therefore, exploring precursors and intermediates of rumen metabolic pathways as candidate additives is essential for comprehending their potential to mitigate CH₄ emissions.

The aim of our present study was to investigate the impact of a novel feed additive, itaconic acid, registered for feeding to sheep, on enteric CH₄ production and utilization in ruminants. Intriguingly, several previous studies have highlighted itaconic acid's anti-inflammatory and anti-pathogenic activities in mammals (Cordes et al., 2015; O'Neill & Artyomov, 2019; Peace & O'Neill, 2022). However, there is limited information available

regarding the use of itaconic acid in ruminant feed. Only Yamada et al. (2023) have described its potential in mitigating CH₄ production from ruminants, albeit in an *in vitro* setting. Our study builds upon this, confirming these *in vitro* results through *in vivo* analysis.

The inclusion of itaconic acid as a registered feed supplement for sheep prompted our investigation into its effects on enteric CH₄ production. Notably, while previous research suggests anti-inflammatory and anti-pathogenic properties in mammals, its application in ruminant feed remains an understudied area. Previously, some studies provided a valuable *in vitro* perspective, demonstrating CH₄ mitigation potential. Our study bridges the gap by extending these findings to *in vivo* conditions, thus contributing to the understanding of itaconic acid's role in ruminant nutrition.

III.2 Material and method

III.2.1 Animals, treatments, and experimental procedures

The experimental design, animal care, and sampling process of the current study were all approved by the Obihiro University of Agriculture and Veterinary Medicine Animal Care and Use Committee (approval numbers 23-37). Each sheep, weighing 76.5 \pm 5.1 kg, was used in a 2 x 2 cross-over design. The experiment spanned a total of 30 days, divided into two 15-day periods. Each period consisted of an adaptation period of 10 days and a 5-day sampling period. The current study treatment comprised a control group (0 *mM* itaconic acid) and a treatment group (20 *mM* itaconic acid). The further methods used are all followed by a second chapter.

The chemical composition of the basal diet is described in Table 1.

Item ¹	Klein Grass	Concentrate	
DM (g/kg fresh	908.6	884.4	
matter)			
ОМ	909.1	943.6	
СР	140.8	180.0	
EE	20.6	35.0	
NDF	696.2	524.4	
ADF	366.7	97.0	
ADL	89.6	20.0	

Table 1. Chemical composition of basal diet (g/kg Dry

Matter)

¹DM: Dry matter; OM: Organic matter; CP: Crude protein; EE: Either extract; NDF: Neutral detergent fiber; ADF: Acid detergent fiber; ADL: Acid detergent lignin

III.3 Results

III.3.1 Addition of itaconic acid to nutrient intake

There were no observable changes in DM and OM intake (p > 0.05) following the inoculation of itaconic acid in the sheep's diet, as indicated in Table 3. Furthermore, the intake of crude protein (g/d) did not exhibit significant variations (p > 0.05), and the intake of neutral detergent fiber (NDF) remained consistent (p > 0.05) between the groups supplemented with itaconic acid and those without supplementation.

Item ¹	Control	Itaconic acid	SEM	<i>P</i> -value
FI	1387.74	1326.07	54.64	0.27
DM	1367.14	1318.84	20.43	0.30
ОМ	1263.26	1218.09	19.11	0.30
СР	193.70	182.39	4.40	0.30
NDF	716.08	678.82	16.56	0.30

Table 2. Addition of itaconic acid on nutrient intake in sheep (g/day).

¹ FI: Fresh Matter Intake; DM: Dry Matter; OM: Organic Matter; CP: Crude Protein; NDF: Neutral Detergent Fiber

III.3.2 Addition of itaconic acid to apparent nutrient digestibility

Table 3 presents a comprehensive overview of the apparent nutrient digestibility in response to dietary treatments, specifically the inclusion of itaconic acid in the sheep's diet. The digestibility of DM, OM, NDF, and CP showed no discernible impact attributable to the addition of itaconic acid (p > 0.05).

Table 3. Addition of itaconic acid on apparent total tract digestibility of nutrients

Item ¹	Control	Itaconic acid	SEM	<i>P</i> -value
DM	694.20	702.21	10.12	0.68
ΟΜ	671.07	678.32	12.12	0.76
СР	754.37	739.46	5.37	0.13
NDF	592.17	589.72	18.48	0.93

(g/kg) in sheep

¹DM: Dry Matter; OM: Organic Matter; CP: Crude Protein; NDF: Neutral Detergent Fiber

III.3.3 Addition of itaconic acid to Rumen Fermentation

The impact of itaconic acid concentration on ruminal parameters was examined in this study. Surprisingly, ruminal pH remained unaffected by the inclusion of itaconic acid in the diet. However, a more detailed analysis revealed significant alterations in total volatile fatty acids (VFAs) and propionate concentration due to the supplementation of itaconic acid, as highlighted in Table 4. While other individual VFAs did not display significant changes, there was a distinct rise in the proportion of acetate by 2.75%. Intriguingly, propionate and butyrate production experienced a reduction of 14.19% and 5.41% respectively, when compared to the control group. These findings emphasize the nuanced effects of itaconic acid on ruminal fermentation dynamics.

Variable ¹	Control	Itaconic acid	SEM	Р-
				value
TVFA (<i>mM</i>)	79.69	74.47	2.92	0.043
Acetate (<i>mM</i>)	56.64	55.22	2.05	0.281
Propionate (<i>mM</i>)	14.32	11.48	0.79	0.013
Butyrate (<i>mM</i>)	8.42	8.18	0.31	0.611
Acetate (%)	71.57	73.54	0.56	0.008
Propionate (%)	17.90 ^a	15.36	0.52	0.028
Butyrate (%)	10.53	11.10	0.45	0.011
A/P ratio	4.01	4.82	0.17	0.383
рН	6.66	6.64	0.05	0.919

Table 4. Effect of Itaconic acid on rumen fermentation profile at different times in sheep.

¹ TVFA: Total Volatile Fatty Acids, A/P ratio: Acetate/Propionate ratio.

III.3.4 Addition of itaconic acid to CH4 emission

In the *in vivo* assessment, the inclusion of itaconic acid in the sheep's diet did not show any significant changes in CH₄ emissions (L/d) or (g/d) (p > 0.05), as indicated in Table 5. However, CH₄ production parameters, such as g/kg OMI, DDM and DOMI, did not demonstrate statistically significant variations.

CH ₄ emission ¹	Control	Itaconic acid	SEM	<i>P</i> -
		Itaconic aciu		value
ml/min/MBW	0.62	0.64	0.02	0.512
L/day	23.13	24.03	0.54	0.486
g/day	16.52	17.16	0.39	0.486
L/kg DMI	16.96	18.21	0.35	0.174
g/kg DMI	12.12	12.95	0.25	0.174
L/kg OM	17.07	18.33	0.35	0.152
g/kg OM	12.19	13.09	0.25	0.152
g/kg DDMI	28.32	30.90	1.63	0.094
g/kg DOMI	18.29	20.80	0.88	0.194

*Table 5. Effect of Citric acid supplementation on CH*⁴ *emissions in sheep.*

¹MBW: Metabolic Body Weight, DMI: Dry Matter Intake, OMI: Organic Matter Intake, DDMI: Digestible Dry Matter Intake, DOMI: Digestible Organic Matter Intake.

III.4 Discussion

CH₄ emissions were not affected when itaconic acid was included in a sheep diet at a dose of 20 *mM*. Several studies, including Yamada et al. (2023), have reported that the addition of 10 to 30 *mM* of organic acids in the ratio of ruminants mitigates CH₄ production significantly, by about 5 to 23% (Bayaru et al., 2001; Carro & Ranilla, 2003; Clemmons et al., 2021; Palangi & Macit, 2021). Carro & Ranilla (2003) reported that inoculating fumarate (from 0 to 10 *mM*) into substrate in batch culture increased the production of VFA, decreased the A:P ratio, and reduced CH₄ production by up to 5%. Similarly, studies (Islam & Lee, 2019) indicate that most organic acids reduce CH₄ production by acting as hydrogen sinks and stimulating the proliferation of ruminal microorganisms.

However, there is less information on the effects of organic acids used *in vivo* Bayaru et al. (2001) added 20 g/kg of DMI of fumaric acid (about 18 *mM*) into a cattle feed and found a 23% reduction in CH₄, along with a significant increase in propionate production. Similar results (Palangi & Macit, 2021) were observed with several organic acids, either separately or in a mixture, at varying levels on VFA and CH₄ mitigation in the Awassi Rams diet. They found a significant reduction in CH₄ production after 24 h of incubation, while some organic acids increased the propionic acid percentage, especially fumaric acid. Contrastingly, McGinn et al. (2004) and Beauchemin & McGinn (2006) fed approximately 15 *mM* and an estimated 50 *mM* of fumaric acids to cattle diets, respectively, reporting no effect on rumen fermentation or CH₄ emissions. Due to the demand for clarity on this topic, it is necessary to investigate recently used organics or suggest novel feed additives, such as new organic acids. Previously, Yamada et al. (2023) reported that itaconic acid suppressed CH₄ production by 60% without affecting rumen

fermentation. In the same study, the proportion of butyrate increased due to the increase of Megasphaera, an important bacterium in the rumen that metabolizes lactate to produce butyrate. According to Cordes et al. (2015), itaconic acid has antipathogenic activities, leading to changes in ruminal microbiome diversity when the sheep diet contains itaconic acid, as observed in our present study. Although our present study results showed a significant reduction in total VFA, propionate and butyrate (p < 0.05), the proportion of propionate in the rumen decreased, and the A/P ratio increased (p < 0.05). The propionate reduction result is in agreement with Yamada et al. (2023); contrastingly, they also described butyrate production as increased by itaconic acid, but our result did not follow this result.

Yamada et al. (2023) described that itaconate did not result in accumulating hydrogen as a feed in ruminants' diets; CH₄ suppression might have been due to the acrylate pathway via lactate production. These results disagree with our findings. In our study, CH₄ production was not suppressed by itaconic acid. Yamada et al. (2023) reported that rumen pH might also be reduced by itaconic acid, but in our findings, there was no effect on rumen pH (p > 0.05).

III.5 Conclusions

The inclusion of itaconic acid at 20 *mM* in sheep diets did not impact CH₄ emissions, contrasting previous findings with organic acids. While some *in vitro* studies have suggested CH₄ reduction potential with fumarate, translating these effects to live animals is less explored. Also, some *in vivo* studies showed promise in reducing cattle CH₄ by 23% with fumaric acid, but inconsistent results from other studies questioned its efficacy. Itaconic acid, whose previous *in vitro* study reported reducing CH₄ by 60%, yielded conflicting results in our study, indicating variability in its effectiveness. Overall, the search for reliable CH₄ suppressors continues, emphasizing the need for nuanced understanding and exploration of novel additives for consistent mitigation in ruminant diets.

III.6 References

- Ahmed, E., Batbekh, B., Fukuma, N., Kand, D., Hanada, M., & Nishida, T. (2021). A garlic and citrus extract: Impacts on behavior, feed intake, rumen fermentation, and digestibility in sheep. *Animal Feed Science and Technology*, 278, 115007. https://doi.org/10.1016/j.anifeedsci.2021.115007
- BAYARU, E., KANDA, S., KAMADA, T., ITABASHI, H., ANDOH, S., NISHIDA, T., ISHIDA, M., ITOH, T., NAGARA, K., & ISOBE, Y. (2001). Effect of Fumaric Acid on Methane Production, Rumen Fermentation and Digestibility of Cattle Fed Roughage Alone. *Nihon Chikusan Gakkaiho*, 72(2), 139–146. https://doi.org/10.2508/chikusan.72.139
- Beauchemin, K. A., & McGinn, S. M. (2006). Methane emissions from beef cattle: Effects of fumaric acid, essential oil, and canola oil1. *Journal of Animal Science*, 84(6), 1489–1496. https://doi.org/10.2527/2006.8461489x
- BROUWER E. (1965). Report of sub-committee on constants and factors. *Energy* Metabolism. Proc. 3rd Symp., 1965.
- Carro, M. D., & Ranilla, M. J. (2003). Effect of the addition of malate on *in vitro* rumen fermentation of cereal grains. *British Journal of Nutrition*, 89(2), 181–188. https://doi.org/10.1079/BJN2002759
- Clemmons, B. A., Schneider, L. G., Melchior, E. A., Lindholm-Perry, A. K., Hales, K. E., Wells, J. E., Freetly, H. C., Hansen, S. L., Drewnoski, M. E., Hartman, S. J., & Myer, P. R. (2021). The effects of feeding ferric citrate on ruminal bacteria, methanogenic archaea and methane production in growing beef steers. *Access Microbiology*, 3(1). https://doi.org/10.1099/acmi.0.000180

- Cordes, T., Michelucci, A., & Hiller, K. (2015). Itaconic Acid: The Surprising Role of an Industrial Compound as a Mammalian Antimicrobial Metabolite. *Annual Review of Nutrition*, 35(1), 451–473. https://doi.org/10.1146/annurev-nutr-071714-034243
- Czerkawski, J. W., & Breckenridge, G. (1972). Fermentation of various glycolytic intermediates and other compounds by rumen micro-organisms, with particular reference to methane production. *British Journal of Nutrition*, 27(1), 131–146. https://doi.org/10.1079/BJN19720077
- de la Fuente, G., Yañez-Ruiz, D. R., Seradj, A. R., Balcells, J., & Belanche, A. (2019). Methanogenesis in animals with foregut and hindgut fermentation: a review. *Animal Production Science*, *59*(12), 2109. https://doi.org/10.1071/AN17701
- Günal, M., McCourt, A., Zhao, Y., Yan, Z. G., & Yan, T. (2019). The effect of silage type on animal performance, energy utilisation and enteric methane emission in lactating dairy cows. *Animal Production Science*, 59(3), 499. https://doi.org/10.1071/AN16435
- Intergovernmental Panel on Climate Change (IPCC). (2023). *Climate Change 2022 Impacts, Adaptation and Vulnerability*. Cambridge University Press. https://doi.org/10.1017/9781009325844
- Islam, M., & Lee, S.-S. (2019). Advanced estimation and mitigation strategies: a cumulative approach to enteric methane abatement from ruminants. *Journal of Animal Science and Technology*, 61(3), 122–137. https://doi.org/10.5187/jast.2019.61.3.122
- McGinn, S. M., Beauchemin, K. A., Coates, T., & Colombatto, D. (2004). Methane emissions from beef cattle: Effects of monensin, sunflower oil, enzymes, yeast,

and fumaric acid1. *Journal of Animal Science*, 82(11), 3346–3356. https://doi.org/10.2527/2004.82113346x

- Newbold, C. J., López, S., Nelson, N., Ouda, J. O., Wallace, R. J., & Moss, A. R. (2005). Propionate precursors and other metabolic intermediates as possible alternative electron acceptors to methanogenesis in ruminal fermentation *in vitro*. *British Journal of Nutrition*, 94(1), 27–35. https://doi.org/10.1079/BJN20051445
- Newbold, C. J., & Ramos-Morales, E. (2020). Review: Ruminal microbiome and microbial metabolome: effects of diet and ruminant host. *Animal*, 14, s78–s86. https://doi.org/10.1017/S1751731119003252
- O'Neill, L. A. J., & Artyomov, M. N. (2019). Itaconate: the poster child of metabolic reprogramming in macrophage function. *Nature Reviews Immunology*, 19(5), 273–281. https://doi.org/10.1038/s41577-019-0128-5
- Palangi, V., & Macit, M. (2021). Indictable Mitigation of Methane Emission Using Some Organic Acids as Additives Towards a Cleaner Ecosystem. *Waste and Biomass Valorization*, 12(9), 4825–4834. https://doi.org/10.1007/s12649-021-01347-8
- Peace, C. G., & O'Neill, L. A. J. (2022). The role of itaconate in host defense and inflammation. *Journal of Clinical Investigation*, 132(2). https://doi.org/10.1172/JCI148548
- Pen, B., Takaura, K., Yamaguchi, S., Asa, R., & Takahashi, J. (2007). Effects of Yucca schidigera and Quillaja saponaria with or without β 1–4 galacto-oligosaccharides on ruminal fermentation, methane production and nitrogen utilization in sheep. *Animal Feed Science and Technology*, 138(1), 75–88. https://doi.org/10.1016/j.anifeedsci.2006.11.018

- Philip Thornton, & Mario Herrero. (2010). The Inter-Linkages between Rapid Growth in Livestock Production, Climate Change, and the Impacts on Water Resources, Land Use, and Deforestation. *World Bank Policy Research Working Paper No.* 5178, 5178.
- Reisinger, A., Clark, H., Cowie, A. L., Emmet-Booth, J., Gonzalez Fischer, C., Herrero, M., Howden, M., & Leahy, S. (2021). How necessary and feasible are reductions of methane emissions from livestock to support stringent temperature goals? *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering* Sciences, 379(2210), 20200452. https://doi.org/10.1098/rsta.2020.0452
- Soren, N. M., Sejian, V., Terhuja, M., & Dominic, G. (2017). Enteric Methane Emission in Sheep: Process Description and Factors Influencing Production. In *Sheep Production Adapting to Climate Change* (pp. 209–233). Springer Singapore. https://doi.org/10.1007/978-981-10-4714-5 10
- Takahashi, J., Chaudhry, A. S., Beneke, R. G., & Young, B. A. (1999). An open-circuit hood system for gaseous exchange measurements in small ruminants. *Small Ruminant Research*, 32(1), 31–36. https://doi.org/10.1016/S0921-4488(98)00163-1
- Terry, S. A., Badhan, A., Wang, Y., Chaves, A. V., & McAllister, T. A. (2019). Fibre digestion by rumen microbiota — a review of recent metagenomic and metatranscriptomic studies. *Canadian Journal of Animal Science*, 99(4), 678–692. https://doi.org/10.1139/cjas-2019-0024

Yamada, K., Iwamae, K., Suzuki, Y., Koike, S., & Kobayashi, Y. (2023). Batch culture analysis to identify potent organic acids for suppressing ruminal methane production. *Animal Science Journal*, 94(1). https://doi.org/10.1111/asj.13873

General Summary

The current thesis study contains two parts: the first one focuses on the use of byproducts from human consumption as feed or feed additives for ruminants *in vitro*, while the other, the main objective of our study, involves the addition of organic acids *in vivo* conditions, exploring the application of novel CH₄ suppressors. Each part includes both *in vitro* and *in vivo* experiments with a specific emphasis on mitigating CH₄ emissions, enhancing animal productivity, and at the same time, addressing economic concerns related to ruminant feed.

Ruminant livestock, notably cattle, supply meat and milk on a global scale. Nevertheless, the expanding global population and its growing appetite for meat and milk present formidable challenges to the livestock sector, particularly in the realm of environmental sustainability. While fulfilling nutritional requirements, ruminants make a substantial contribution to GHG emissions, prominently featuring CH₄ as a major constituent.

The escalating levels of GHGs, especially CH₄, raise concerns about climate change, prompting efforts to find solutions in the livestock industry. Nowadays, researchers highlight the inefficiency of ruminant digestion, leading to CH₄ emissions and energy loss. Researchers are actively seeking strategies to mitigate these emissions without compromising animal welfare or farmer profits. In the last decades, increased attention has been given to CH₄ mitigation technologies, with promising findings from dietary interventions. However, most of these strategies have economic disadvantages, conflict with animal welfare regulations, adverse effects of some approaches on animal performance, and rumen fermentation. Due to this, there is a need to find new materials

that can mitigate CH₄ and be beneficial for farmers and animals. Dietary interventions, including feed additives and by-products, are being explored as potential solutions. By-products, especially those derived from human food, are considered a potential idea for inclusion in animal diets. In the first chapter of our thesis, we explored a specific by-product, namely spent coffee waste (SCW), which comprises products deemed unconsumable by humans for ruminants' diets. SCW, rich in organic compounds, is examined for its potential as ruminant feed, with attention given to dosage, processing methods, and effects on animal health.

The current first chapter is conducted *in vitro* batch culture using two different experimental designs with three TRIALS. The first experimental design, TRIAL. 1, was performed with a control diet of 500 mg (fresh matter basal diet, 60% hay/40% concentrate), and coffee waste was used as a feed additive at 1%, 10%, and 20% of the substrate. The second experimental design, TRIAL. 2, and TRIAL. 3, were performed with the control group and coffee waste replaced part of the grass (TRIAL. 2) or the concentrate mixture (TRIAL. 3). The four different dosages (30:70, 50:50, 70:30, and 100) inclusion levels of SCW in the basal diet (raw or ensilage) were achieved by replacing the grass or concentrate.

During our *in vitro* batch culture study, TRIAL 1, SCW was supplemented as a feed additive, resulting in an increase in the production of volatile fatty acids by up to 2.7% and gas production by 10.86%, but it did not show a suppressive effect on CH₄ production. In contrast, in TRIAL 2 and 3, when SCW was included as a replacement for grass or concentrate for substrate, there was a significant reduction in CH₄ production with increasing levels of SCW inclusion. This reduction in CH₄ production was accompanied by negative effects on nutrient digestibility and total volatile fatty acid production. These

findings demonstrate that SCW could be used as a potential prebiotic feed additive. However, when SCW is used as a replacement at 70:30 and 50:50 for silage, these dosages appear to be feasible for use in the field as a substitute for animal feed (grass and concentrate).

Amidst the exploration of new strategies to mitigate CH₄ emissions, feed additives, particularly organic acids, emerge as a potential avenue. Previous studies have investigated various organic acids as CH₄ suppressors for ruminants; however, their results exhibit variability and inconsistency. Furthermore, the majority of the data stems from *in vitro* studies, with limited information from *in vivo* experiments. Given these challenges, there is a pressing need to identify a new supplement that can effectively suppress enteric CH₄.

In our second chapter, we explored the use of citric acid as a feed additive for sheep. Recent research has expressed interest in citrus-based products as CH₄ suppressors or protein sources for animal diets (Yamada et al., 2023). Notably, previous studies have utilized citric acid by-products and citric acid waste-fermented yeast waste as alternative energy sources, demonstrating no adverse effects on chicken farming or tropical lactating cows.

In this *in vivo* study, we measured nutrient intake, total tract digestibility, ruminal fermentation (at 0, 3, 6, 9, and 12 hours after feeding), and CH₄ emissions in Corriedale sheep fed a diet supplemented with citric acid. The experiment followed a replicated 2×2 crossover design with 15-day periods and involved 4 Corriedale wether sheep (initial BW of 77.5±2.75 kg). Treatments included a control group (no additive) and the addition of citric acid (40 mM). The basal diet comprised Kleingrass hay and a commercial concentrate mixture at a ratio of 2:1. The animals in the four respiration chambers were

equipped to measure CH₄ emissions for two days each period, with two 24 h measurement cycles. In the present study, the addition of citric acid reduced feed intake and digestibility, though it did not exhibit suppressing effects on CH₄ production.

Our third chapter investigated the effects of itaconic acid as a feed additive at a dosage of 20 mM. Given its potential anti-inflammatory and anti-pathogenic activities, itaconic acid was hypothesized to counteract methanogens in the rumen, presenting a possible avenue for mitigating CH₄ production from ruminants. However, information on the use of itaconic acid as a feed additive in ruminant diets is exceedingly limited. Only one study has reported its potential to reduce CH_4 in *in vitro* conditions. Therefore, the objectives of our current study mirror those of the second chapter, measuring the same parameters using the same methods.

Despite its potential, itaconic acid did not yield significant results in CH₄ production for all analyzed parameters, except for CH₄ emissions (L/kg DMI). Our study observed a significant increase in CH₄ emissions (L/kg DMI) in sheep whose diet contained itaconic acid. All rumen fermentation parameters, including statistically significant changes in propionate and non-statistically significant changes in acetate and butyrate, exhibited a reduction in our third chapter, leading to a significant decrease in total VFA. While itaconic acid demonstrates potential as a feed additive for the livestock sector, there is still limited information on its advantages and disadvantages. Further research is needed to clarify the full potential of this product. 本論文の第1章は、反芻動物の飼料または飼料添加物としてのヒトの食品副 産物の利用に関して *in vitro* 実験によって評価したものであり、第2章は、本研 究の主な目的である、反芻動物に対する有機酸の添加によるメタン(CH4)抑制の 効果を検討した。第1章では *in vitro*、第2章では *in vivo* 実験を行い、CH4 排出 の抑制、家畜の生産性向上、同時に反芻家畜の飼料に関する経済的懸念への対 応に重点を置いて検討した。

反芻家畜、特に牛は、世界規模で肉と牛乳を供給している。とはいえ、世界 人口の増加と肉と牛乳に対する需要の増大は、畜産部門、特に環境の持続可能 性の点において課題が突きつけられている。反芻家畜は栄養要求を満たす一方 で、温室効果ガス排出に大きく寄与しており、特に CH4 が主要な構成要素とな っている。

大気中では温室効果ガス、特にCH4のレベルが上昇していることから、気候変 動に対する懸念が高まり、畜産業においてその解決策を見出す努力が続けられ ている。現在、研究者たちは、CH4の排出とエネルギーロスにつながる反芻動 物の消化効率の悪さに着目している。研究者たちは、動物福祉や酪農家の利益 を損なうことなく、これらの排出を軽減する戦略を積極的に模索している。こ こ数十年、CH4緩和技術に注目が集まっており、飼養管理から有望な知見が得 られている。しかし、これらの戦略の大半は経済的デメリット、動物福祉規制 との抵触、畜産物の生産性やルーメン発酵への悪影響がある。このため、CH4

を緩和し、酪農家と家畜にとって有益な新素材を見つける必要がある。飼料添加物や副産物を含む食餌への介入が、解決策として検討されている。

副産物、特に人間の食物に由来する副産物は、家畜の飼料に配合できる可能 性があると考えられている。本論文の第1章では、特定の副産物、すなわち使 用済みコーヒー廃棄物 (SCW) について検討した。有機化合物を豊富に含む SCW について、反芻動物の飼料としての可能性を、投与量、加工方法、動物の 健康への影響に注意を払いながら検討した。

第1章では、3つのTRIALによる2つの異なる実験計画を用いて、試験管内バ ッチ培養を行った。最初の実験デザインであるTRIAL1では、対照飼料を 500mg(乾草60%/濃厚飼料40%)とし、飼料添加物としてSCWを基質の1%、 10%、20%として用いた。第二の実験計画であるTRIAL2、およびTRIAL3は 対照群とし、SCWを牧草の一部(TRIAL2)または濃厚飼料の一部(TRIAL3) に置き換えた。基礎飼料(生またはサイレージ)中のSCWの含有量は、牧草ま たは濃厚飼料を置き換えることで4種類(30:70、50:50、70:30、100)とした。 試験管内バッチ培養試験であるTRIAL1では、SCWを飼料添加物として投与 した結果、揮発性脂肪酸の生産量が最大で2.7%増加し、ガス生産量が10.86%増 加したが、CH4生産量の抑制効果は認められなかった。対照的に、TRIAL2と3 では、SCWを牧草や濃厚飼料に置き替わる基質として配合した場合、SCW 配 合量の増加に伴いCH4生産量が有意に減少した。このCH4産生量の減少は、栄 養消化率および総揮発性脂肪酸産生量への負の影響を伴っていた。これらの結 果は、SCWがプレバイオティック飼料添加物として利用できる可能性を示して

いる。しかし、SCW をサイレージの代替物として 70:30 および 50:50 で使用 する場合、これらの用量は動物飼料(牧草および濃厚飼料)の代替品として牧 場で使用することが可能であると思われる。

CH4 排出を抑制する新たな戦略が模索される中、飼料添加物、特に有機酸が手 段として浮上してきた。これまでの研究では、反芻家畜の CH4 抑制剤として 様々な有機酸が研究されてきたが、その結果にはばらつきがあり、一貫性がな い。さらに、データの大半は *in vitro* 試験によるもので、*in vivo* 実験による情報 は限られている。これらの課題を考えると、消化管内 CH4 を効果的に抑制でき る新しいサプリメントを特定することが急務である。

第2章では、羊の飼料添加物としてのクエン酸の利用について検討した。最近 の研究では、柑橘類をベースとした製品が CH4 抑制剤や動物用飼料のタンパク 質源として注目されている(Yamada 6、2023)。注目すべきは、これまでの研 究で、クエン酸副産物やクエン酸廃棄物-発酵酵母廃棄物を代替エネルギー源と して利用し、養鶏や熱帯泌乳牛に悪影響がないことを実証していることである。 本 *in vivo* 試験では、クエン酸を添加した飼料を与えたコリデール種羊の栄養摂 取量、消化率、ルーメン発酵(給餌後 0、3、6、9、12 時間)、CH4 排出量を測 定した。実験は1期15日間の2×2クロスオーバーデザインに従い、4 頭のコリ デール種ヒツジ(初期体重 77.5 ± 2.75kg)を用いた。対照群(無添加)とクエ ン酸(40 mM)添加群に分けた。基礎飼料は、クレイングラス乾草と市販の濃 厚飼料を2:1の割合で混合したものとした。4 つの呼吸試験チャンバー内の動 物は、各期間2日間、24時間の測定サイクルで2回、CH4排出量が測定された。

本研究では、クエン酸の添加は飼料摂取量と消化率を低下させたが、CH4産生 を抑制する効果は認められなかった。

第3章では、飼料添加物としてのイタコン酸の効果を20mMの用量で調査した。抗炎症作用および抗病原性作用が期待されることから、イタコン酸はルーメン内のメタン生成菌を抑制し、反芻動物からのCH4産生を低下させる可能性があると考えられた。しかし、反芻胃動物の飼料添加物としてのイタコン酸の利用に関する情報は極めて限られている。試験管内条件下でCH4削減の可能性を報告した研究は1件のみである。したがって、今回の研究の目的は第2章と同じであり、同じ方法で同じパラメーターを測定した。

上記の可能性にもかかわらず、イタコン酸は CH4 排出量(L/kg DMI)を除く すべての分析パラメータにおいて、CH4 産生に有意な結果をもたらさなかった。 我々の研究では、飼料にイタコン酸を配合したヒツジにおいて、CH4 排出量

(L/kg DMI)の有意な増加が観察された。第3章では、プロピオン酸の統計的 に有意な変化を含む全てのルーメン発酵パラメータが減少し、総揮発性脂肪酸 が有意に減少した。イタコン酸は畜産部門の飼料添加物としての可能性を示し ているが、その利点と欠点に関する情報はまだ限られている。この物質の可能 性を完全に解明するためには、さらなる研究が必要である。

ACKNOWLEDGEMENT

General Conclusion

The findings from this comprehensive study highlight the potential of alternative feed ingredients in addressing the challenges of ruminant agriculture, particularly in reducing CH₄ emissions. Spent coffee waste (SCW) emerges as a promising alternative due to its nutrient composition, showing potential to mitigate the environmental impact of ruminant farming. However, caution is warranted regarding higher dosages, as they may adversely affect animal production. Further research is essential to assess the implications, optimal dosages, and administration methods of SCW in ruminant diets for a more informed and sustainable approach.

Examining citric acid and itaconic acid as feed additives revealed insights into their effects on feed intake, digestibility, and CH₄ emissions. Citric acid, despite reducing feed intake and digestibility due to the higher dosage, did not impact rumen fermentation. However, the translation of *in vitro* CH₄ suppression to *in vivo* scenarios remains complex and requires additional research to determine optimal levels and usage methods in commercial ruminant feed.

Itaconic acid at 20 *mM* in sheep diets did not significantly impact CH₄ emissions, emphasizing the variability in the effectiveness of organic acids. The search for reliable CH₄ suppressors continues, underscoring the importance of nuanced understanding and exploration of novel additives for consistent mitigation in ruminant diets. Overall, these studies contribute valuable insights to the ongoing efforts to develop sustainable feeding strategies that optimize animal health, productivity, and environmental sustainability in the field of ruminant nutrition.

ACKNOWLEDGEMENT

Acknowledgement

The work presented in this thesis was conducted at Obihiro University of Agriculture and Veterinary Medicine. Professor Takehiro Nishida provided supervision, with additional assistance from co-supervisors Professors Masaaki Hanada and Naoki Fukuma.

I want to begin by expressing profound gratitude to my main supervisor, Prof. Takehiro Nishida. From the inception of my study until now, he has not only served as my supervisor but has also been an exemplary mentor, providing invaluable guidance and encouragement. I take pride in being his student, and his unwavering support has been instrumental in my academic journey.

I also extend my sincere appreciation to my co-supervisors, Prof. Masaaki Hanada and Prof. Naoki Fukuma, for their continued guidance and support. Dr. Eslam Khalifa deserves special thanks for his collaboration, encouragement, and invaluable guidance throughout my research.

I would like to express my gratitude to the Moonshot project and Sanyu Group Co., Ltd., Japan, for funding and providing me the opportunity to complete certain parts of my thesis.

Acknowledgment is due to my friends and fellow students in the Animal Husbandry section and those I met during my study period for their support. Their camaraderie has been a source of strength, and I am truly thankful to all of them. Special thanks to my friends in Obihiro and the Mongolian committee, and to B. Tugsbaatar's family, and B. Tuvshintulga's family, who motivated me to start this study.

Heartfelt thanks are extended to my beloved family, parents, and sisters, whose constant love and support have been my pillars of strength. Their unwavering encouragement keeps me motivated and confident, playing a pivotal role in my

accomplishments. I want to express special gratitude to my parents, father P. Batbekh and mother Ts. Otgontuya, for their love and guidance, which have been indispensable to my success.

Finally, I would like to express special appreciation to my mother, Ts. Otgontuya, who has a warm heart, the strongest mind, and is the most beautiful lady in the world. I am always filled with your abundant love, and without your immense motivation, I could not have completed this thesis.