# IN VITRO GROWTH INHIBITORY EFFECTS OF COMBINED APPLICATIONS OF CLOTRIMAZOLE, KETOCONAZOLE, AND CLODINAFOP-PROPARGYL ON EQUINE BABESIA PARASITES

Sabine Bork, Naoaki Yokoyama, Masashi Okamura, and Ikuo Igarashi\*

National Research Center for Protozoan Diseases (NRCPD), Obihiro University of Agriculture and Veterinary Medicine, Inada-cho, Obihiro, Hokkaido 080-8555, Japan.

\*Corresponding author:NRCPD, Obihiro University of Agriculture and Veterinary Medicine, Inada-cho, Obihiro, Hokkaido 080-8555, Japan.Tel.: +81-155-49-5641; Fax: +81-155-49-5643

#### Abstract

Recently, we reported the *in vitro* growth inhibitory effects of the imidazole derivatives, clotrimazole (CLT) and ketoconazole (KC), and the herbicide, clodinafop-propargyl (CP), against bovine and equine *Babesia* parasites. Additionally, in the bovine species, several combined applications of the CLT, KC, and CP were reported to significantly increase the inhibitory effects. In this study, similar combination tests were carried out in order to confirm possible synergistic effects in the equine species, *Babesia caballi* and *Babesia equi*. Combinations of CLT/KC, CLT/CP, and CLT/KC/CP exerted a significantly enhanced growth inhibitory efficacy in *B. caballi*, in contrast to *B. equi*, where no synergistic effects were observed in any of the combinations. Our results suggest that the combined usage of these drugs may have a high enough potential to be considered applicable in practice.

Keywords: in vitro, Babesia, clotrimazole, ketoconazole, clodinafop-propargyl

#### Introduction

Equine babesiosis, also known as biliary fever, is a tick-borne disease of equidae caused by two Apicomplexan parasites, *Babesia caballi* and *Babesia equi*. These intraerythrocytic protozoa occur in many regions of the world (Friedhoff et al., 1990) and are considered to be an important threat for livestock (Schein, 1988). Infected animals suffer from fever, anorexia, severe anemia, and apathy (Holman et al., 1998), leading to their restricted use in sports and in the meat/horse industry. The mortality rate is as high as 50% (Levine, 1985), and the surviving horses are likely to remain subclinical carriers for life. Currently, horses awaiting importation into countries considered free of the equine babesiosis (such as Japan) have to adhere to strict regulations, including serological tests (Holman et al., 1998).

Although many anti-babesial drugs have been developed and applied more or less successfully for several years (Kuttler et al., 1987; Singh et al., 1980; Bruning, 1996), many compounds have been shown to have severe side effects or to produce drug-resistant parasites (Adams, 1980; Granwell, 1990; Upcroft, 1994; Wijaya et al., 2000). With regard to these facts, further development of potent and safe drugs is highly required. We recently demonstrated the efficacy of the imidazole derivatives and anti-fungal agents, clotrimazole (CLT) and ketoconazole (KC), and the herbicide, clodinafop-propargyl (CP), when applied singly (as mono-drug) to equine Babesia parasites (Bork et al., 2003a) and singly as well as in combinations to bovine Babesia species (Bork et al., 2003b) by using the in vitro cultivation system (Igarashi et al., 1998; Nagai et al., 2003; Bork et al., 2003c, 2004). Since several studies reported the usefulness of combined drug applications with the benefit of higher sensitivity and fewer adverse effects (Krause et al., 2000; Wijaya et al., 2000), we were encouraged to evaluate the efficacy of drug combinations against the in vitro growth of B.

equi and B. caballi.

# Materials and methods

Parasites and in vitro cultivation: United States Department of Agriculture (USDA) strains of B. caballi and B. equi, maintained in the NRCPD for several years, were grown in equine erythrocytes as described previously (Bork et al., 2003c).

Chemicals: Clotrimazole (CLT) (1-[(2-chlorophenyl) diphenylmethyl]-1*H*-imidazole) and ketoconazole (KC) (cis-1-acetyl-4-[4-[2- (2,4-dichlorophenyl) -2- (1*H*-imidazole-1-ylmethyl)-1,3-dioxolan-4-yl] methoxy] phenyl] piperazine) were obtained from Sigma Chemical Co. (St. Louis, Missouri, U S A). Clodinafop-propargyl (CP) ((2*R*)-2-[4-[(5-Chloro-3-fluoro-2-pyridinyl) oxy]phenoxy] propanoic acid-2-propynyl ester) was purchased from Riedel-de Haën Laborchemikalien (Seelze, Germany). The stock solutions were prepared as described previously (Bork *et al.*, 2003*a*).

Drug combination tests: The concentrations of CLT, KC, and CP, acting suppressive but not destructive to the parasites, were determined according to previous experiments (Bork et al., 2003b) and applied in the present study. The classification into "synergistic," "antagonistic," or "no effect" was made as follows: The inh.% value of the drug combination was compared to that of the drug applied singly. When the inh.% value of the combination was higher than that of the most effective singly applied drug, the result was considered to be "synergistic." In the case that the inh.% value of the drug combination was lower, it was determined to be "antagonistic." Finally, when both inh.% values (drug combination and singly applied) were almost identical, it was determined to have "no effect."

## Results

Table 1 shows the percentage of growth inhibition (inh.%) of the combined applications of CLT, KC, and CP against equine *Babesia* parasites in comparison to the control inh.% of the mono-drug applications. The inh.% varied from 1.6% in *B. caballi* exposed to KC/CP to 96.2% in the same parasite exposed to the triple combination of CLT/KC/CP. In addition, in *B. caballi*, the CLT/KC, CLT/CP, and CLT/KC/CP were found to act synergistically and reveal a stronger enhancing inh.% compared to that of the single application. Only the combination of KC/CP had a significantly attenuated effect. On the other hand, in *B. equi*, the inh.% ranged from 29.4 (KC/CP) to 92% (CLT/KC). Combinations of KC/CP and CLT/KC/CP acted antagonistically; especially, the inh.% of the KC/CP combination revealed a 0.4-fold lower sensitivity in *B. equi* than that of the KC single application.

# Discussion

In this study, the applicability of drug combinations containing CLT, KC, and CP was evaluated against the *in vitro* growth of *B. equi* and *B. caballi*. In *B. caballi*, all combinations containing CLT showed strong enhancing efficacies. These findings correlate with previous reports, according to which these combinations significantly enhanced the destructive effects in *B. bovis* and *B. bigemina* (Bork *et al.*, 2003b), but were clearly different from the results of *B. equi*. All but two combinations (KC/CP and CLT/KC/CP) acted antagonistically for *B. equi*, whereas other combinations did not show any enhancing or weakening effects. This interesting observation is consistent with previous reports describing several differences between the two equine species in the level of drug sensitivity (Schein, 1988). It is well understood that the sensitivity of

B. equi to drugs is usually lower than that of B. caballi (Bork et al., 2003c; Nagai et al., 2003), which might be explained through the probably distinct generic classification of B. equi as Theileria equi (Mehlhorn et al., 1998). In contrast, the occurrence of strong antagonistic effects of the KC/CP combination in both equine parasites is also consistent with the results obtained in B. bovis, but in complete contrast to the findings in B. bigemina (Bork et al., 2003b). These findings possibly suggest a common metabolic pathway among the Babesia parasites except B. bigemina.

Synergy between drugs was proved to be a valuable tool to improve treatment efficacy in several diseases (Radloff et al., 1996). In a previous report, CLT exerted synergistic effects when combined with the antibiotic mefloquine, but it was found to be antagonistic with the widely used antibiotic chloroquine against Plasmodium parasites (Tiffert et al., 2000). Combinations of KC with either a lactone derivative (lovastatin, imidazole acetamide, or benznidazole) or an ergosterol biosynthesis inhibitor (terbinafine) were found to be highly successful against Trypanosoma cruzi infection (Araujo et al., 2000; Urbina et al., 1993). In P. falciparum, the CLT may interfere with the hemobiosynthesis (Huy et al., 2002), and, in fungi, the KC is considered to act on the microsomal P450-dependent enzyme system (Matthew et al., 1993), while the CP inhibits one key enzyme in the de novo fatty acid biosynthesis of plants (Zuther et al., 1999). Although Plasmodium and Babesia species are known to have close biological similarities (Clark and Jacobson, 1998), the action-mode of the three compounds may differ among Babesia parasites and, therefore, requires further clarification.

In conclusion, the usefulness of combined applications of CLT, KC, and CP was clearly demonstrated against the *in vitro* growth of *B. equi* and *B. caballi*.

Table 1. Growth inhibitory efficacy (inh.%) at suppressive concentrations of CLT, KC, and CP to the in vitro growth of B. caballi and B. equi

Drugs	Babesia caballi			Babesia equi		
	drug conc.	inh.% <sup>*</sup>	effects <sup>\$</sup>	drug conc.	inh.%*	effects <sup>§</sup>
CLT	15µM	50.5%		2.5μΜ	71.4%	
KC	10μΜ	18.2%		$10 \mu M$	93.6%	
CP	250μΜ	14.5%		500μΜ	57.0%	
CLT/KC	15μΜ/10μΜ	85.4%	syn	2.5μΜ/10μΜ	92.0%	ne
CLT/CP	15μΜ/250μΜ	92.0%	syn	2.5μΜ/500μΜ	74.0%	ne
KC/CP	10μΜ/250μΜ	1.6%	ant	10μΜ/500μΜ	29.4%	ant
CLT/KC/CP	15μΜ/10μΜ/250μΜ	96.2%	syn	2.5μΜ/10μΜ/500μΜ	70.7%	ant

<sup>\*</sup> inh.% indicates parasitemia/control parasitemia without drug addition.

## Acknowledgments

This study was supported by a Grant-in-Aid for Scientific Research (A) from the Japan Society for the Promotion of Science and by grants from Special Coordination Funds for Science and Technology from the Science and Technology Agency and from The 21st Century COE Program (A-1), the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

## References

<sup>\$</sup> synergistic (syn); antagonistic (ant); no effect (ne)

- Adams, L.G. 1980. Clinicopathological aspects of imidocarb diproprionate toxicity in horses. *Res. Vet. Sci.* 31: 54-61.
- Araujo, M.S.S., Martins-Filho, O.A., Pereira, M.E.S., & Brener, Z. 2000. A combination of benznidazole and ketoconazole enhances efficacy of chemotherapy of experimental Chagas' disease. *J. Antimicrob. Chemother.* 45: 819-824.
- Bork, S., Yokoyama, N., Matsuo, T., Claveria, F.G., Fujisaki, K., & Igarashi, I. 2003a. Clotrimazole, ketoconazole, and clodinafop-propargyl as potent growth inhibitors of equine *Babesia* parasites during *in vitro* culture. *J. Parasitol.* 89: 604-606.
- Bork, S., Yokoyama, N., Matsuo, T., Claveria, F.G., Fujisaki, K., & Igarashi, I. 2003b. Clotrimazole, ketoconazole, and clodinafop-propargyl inhibit the *in vitro* growth of *Babesia bigemina* and *Babesia bovis* (Phylum Apicomplexa). *Parasitology* 127: 1-5.
- Bork, S., Yokoyama, N., Matsuo, T., Claveria, F.G., Fujisaki, K., & Igarashi, I. 2003c. Growth inhibitory effects of triclosan on equine and bovine parasites. *Am. J. Trop. Med. Hyg.* 68: 334-340.
- Bork, S., Yokoyama, N., Ikehara, Y., Kumar, S., Sugimoto, C., & Igarashi, I. 2004. Growth-inhibitory effect of heparin on *Babesia* parasites. *Antimicrob. Agents Chemother.* 48: 236-241.
- Bruning, A. 1996. Equine piroplasmosis: An update on diagnosis, treatment, and prevention. *Br. Vet. J.* 152: 139-151.
- Clark, I.A., & Jacobson, L.S. 1998. Do babesiosis and malaria share a common disease process? *Ann. Trop. Med. Parasitol.* 92: 483-488.
- Friedhoff, K.T., Tenter, A.M., & Muller, I. 1990. Hemoparasites of equines: Impact on international trade of horses. *Rev. Sci. Tech.* 9: 1187-1194.
- Granwell, M.P. 1990. Efficacy of long-acting oxytetracycline for the prevention of tick-borne fever in calves. *Vet. Rec.* 126: 334-336.
- Holman, P.J., Becu, T., Bakos, E., Polledo, G., Cruz, D., & Wagner, G.G. 1998. *Babesia equi* field isolates cultured from horse blood using a microcentrifuge method. *J. Parasitol.* 84: 696-699.
- Huy, N.T., Kamei, K., Yamamoto, T., Kondo, Y., Kanaori, K., Takano, R., Tajima, K., & Hara, S. 2002. Clotrimazole binds to heme and enhances heme-dependent hemolysis: Proposed antimalarial mechanism of clotrimazole. *J. Biol. Chem.* 277: 4152-4158.
- Igarashi, I., Njonge, F.K., Kaneko, Y., & Nakamura, Y. 1998. Babesia bigemina: In vitro and in vivo effects of curdlan sulfate on growth of parasites. Exp. Parasitol. 90: 290-293.
- Krause, P.J., Lepore, T., Sikand, V.K., Gadbaw, J. Jr., Burke, G., Telford, S.R. 3rd, Brassard, P., Pearl, D., Azlanzadeh, J., Christianson, D., McGrath, D., & Spielman, A. 2000. Atovaquone and azithromycin for the treatment of babesiosis. *N. Engl. J. Med.* 343: 1454-1458.
- Kuttler, K.L., Zaugg, J.L., & Gispon, C.A. 1987. Imidocarb and parvaquone in the treatment of piroplamosis (*Babesia equi*) in equids. *Am. J. Vet. Res.* 48: 1613-1616.
- Levine, N.D. 1985. Veterinary protozoology. p. 414. Iowa State University Press, Ames, Iowa, U.S.A.
- Matthew, D., Brennan, B., Zomorodi, K., & Houston, J.B. 1993. Disposition of azole antifungal agents. I. Nonlinearities in ketoconazole clearance and binding in rat liver. *Pharm. Res.* 10: 418-422.
- Mehlhorn, H., & Schein, E. 1998. Redescription of *Babesia equi* Laveran 1901 as *Theileria equi*. *Parasitol Res.* 84: 467-475.
- Nagai, A., Yokoyama, N., Matsuo, T., Bork, S., Hirata, H., Xuan, X., Zhu, Y., Claveria, F.G., Fujisaki, K., & Igarashi, I. 2003. Growth inhibitory effects of artesunate, pyrimethamine, and pamaquine against *Babesia equi* and *Babesia caballi* in *in vitro* cultures. *Antimicrob. Agents Chemother.* 47: 800-803.

- Radloff, P.D., Philipps, J., Nkeyi, M., Hutchison, D., & Kremser, P.G. 1996. Atovaquone and proguanil for *Plasmodium falciparum* malaria. *Lancet* 347: 1511-1514.
- Schein, E. 1988. Equine babesiosis. pp. 197-208. *In:* Babesiosis of domestic animals and man. Ristic, M. (Ed.), CRC Press, Boca Raton, Florida.
- Singh, B., Banerjee, D.P., & Gautam, O.P. 1980. Comparative efficacy of diminazene diaceturate and imodocarb diproprionate against *Babesia equi* infection in donkeys. *Vet. Res.* 7: 173-179.
- Tiffert, T., Ginsburg, H., Krugliak, M., Elford, B.C., & Lew, V.L. 2000. Potent antimalarial activity of clotrimazole in *in vitro* cultures of *Plasmodium falciparum*. *Proc. Natl. Acad. Sci. U.S.A.* 97: 331-336.
- Upcroft, P. 1994. Multiple drug resistance in the pathogenic protozoa. Acta Trop. 56: 195-212.
- Urbina, J.A., Lazardi, K., Marchan, E., Visbal, G., Aguirre, T., Piras, M.M., Piras, R., Maldonado, R.A., Payares, G., & de Souza, W. 1993. Mevinolin (lovastatin) potentiates the antiproliferative effects of ketoconzole and terbinafine against *Trypanosoma (Schizotrypanum) cruzi: In vitro* and *in vivo* studies. *Antimicrob. Agents Chemother.* 37: 580-591.
- Wijaya, A., Wulansari, R., Ano, H., Inokuma, H., & Makimura, S. 2000. Therapeutic effect of clindamycin and tetracycline on *Babesia rodhaini* infection in a mouse model. *J. Vet. Med. Sci.* 62: 835-839.
- Zuther, E., Johnson, J.J., Haselkorn, R., McLeod, R., & Gornicki, P. 1999. Growth of *Toxoplasma gondii* is inhibited by aryloxyphenoxyproprionate herbicides targeting acetyl-Co A carboxylase. *Proc. Natl. Acad. Sci. U.S.A.* 96: 13387-13392.